

Doctoral Thesis (Abridged)

博士論文 (要約)

Endocrine network essential for energy homeostasis  
in the two-spotted cricket, *Gryllus bimaculatus*

(フタホシコオロギにおけるエネルギー恒常性に重要な内分泌系ネットワーク)

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Graduate School of Frontier Sciences  
The University of Tokyo

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## Abbreviations

7-TM: Seven-transmembrane

AcG: Accessory gland

ACP: AKH/Corazonin-related peptide

AG: Abdominal ganglia

AKH: Adipokinetic hormone

APC: AKH-producing cell

Ast-C: Allatostatin-C

BLAST: The Basic local alignment search tool

Br: Brain

Burs- $\alpha$ : Bursicon- $\alpha$

CA: Corpora allata

cAMP: Cyclic Adenosine monophosphate

CC: Corpora cardiaca

CCAP: Crustacean cardioactive peptide

CCAPR: CCAP receptor

CHCA:  $\alpha$ -Cyano-4-hydroxycinnamic acid

CNS: Central nervous system

CoG: Cluster of orthologous groups of proteins

CRZ or Crz: Corazonin

DAG: Diacyl glycerol

DCM: Dichloromethane

DH: Diuretic hormone

DNA: Deoxyribonucleic acid

dsRNA: Double-stranded RNA

EDT: 3,4-Ethoxylene dioxythiophene

EH: Eclosion hormone

FB: Fat body

FG: Foregut

G-6-DH: Glucose-6-phosphate dehydrogenase

Glut1: Glucose transporter

GO: Gene ontology

Gp: Glycogen phosphatase

GPCR: G-protein coupled receptor

HG: Hindgut

HL: Hemolymph

HPLC: High performance liquid chromatography

IPC: Insulin-producing cell

IRS: Insulin receptor substrate

KEGG: Kyoto Encyclopedia of genes genomes

MALDI-TOF: Matrix-assisted laser desorption ionization-time of flight

MG: Midgut

MIP: Myoinhibitory peptide

MS: Mass spectrometry

MT: Malpighian tubules

Mu: Muscle

n.s.: Not significant

NCBI: National center for biotechnology information

NCC: Nervi corporis cardiaci

NGS: Next-generation sequencer

Op: Optic lobe

PBS: Phosphate-buffered saline

PCR: Polymerase chain reaction

PDF: Pigment dispersing factor

PI: Pars intercerebralis

Real time qRT-PCR: Real time quantitative RT-PCR

RNA: Ribonucleic acid

RNAi: RNA interference

RT-PCR: Reverse transcription-PCR

SG: Subesophageal ganglion

sNPF: Short neuropeptide F

TAG: Triacyl glycerol

Te: Testis

TFA: Trifluoroacetic acid

TG: Thoracic ganglia

TMSBr: Trimethylsilyl bromide

TPP: Trehalose phosphate phosphatase

TPS: Trehalose phosphate synthetase

Tre: Trehalase

Tret1: Trehalose transporter

VNS: Ventral nervous system



## **Introduction**

### **Energy homeostasis and its regulation by endocrine factors**

Insects, as heterotrophic organisms, have to obtain energy from outside sources. The dietary energy is allocated for all living processes *in vivo* (Fig. 0-1 A). The concept of energy homeostasis was elaborated by Walter Cannon (Cannon, 1929) and Claude Bernard (Holmes, 1986) with an essential role for survival, which promises a coordinated regulation on food intake and energy expenditure to maintain a fairly stable condition. In related researches, energy homeostasis has been frequently illuminated as the mechanisms by which all living creatures satisfy their energy demands, but also manipulates food intake when expenditure changes (Chapelot and Charlot, 2019). However, a linear relationship is not adequate for describing energy expenditure and food intake. In mammals, overfeeding leads to a 7 to 18% increase in total energy expenditure, allowing ~25% of excess energy to be dissipated. Excess amount of food intake causes a 7 to 18% increase in total energy expenditure, allowing approximately 25% of over-loaded energy to be consumed (Klein and Goran, 1993), indicating the dynamic correlation between energy intake and expenditure for maintaining energy homeostasis.

To maintain energy homeostasis, animals rely on various endogenous signals, such as circulating metabolites, neuropeptides and neuromodulators (Fig. 0-1 B). Flexible physiology and behavior enable animals to adapt against extendedly varied challenges. The nervous system and endocrine control organize effectively in these adaptations and guarantee coordination between the external and the internal environments, in addition to the regulations of appropriate balanced nutrients and various living processes. Endocrine factors have been demonstrated as regulators in energy homeostasis in various insects. Different from synaptic transmission, endocrine factors deliver their signals slowly to their target tissues or cells, because neuropeptides and peptide hormones are synthesized and released into hemolymph. Second messages mediate intracellular signals after the combination of endocrine factors (ligands) and their corresponding receptors, which are G-protein coupled receptors (GPCRs) in most cases. The activated intracellular signals with specific spatial distributions promise the specificity of neuropeptide signals. The variety of endocrine factors and the activated downstream signals enable them to involve the regulations in almost all the living processes, especially the energy homeostasis that is composed of complex collaboration between different organs and different living processes. Hence, multiple neuropeptides are recognized to contribute to an extensive range of temporal and spatial harmonization (Kim et al., 2017).

To date, energy homeostasis has been imagined as a fixed programmed value, but only studies about adaptive responses to external stimulus are understood. In a review of energy homeostasis, Chapelot and Charlot elaborate that energy homeostasis can be considered as a stochastic value with some possibility to be predicted with an expected accuracy if enough determinants are known (Chapelot and Charlot, 2019). However, accumulation of these determinants and mathematically calculating the index will be daunting. Indeed, multiple metabolites, hormones, neurotransmitters and neurons, and new molecules with regulatory roles in energy homeostasis have been constantly discovered in terms of their functions in food intake and energy expenditure (Gavrieli and Mantzoros, 2016).

## **Communications or cross-talk between neuropeptides**

Gastrointestinal tract collects dietary information and CNS activates adaptive feeding behavior and metabolic flux. The coordinated regulation of the gastrointestinal tract and brain is elaborated as brain-gut axis, which are composed of bidirectional passes from both the CNS and the intestine. The gastrointestinal tracts of animals have enteroendocrine cells (EECs), which perceive the internal intestinal information and produce neuropeptide and peptide hormones to secrete for control of various physiological events including intestinal contraction, nutrient homeostasis and appetite (Miguel-Aliaga et al., 2018).

In mammals, Glucagon-like peptide-1, pancreatic polypeptide, peptide tyrosine-tyrosine and ghrelin are experimentally proved to be intestinal peptides that contribute to energy metabolism (Maric et al., 2014, Murphy et al., 2006). Moreover, ghrelin secreted from intestine suppresses insulin secretion. By contrast, cholecystokinin can enhance the secretion of insulin and other hormones (Lo et al., 2011); (Rehfeld et al., 1980). Hence, the nutritional states are regulated by intestinal peptides relaying the modulation of the secretion of insulin in the beta-cells and other neuropeptides in CNS. Such endocrinal situations are similar to those in invertebrates. To date, in the fruit fly, *Drosophila melanogaster*, the midgut enteroendocrine cells express 11 neuropeptides, including Allatostatin-A, Allatostatin-B, Allatostatin-C, Bursicon- $\alpha$ , CCHamide-1 and -2, Diuretic hormone 31, Neuropeptide F, Orcokinin-B, Short neuropeptide F and Tachykinin, although their distributions are varied in larvae and adult (Nassel and Zandawala, 2019). Besides, *D. melanogaster* intestine expresses 23 neuropeptide receptors, whose ligands are all express in the brain. These neuropeptide signals in intestine transfer the ‘information’ of the fluctuation of the digested compounds or metabolites, and sequentially activate downstream

signals for adaptive transitions. For example, DH31-producing cells in intestine can sense the dietary proteins and amino acids (Park et al., 2016). CCHamide-2 from intestine is thought to stimulate brain insulin-producing cells to regulate food intake (Ren et al., 2015). Not only the brain-gut axis, communication cross organs occurs between brain and fat body. For example, Bursicon- $\alpha$ -producing neurons are activated by dietary sucrose, and Bursicon- $\alpha$  can activate Lgr2 receptors in the brain neurons, which downregulate Adipokinetic hormone (AKH) signaling to involve in regulation of energy homeostasis (Scopelliti et al., 2019). Indeed, unidirectional signal activities are discovered in other organs. For example, Allatotropin (AT) stimulates juvenile hormone biosynthesis in the corpora allata in *Bombyx mori* (Yamanaka et al., 2008). Not only organ-cross communications, an antagonistic effect of sNPF in modulating feeding motivation was proved within intestine of *B. mori* (Matsumoto et al., 2019).

The study on neuropeptides communication reveals the high possibility that the neuropeptides have been practically collaborated each other or are coordinately regulated.

### **Endocrine network for understanding neuropeptides in energy homeostasis**

The concept of networked regulation has been discussed for a long time. In mammals, networks involved in the regulation of appetite, thermogenesis and energy homeostasis are concluded in several specific regions in the brain which generate cognitive and autonomic systems of the brain. These networks are modulated by peripheral factors including hormones from adipose and intestinal tissues, addressing the brain via the circulation and neuronal signals to transmit the information about energy balance and nutritional state to CNS. Actually, the endocrine network which is constructed by the neuropeptides and peptide hormones related to food intake has been reported recently with abundant connections between nodes (Fig. 0-2). However, as the network is generated from conclusions, which are collected from different studies, the validity in individual levels cannot be promised.

For endocrinal regulation of energy homeostasis, it is almost a common sense that a single neuropeptide is unlikely to be the only player capable of making decisions in entire metabolic states. Instead, endocrine network can mediate the cues from multiple factors to make a 'metabolic' decision (Banerjee et al., 2017). The orchestration of multiple neuropeptides promises energy homeostasis. For example, sNPF and TK receptors expressed in olfactory sensory neurons modifies the sensitivity to specific odorant inputs in the hungry fly, eventually increasing food odor firing recurrence and activating food-searching behavior (Root et al., 2011); (Ko et al., 2015). Meanwhile, NPF enhances the sensitivity of the receptor against sweet compounds, by contrast,

sNPF reduces the sensitivity of the receptor against bitter compounds (Inagaki et al., 2014); (Yu et al., 2016); (Sayin et al., 2018). The systemically integrated neuropeptide information results in the adaptive feeding behavior to maintain energy homeostasis. Thus, it is necessary to understand if a specific neuropeptide serves multiple disparate functions to orchestrate systemically unified functions.

Collectively, a comprehensive understanding of neuropeptides involved in energy homeostasis is imperative for insects, which could be realized at an individual level.

### **Advantages of the study using *G. bimaculatus***

The two-spotted cricket *Gryllus bimaculatus* (Fig. 0-3), which is used as a research target for investigating mechanisms of development and regeneration from early 1990s. Developmental mode of *G. bimaculatus* is the typical of arthropods compared to that of *D. melanogaster* (Hadley and Mito, 2017). In contrast to mammals, one predominantly different structure in insects is the circulatory system, indicating that insects have the distinctive nature of delivering system of oxygen and metabolites to the CNS (Maier and Orion, 1969).

All stages' animals of *G. bimaculatus* are easily maintained and prepared in the laboratory, which might be the biggest reason to be widely used for studying insect physiology and neurobiology so far; for example, Allatostatin B (MIP) was originally identified in *G. bimaculatus* as early as 1990s (Lorenz et al., 1995). Until now, *G. bimaculatus* has become suitable as an appropriate model insect for addressing the molecular mechanisms of feeding behavior and more applicable for observing behavior because of its high locomotor activity (Hadley and Mito, 2017). In addition, high efficiency of RNA interference (RNAi) in cricket is convenient for design of the RNAi-based experiments on behavioral assays through loss of function.

### **Objectives of this study**

In this study, I sought to describe a prototype of endocrine network that is essential for energy homeostasis at an individual level in *G. bimaculatus*. To address the endocrine network, following questions would be answered through this study.

- 1) Is endocrine network essential for maintaining energy homeostasis?

Binary communications, such as cross-talk between two neuropeptides, have been studied for a long time.

However, multivariant communications are poorly understood. Importance of endocrine network should be

confirmed at an individual level in insects.

- 2) What is the arrangement of endocrine network in maintaining energy homeostasis? Are there any important regulators that are extensively related to other neuropeptides?

The involvement of multiple neuropeptides determines the complexity of endocrine network in maintaining energy homeostasis. Although proof of communications between neuropeptides are plentiful, especially for Insulin/AKH signals, the frequently co-researched endocrine factors have not been considered as an extensive regulator among neuropeptides. Are there any possibilities for several neuropeptides to dominate the regulation of energy homeostasis?

- 3) How can 'information' from peripheral or circulating metabolites be delivered to CNS to maintain energy homeostasis, especially with the existence of hemolymph-brain barrier (HBB)?

The existence of brain-gut axis in insects somehow proves the possibility of some neuropeptides passing through the HBB and functioning as neurotransmitters. However, except those neuropeptides, how do most neuropeptides 'send' information to brain? Is there any tissue in insects that resembles hypothalamus in mammals?

- 4) What relationship is present between circulating metabolites and endocrine network in maintaining energy homeostasis in insects?

As stated above, energy from diets is metabolized and turns into different structures of metabolites, such as carbohydrates and lipids. The regulation of metabolites in circulation by neuropeptides needs to be elucidated in the endocrine network.

## **Challenges of this study**

As stated above, more and more neuropeptides have been proved as regulators of biological and physiological processes. In this study, other than conventional neuropeptide research and characterization of a specific neuropeptide in a specific function, I focused on the overview of communications between neuropeptides across cerebral and peripheral tissues based on the hypothesis of endocrine network. However, since endocrine network-related literatures are extremely limited, exploring feasible methods is the most challenging part in this study. Highly conserved endocrine network has been studied in the reproductive system of *D. melanogaster* (Meiselman et al., 2017). However, in the above study, the appearance of endocrine factors is limited to Ecdysis-

triggering hormone (ETH), ecdysone (20E) and Juvenile hormone (JH), whereas more endocrine factors are proved to be involved in the reproductivity (Arakane et al., 2008). The investigation of endocrine network needs to include neuropeptides as much as possible for vividly revealing the dynamic regulation of energy homeostasis. Thus, the massive work of synthesizing neuropeptides and figuring out their regulatory roles in energy homeostasis are tough. Due to the experimental limitation and massive amounts of research targets, I decided to construct an endocrine network based on key-regulators and their associated signals (Fig. 0-4). Key regulators can be determined after screening of neuropeptides. Also, its representativeness provides the possibility of study on the endocrine network through manipulation of the key regulators.

Another challenging issue is to understand the comprehensive interplay among neuropeptides from the obtained results of each neuropeptide's role in energy homeostasis to generate the endocrine network. Due to the above limitation, research on the detail mechanism of each neuropeptide involving in energy homeostasis is difficult in this study. Instead, I presented here their function and communication between neuropeptides by transcriptional regulation and conditioned peptidome analyses.

## **Components of this study**

Four chapters are enclosed in this study for revealing the prototype of endocrine network (Fig. 0-5): (1) A virtual network of neuropeptides in energy homeostatic regulation is described by bibliometric mapping. Followingly, all neuropeptides in *G. bimaculatus* were identified and evaluated their regulatory function responding to nutrient, feeding motivation, food intake and excretion. AKH and AKH/Corazonin-related peptide (ACP) were screened as key regulators. Adaptive modulation by transcriptional alteration of the selected GPCRs in fat body and corresponding ligands in brain were evaluated. (2) RNA-sequencing analysis of CC was performed for screening GPCRs in the CC and CC-synthesizing neuropeptides. Also, the whole transcriptome of CC was annotated. (3) Elucidating the counteractive regulation of energy homeostasis by crustacean cardioactive peptide (CCAP) and Myosuppressin (MS), which is revealed by their alterations in food intake, excretion, carbohydrate and lipid level in hemolymph. (4) As main hemolymph carbohydrates in cricket, trehalose and glucose levels were evaluated for their influences on energy homeostasis. Abnormal carbohydrate concentration was utilized to evaluate endocrine network activity. Collectively, I described the prototype of endocrine network that is essential for energy homeostasis in the circumstance of diet insufficient, in which neuropeptides are considered as nodes

and communication between them are considered as linkages.

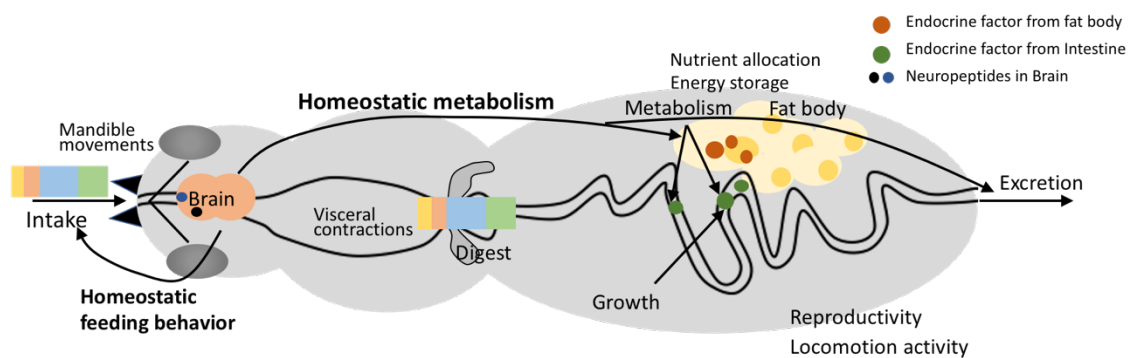
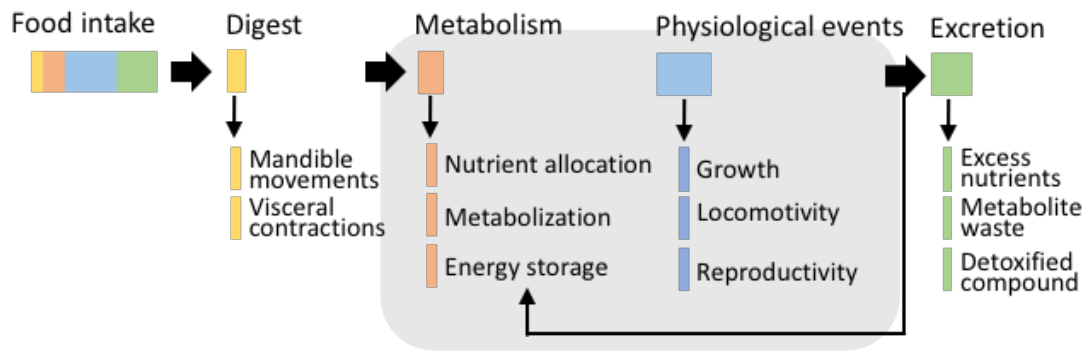


Figure 0-1 Physiological flow of energy and its maintaining mechanism by endocrine network

(A) Dietary nutrient flow in insect body (Nagata and Zhou, 2020). Dietary source changes by several feeding processes from ingestion to excretion. All steps include further detailed physiological processes. Harmonization of all processes accomplishes to obtain nutrients effectively and to maintain energy homeostasis.

(B) Schematic endocrine network of how to maintain energy homeostasis along with feeding behavior in insects. The stored energy in fat body and free energy in intestine give endocrinal signals, which target at central nervous system and activate homeostatic feeding behavior and metabolic flux. The metabolic flux and signals in brain control the balance between energy storage and excretion of excess energy.



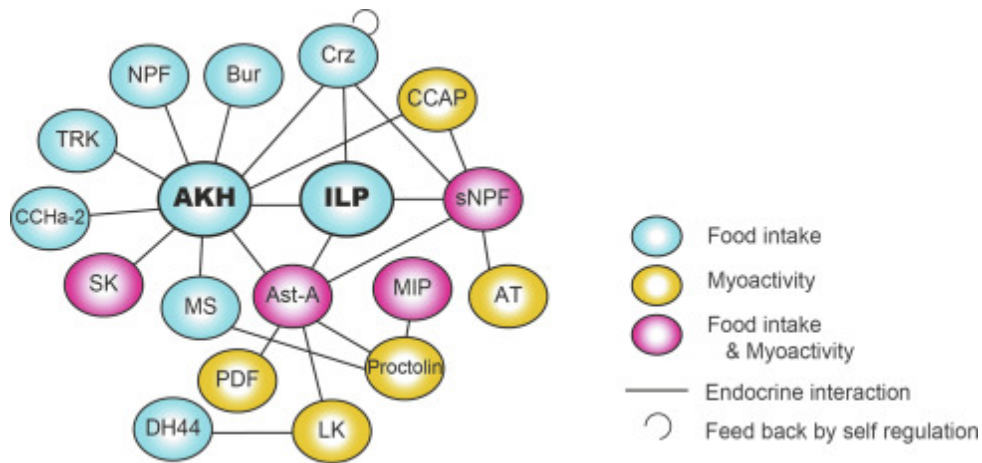


Figure 0-2. Schematic simple network graph generated by knowledges of neuropeptides and peptide hormones related to food intake (Nagata and Zhou, 2020).

All nodes are corresponding to neuropeptides and peptide hormones, which are selected by the previous literatures related to food intake in insects. –AKH, adipokinetic hormone; Ast-A, allatostatin-A; AT, allatotropin; Bur, bursicon; CCAP, crustacean cardio-active peptide; Crz, corazonin; DH44, diuretic hormone 44; ILP, insulin-like peptide; MIP, Myoinhibitory peptide, Ast-B, allatostatin-B; MS, myosuppressin; NPF, neuropeptide F; PDF, pigment dispersing factor; sNPF, short neuropeptide F; SK, sulfakinin; TRK, tachykinin-related peptide.

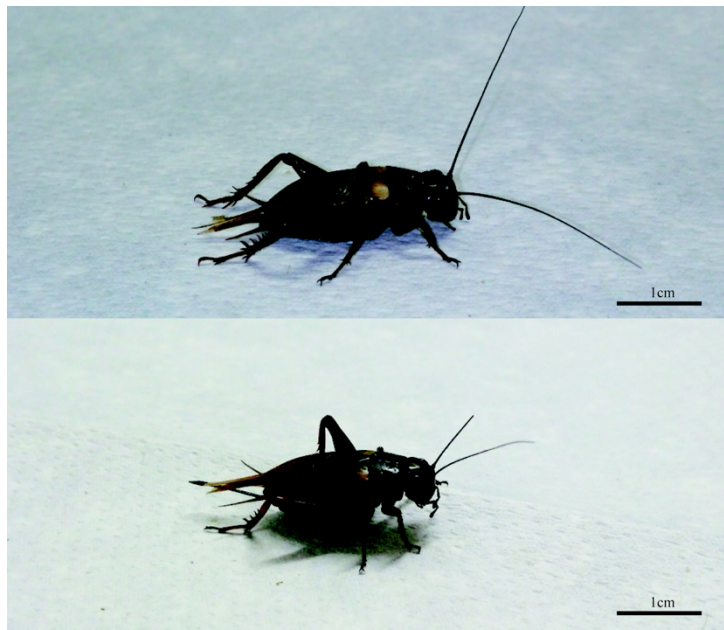


Figure. 0-3. The two-spotted crickets, *G. bimaculatus*; male (upper) and female (below).

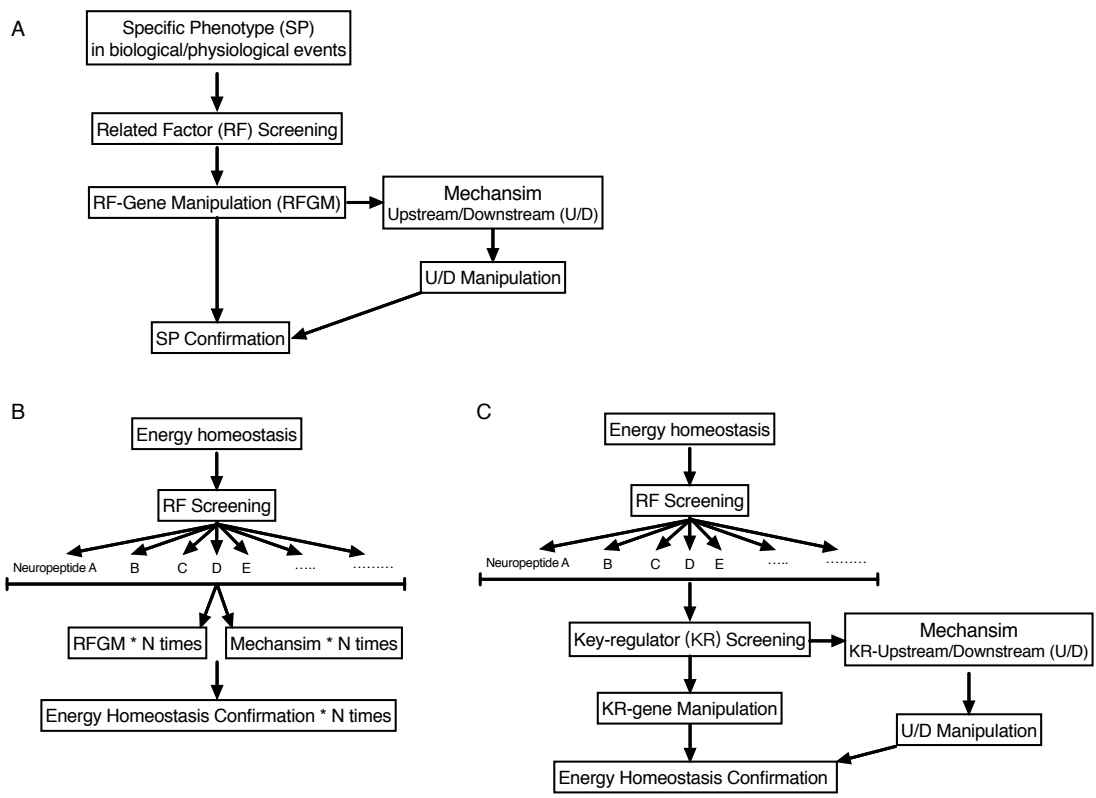


Figure 0-4. Schemes of conventional neuropeptide studies and in this study.

(A) Conventional neuropeptide studies. A specific phenotype is observed, and the endocrinal factors are screened for its regulatory role. Gene manipulation is used both for phenotype confirmation and for understanding its mechanism. The upstream/downstream signals are elucidated and their regulation on the specific phenotype are studied as well. (B) The summarized study scheme based on the classic neuropeptides studies. (C) The practical study scheme in this study. To avoid the unreachable experiments, key regulators screening is employed and manipulated.

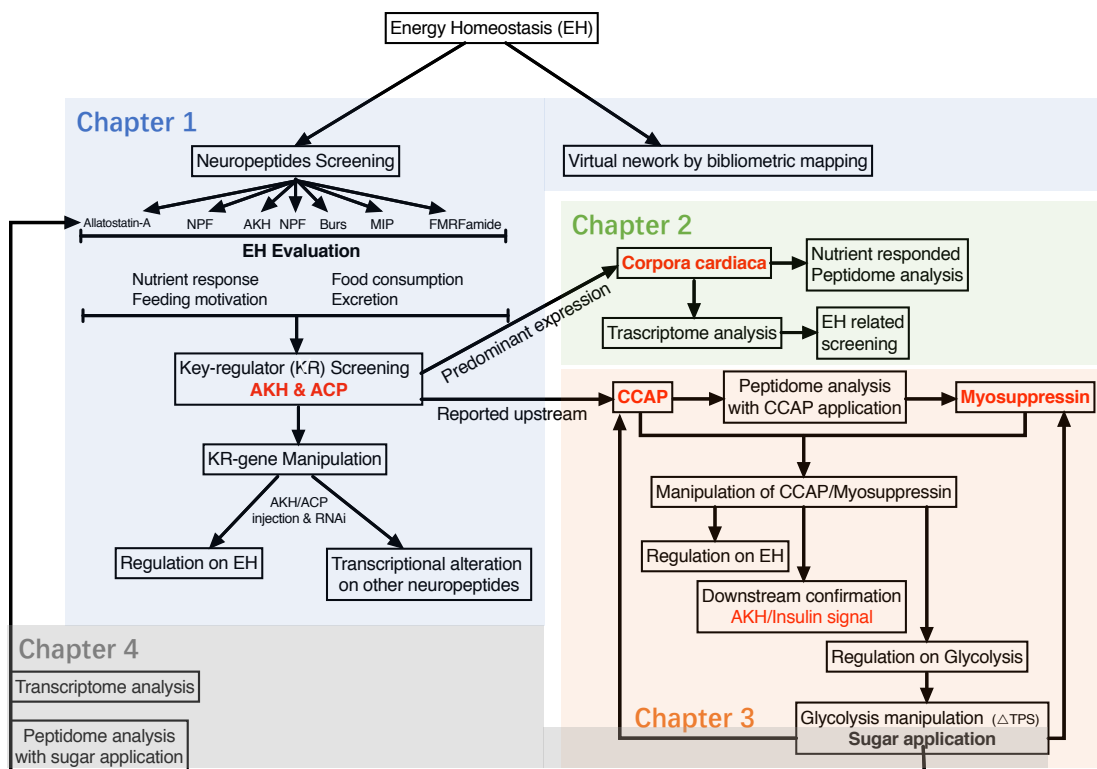


Figure 0-5. Schematic experimental flow in this study.

Red words represent important factors in each chapter.

AKH: Adipokinetic hormone; CCAP: Crustacean cardioactive peptide.

## Supplemental figures

A

```
1 GCCCCGCCGCCGCCGCTCCCGCCGCGCCCGCGCCCTCCGATCACGCCCCACGCCTCCAC
1 A P A A A A P A A P A P S D H A P R L H
61 AAGAGGCAAGCGGACCCCGCAGAGCTGGAGCGCCTCGCAGCGGAGCCCAAACGCAAGCGG
21 K R Q A D P A E L E R L A A E P K R K R
121 CCCTTCTGCAACGCTTCACCGGGTGC GGCAAGAAGCGCGGACGAGAGCCTGGGCACG
41 P F C N A F T G C G K K R A D E S L G T
181 CTGGTGGAGCTGAACTCGGAGCCCGCCGTGGCGGAGCTCAGCCGGCAGATCCTGTCCGA
61 L V E L N S E P A V A E L S R Q I L S E
241 GCCAAGCTGTGGGAGGCCATCCAGGAGGCACGCGGGAGCTCCTGCGCCGCCGCCAGCAG
81 A K L W E A I Q E A R A E L L R R R Q Q
301 CAGCTGCAGACGAACCGCATCGCGGCCGACGGCCCGCTGCCGCTGCCGCTACCAGCTTC
101 Q L Q T N R I A A D G P L P L P L T S F
361 CGAAGCGGCGCGCCCGCTCGCCGCCCCGAGGCCCGCCCGCCGCCGACGCCGCGCC
121 R K R R A A L A A P E A A P A A D A A P
421 GCCC
141 A
```

B

```
1 TGCAGATGGCGGGAGGCCCGACTTGTATTTCGAGAGGACATTCAAGTGCTTGTAACGG
61 TTGTGGAAGATTTAGAATTGACGTGTCAATACACATACCTGCAAGTGGTGTGGATATTC
121 ATTATTTCTGGTAGTAGATTTTTTTTTCTGCTGGAATTGAAGAAATGCAATTTTCATCCA
1 M R R Q Q Q R P P S S A T S
181 TACATTACATCACAAAAATGCGACGACAGCAGCAGCGGCCCTCCGTCGTCGGCGACGTCG
15 A A A S A L A T A T P C S S G R W G T A
241 GCGGCGGCTTCGGCGTTCGGCGACGGCCACACCATGCTCAAGTGGACGGTGGGGCACCGBA
35 R A A A A A D T D A E T A A A A A T A A
301 AGGGCGGCGGCAGCAGCCGACACGGACGCCGAGACGGCGGGCGGGCGGCAACGGCGGGC
55 T P L L D D A A T V P A P D L D S R P
361 ACGCCGCTGCTGGACGACGCCGCGACGGTGC CGGCGCCGACCTCAACGACTCCCGGCC
75 D R E A Q N V T F S F Y F Y Q T E Q F
421 GACCGGGAGGCGCAGAACGTCACCTTCAACTCCTTCTACTTCTACCAGACGGAGCAGTTC
95 T V L W I L F A L I V L G N L A V L A A
481 ACCGTGCTGTGGATCCTGTTCGCGTGTATCGTGTGGGCAACCTGGCCGTGTGGCGGGC
115 L A M N K R R K S R M N F F I M Q L A L
541 CTCGCCATGAACAAGCGCCGCAAGTCGCGCATGAACTTCTTCATCATGCAGCTCGCCCTC
135 A D L S V G L I S V L T D I V W R H T I
601 GCAGATTTGTCGGTGGGGCTGATCAGTGTGCTCACCAGACATCGTTTGGCGTCATACCATC
155 A W N A G N A A C K V I R F L Q A V V T
661 GCCTGGAATGCAGGAAATGCCGCTGCAAGGTCATTCGCTTCTGCAGGCTGTGCTCACC
175 Y S S T Y V L V A L S I D R Y D A I T H
721 TACTCGTCCACCTACGTGCTGGTGGCGCTTAGCATCGATCGCTACGACGCCATCACCAC
195 P M F S G S W R R A R L L V G V A W V
781 CCCATGAACTTCTCTGGCAGCTGGCGGCGCGCGGGCTGCTGGTGGGAGTGGCGTGGGTG
215 I S A L F S V P I L F L Y Q E Q P V E G
841 ATCAGCGCCCTCTTCTCGGTGCCATCCTTCTTCTTCTACCAGGAGCAGCCCGTTGAAGGT
235 H L Q C W I D F E K Q W M W Q L W V T L
901 CACCTCCAGTGTGGATCGACTTCGAGAAGCAGTGGATGTGGCAGTTGTGGGTGACGCTG
255 V A L T L F V L P A F I I S A C Y I V I
961 GTGGCCCTGACGCTGTTCTGCTGCTGCCCCCTTCATCATCTCGGCGTGCTACATCGTCATC
```

275 V S T I W T K S K Q L T P D P T R R T S  
 1021 GTCTCCACCATTTGGACCAAGAGCAAGCAACTCACGCCGATCCCACGCGCCGCACGTCA  
 295 R S S V Q Q L Q Q T H Q Q L Q L Q N S  
 1081 CGCAATAGTTCGTCCAGCAACTGCAACAGACACATCAGCAACTACAGCTGCAAAACAGT  
 315 G K P R G V A S R L Q Q L Q Q E D Q D S  
 1141 GGCAAGCCGCGAGGTGTGGCTTCGCGATTGCAACAAC TTCAGCAAGAGGACCAGGACAGT  
 335 R R A S S R G I I P K A K V K T V K M T  
 1201 CGAAGAGCTTCTTCGCGAGGAATCATTCCAAAGCAAAGTCAAGACTGTAAAGATGACC  
 355 F V I V I V F V L C W S P Y F V F D L L  
 1261 TTTGTAATTGTAATAGTATTTGTACTCTGCTGGAGTCCATATTTTCGTATTTGACCTACTT  
 375 Q V Y G Y V P R T Q T N I A V A T F I Q  
 1321 CAAGTATATGGGTATGTTCCGAGGACACAGACCAACATTGCTGTAGCTACTTTTCATTACG  
 395 S L A P L N S A A N P V I Y C L F S T T  
 1381 AGTTTGGCTCCCTAAATTTCTGCAGCAAATCCTGTCATCTATTGTCTTTTTCTCTACTACA  
 415 I G R T L R K I P P I S W A V A A L Q P  
 1441 ATTGGTCGAACACTCAGGAAAATCCCACCAATCAGTTGGGCAGTAGCTGCTCTCCAGCCT  
 435 C C P G L R P V P P D E E E P A L L D A  
 1501 TGCTGTCCCGTGTGCGCCCTGTCCCTCCTGACGAGGAGGAACCAGCTCTTTTAGACGCA  
 455 R M R W A S Q H H K A H P H L D A A V R  
 1561 CGCATGAGATGGGCATCTCAACATCACAAAGCGCATCCACATTTGGATGCAGCAGTGAGA  
 475 R G T A V A A V V S I V \*  
 1621 AGAGGTAAGTCTGTTGCAGCAGTGGTTTCTATTGTGTAAAGAGACAGTGATTTAATACTG  
 1681 CTCATTTTCTATAAGAGATTCTCAGAAATGTGGACATGAGTGTATGTCAAAGAATGACT  
 1741 GACTGGAGGTACCTGCAAGTTATTATACTAGTGCACACACTTTCTCAAATTTGTATTAGT  
 1801 TCAAGAGTGCAAATTTTCTAAAAACGCTCAATGCAAGGAAGATGATTTACATGCAAAGTT  
 1861 TATTTCTCAGCTTAAAAAGTTCAGTTTTCATAGTTTCTCAACAGATCTGTTCTATTCCAGA  
 1921 TGAAGCCTCATCTGTAAAAATTTGTTATATCCAGGTAGTGTAAACATACAAAGAAATAATA  
 1981 CAGGACCTTCAAAAATTTCTGAATTATCCAACATATGGCAATGTTACCTTTTCTCTATTCA  
 2041 AAAAAGGTGCAATAAATATGTGGTAAAATACATGCAGGGAAAGCCAGGAGACAT

C

1 P S P T P A N A G S Y L S Y P L L S  
 1 CCCGTCCCCACCCCGCCAACGCCAAGGCTCCTACAACCTGAGCTACCCGCTGCTGTGCT  
 21 T V A S A S A V P D L N S F Y F Y E T E  
 61 CACCGTTGCCTCCGCTCCGCGAGTCCCGGACCTCAACAGCTTCTACTTCTACGAGACTGA  
 41 Q F A V L W I L F T L I V L G N S A V L  
 121 GCAGTTTGCCTGCTGTGGATCCTCTTACCCTGATCGTGTGGGTAACCTCTGCTGTGCT  
 61 V A L F L N K N R K S R M N F F I M Q L  
 181 TGTGCACTTTTCTCAACAAAAACAGAAAATCGCGAATGAACTTCTTTCATCATGCAGCT  
 81 A L A D L T V G L I S V L T D I V W R I  
 241 CGCACTAGCCGATCTGACCGTGGGGCTGATCAGCGTGCTGACGGACATCGTGTGGCGCAT  
 101 T V A W H A G N A A C K I I R Y L Q A V  
 301 CACCGTGGCGTGGCACGCGGGCAACGCAGCCTGCAAGATCATACGCTATCTGCAGGCCGT  
 121 V T Y S S T Y V L V A L S I D R Y D A I  
 361 GGTCACTACTCGTCCAGTACGTGCTGGTGGCGCTGAGCATCGACCGCTACGACGCCAT  
 141 T H P M F S G S W R R A R V L V A V A  
 421 CACGCACCCCATGAACTTCTCCGGCAGCTGGCGGGCGGGCGCGTGTGGTGGCTGTGCGC  
 161 W F L S A I F S I P I I V F Y Q E Q S V  
 481 CTGGTTCCCTCAGCGCCATCTTTTCCATCCCGATCATCGTCTTCTACCAGGAGCAGTCGGT  
 181 Q G H Q Q C W I D F P E Q W Q W Q L Y M  
 541 GCAAGGGCACCAGCAGTGTGGATCGACTTCCCCGAGCAGTGGCAGTGGCAGCTATACAT

201 T L V A V L L F V F P A I I I S F C Y T  
 601 GACGCTGGTGGCCGTGTTGCTGTTTCGTGTTCCCGGCCATCATCATCTCCTTCTGCTACAC  
 221 V I V F T I W S K S K L L T P S G R P R  
 661 CGTCATCGTCTTCACCATCTGGAGCAAGCAAGCTGCTCACCCCGTCCGGTCGCCCCGG  
 241 G A T R N S E R R A L R Q D D Q D S R R  
 721 CGGGGCTACGCGCAACAGCGAGCGGCGCGCTGCGCCAGGACGACCAGGACAGCCGCCG  
 261 A S S R G L I P R A K I K T V K M T F V  
 781 CGCGTCGTCGCGGGACTTATCCC GCGGCCAAGATCAAGACGGTCAAGATGACCTTCGT  
 281 I V F V F I L C W S P Y F V F D L L Q V  
 841 CATCGTCTTCGTGTTTCATCCTGTGCTGGAGCCCCCTACTTTCGTGTTTCGACCTGCTGCAGGT  
 301 Y G H V P S T Q T N I A V A T F I Q S L  
 901 GTACGGCCACGTGCCGTCCACGCAGACCAACATCGCCGTGGCCACTTTCATCCAGAGCCT  
 321 A P L N S A A N P L I Y C L F S T H I C  
 961 GGGCCCGCTCAACTCGGCCGCAACCCGCTCATCTACTGCCTCTTCTCCACGCACATCTG  
 341 R T L R K I P P F S W V A A G L S L C F  
 1021 CCGCACGCTCAGGAAGATCCCCGCCCTTCAGCTGGGTGGCGGCCGGGCTGTCGTTGTGCTT  
 361 P A L R S G D G R C L R Y A T G D S S S  
 1081 CCCGGCGCTGCGTTCGGGTGACGGGCGCTGCTTGGCGTACGCCACGGGCGACTCCAGCTC  
 381 T V T E T L T Q Q S S R R S T S L R H T  
 1141 CACCGTCACCGAGACGCTCACGCAACAGTCTCGCGCCGCTCCACCTCGCTGCGCCACAC  
 401 M Q V R L P V S G A A V G A C G A T S S  
 1201 CATGCAGGTGCGCCTGCCTGTGTCGGGCGCGGCGGTGGGCGCGTGGGGGGCGACCAGCTC  
 421 H R R K V A V S V V \*  
 1261 GCACCGCCGCAAGGTGGCCGTGCTGTGGTGTAGGCGCCCGGGCCAGCCTCCGCCCCGA  
 1321 CCGCGACGCCGCTGGTGAAGGAAGTGGCGAGCGGCATGCACCGGCGCTGTGCACCGCTT  
 1381 TCTGCTCTACACTTCAACGCCGAGTTCATCCGGAATGGATTACGCAGCTTACCCCTCCA  
 1441 GTGGTCATAGGTTGGATCACCTCTGCGGTTTCGCAGCCGGGCAGATGGGAATGCGAGCG  
 1501 GTTTGAATCGGAGCTGTGCACCACTTGGTCTCTATTCCTCAAGGCAGTGTTCAGCCAAA  
 1561 ATGGATTTCATGGATCTGCTTCGCCCTCAAATGGTCTTAGTGTGGAATTGTATTGTTGTTT  
 1621 AATGTAAGAATCGGTTATCAAGTGGTTTCGGAGTACGAAGTGTCTATGACTTCTACAAT  
 1681 GCAAGGGAAAAGTTAGTTCTTTAAGATTTGGAAAATATAATATCCATAGTAAAAACCTTCA  
 1741 ATGTGGTTATTGGGTCTATTTAGTGATGTATGAAGTCTGACCTGTGACTCTGAACGTGTA  
 1801 CCTCAATTAAACCACCAATATCATTTGCGATCTGTGGATATTTTGAAAACCTGAATTAGT  
 1861 AAGTTCAATTTCTGCACGAAGCAGACCTATTGTATCATTTGTATATGTGTGTGAAAACCT  
 1921 AACGCAACACAGTGAACCGCAAGC

Figure S1 cDNAs and deduced amino acid sequences of GbCCAP (A), GbCCAPR-1 (B) and GbCCAPR-2 (C).

The mature sequence of GbCCAP is shallowly shaded in grey. The cleavage site is wavy underlined. The transmembrane areas of receptors are darkly shaded in grey. The putative glycosylation sites in receptors are shaded in black.

A

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1      M T K G Q E L A V V T E E L L H G K
1 CGCCGCCATGACGAAGGGCCAGGAGCTGGCGGTTCGTACCCGAGGAGCTGCTCCACGGGAA
101 P K Q S F G S Q P L Q P Q P L Q G L G R
301 GCCGAAGCAGAGCTTCGGCTCGCAGCCGCTGCAGCCGAGCCACTGCAGGGCCTCGGCCG
121 Q M F A S V A V A L G S M L V G F S S A
361 ACAGATGTTTCGCGTTCGGTGGCGGTGGCGTTGGGCTCGATGCTCGTGGGCTTCTCTTCCGC
141 Y T S P A L V S M A E P N S T L S V T P
421 CTACACGTGCCCCGCGCTCGTGTCCATGGCGGAGCCCAACAGCACGCTCTCAGTCACCCC
161 S Q G S W V G S L M P A A A L F G G M A
481 GTCGCAGGGCTCGTGGGTGGGCAGCCTGATGCCGGCAGCGGCGTGTTCGGAGGGATGGC
181 G G F L I E W L G R K T T I L A T A P P
541 GGGCGGTTTTCTCATCGAGTGGCTGGGCCGCAAGACTACCATCCTCGCCACCGCGCCGCC
201 S A I A W L L I F G A S N V G M L Y A G
601 GTCCGCCATTGCGTGGCTTCTCATCTTCGGCGGAGCAACGTGGGCATGCTGTACGCGGG
221 R I L S G F C V G V T S L S L P V Y M G
661 CCGCATCCTCTCGGGCTTCTGCGTGGGCGTCACGTCCCTCAGCCTGCCCGTACATGGG
241 E T I Q P E V R G M L G L I S T T L G N
721 CGAGACCATTAGCCCGAGGTGCGAGGCATGCTTGGCCTCATCTCCACAACACTAGGCAA
261 I G I L L C F V L G K F L D W R Y L A M
781 CATCGGCATTCTGCTGTGCTTCGTCTGGGCAAGTTCTCGACTGGCGCTACCTGGCCAT
281 M G A L C S L P Y G L C T M F A P E T P
841 GATGGGCGCGCTGTGCTCGCTGCCGTACGGGCTCTGCACCATGTTTCGCCCCGAAACGCC
301 H W Y L S K G R R G D A E T A L R W L R
901 GCACTGGTACCTCTCAAAGGGCGGGGGCGACCGGAGACGGCGCTGCGCTGGCTGCG
321 G G A G A D V A Q E L R A T E Q G L G K
961 CGGCGGCGGGGCGCCGACGTGGCGCAGGAGCTGCGCGCCACCGAGCAGGGCCTGGGCAA
341 G A E A A I G L P R S V T S I N L Q A Q
1021 GGGCGCCGAGGCCGCCATCGGCCTGCCGCGCAGCGTCACCAGCATCAACCTGCAGGCGCA
361 A A P S D K S G L A L L F T R D N M R P
1081 GGCCGCGCCCTCCGACAAGAGCGGCCTGGCGCTGCTCTTCACGCGGACAACATGCGGCC
381 V G V A L G L M L F Q Q F S G I N A V I
1141 CGTCGGCGTTCGCGCTGGGCTCATGCTCTTCCAGCAGTTCAGCGGCATCAACGCCGTCAT
401 F Y T V K I F Q D A G S T I D E F V S T
1201 CTTCTACACGGTCAAGATCTTCCAGGACCGGGCAGCACCATTGACGAGTTCGTGAGCAC
421 M V V G A V N L G A V V L A T A L I D R
1261 GATGGTGGTGGGCGCGGTGAACCTGGGCGCCGTGGTGTGCGGACGGCGCTCATCGACCG
441 L G R R V L L A A S A A A M A L A L A T
1321 CCTGGGGCGGGGCTGCTGCTGGCGGCGTCGGCGGCCCATGGCGCTGGCGCTCGCCAC
461 L G T F F Y L S T C P D V D V S G F G W
1381 GCTGGGCACCTTCTTCTACCTCAGCACCTGCCCCGACGTCGACGTCAGCGGCTTCGGCTG
481 L P L V S F V I F V V G F S V G F G P I
1441 GCTGCCGCTGGTCAGCTTCGTTCATCTTCGTGGTGGGCTTCTCCGTGGGCTTCGGGCCCAT

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501 P W L M M G E I L P P D I R G P A A S L  
 1501 CCCCTGGCTCATGATGGGCGAAATTTTACCACCGGACATCCGCGGGCCGGCGGCGTCGCT  
 521 A T A V N W S C T F M V T K M F A D V V  
 1561 GGCGACGGCCGTCAACTGGTTCGTGCACGTTTCATGGTGACGAAGATGTTTCGCGGACGTGGT  
 541 E G V G S H A A F W G F C A V C V G A F  
 1621 GGAGGGCGTGGGCAGCCACGCGGCCTTCTGGGGCTTCTGCGCCGTGTGCGTGGGCGCCTT  
 561 V F V W A C V P E T R G R T L A D I Q R  
 1681 CGTGTTTCGTGTGGGCGTGCCTGCCCCGAGACGCGCGGCCGCACGCTCGCAGACATCCAGCG  
 581 R M A G R P A P Q R K R R L T V S E S V  
 1741 GCGCATGGCCGGGCGGCCGCCCCGAGCGCAAGCGCCCTCACCGTCAGCGAGTCGGT  
 601 A G W T A S M A S L K P M P T G A \*

**B**

1 M A D V A S S T G S M I V V S N R L P F  
 1 ATGGCTGATGTAGCGTCGTCCACTGGCAGTATGATCGTAGTATCGAACAGATTACCTTTC  
 21 V L K R N A L T Q L L E R K A S A G G L  
 61 GTTTTAAAGCGAAATGCACTGACACAATTATTGGAACGAAAGGCCAGCGCCGGCGTCTT  
 41 V T A V A P V V I Q S G G L W V G W P G  
 121 GTAAGTGTGTAGCACCTGTAGTAATACAGAGTGGCGGATTGTGGGTGGGCTGGCCAGGT  
 61 I H L N N P N E P I P E S D P N D K T P  
 181 ATTCACTTGAATAATCCAAATGAGCCCATACCTGAATCAGACCCAAATGATAAAACACCC  
 81 T A G L L S K K V V S V H I N A D V F D  
 241 ACTGCTGGACTTCTGTCAAAGAAGGTTGTTTTCAGTTCATATAAAACGCTGATGTGTTTGAT  
 101 S Y Y N G C C N G T F W P L F H S M P D  
 301 TCATATTACAATGGTTGTTGTAATGGTACATTTTGGCCCTTGTTTTCACTCAATGCCAGAT  
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 421 GATAAAACCATTGAAGCATTGACAAGCTGATAGCTGAACAGGGAAACAGTGCCGGAAC  
 161 P L I W I H D Y H L M L A A N W V R Q V  
 481 CCGTTAATATGGATTTCATGATTACCATTTGATGTTGGCTGCAAATTGGGTTAGACAGGTG  
 181 A E E R D L R C K L G F F L H I P F P P  
 541 GCTGAAGAAAGGGATCTTCGTTGCAAATTGGGTTTCTTTTTTACATATTCCCTTTTCCCCCA  
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 281 A P K V M L T N Q K I I L G V D R L D Y  
 841 GCTCCCAAGGTAATGCTGACCAATCAGAAGATTATCCTTGGTGTGATCGCTTAGATTAC

301 T K G L V H R L K A I E T L F E K Y P E  
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321 H I E K A T M L Q I S V P S R T D V K E  
961 CACATTGAAAAGGCAACAATGTTGCAAATATCCGTCCCTTCACGCACAGATGTGAAAGAG  
341 Y Q D L K E E M D Q L V G R I N G R F T  
1021 TATCAAGATCTGAAGGAAGAAATGGATCAACTTGTGGGACGCATCAATGGGCGTTTCACA  
361 T Y N W S P I R Y I Y G C V S Q D E L A  
1081 ACTTACAACCTGGTCTCCTATTTCGTTATATATATATGGTTGTGTCAGCCAAGATGAATTGGCA  
381 A F Y R D A A V A L V T P L R D G M N L  
1141 GCTTTTTACCGGGATGCAGCTGTAGCTCTTGTTCACACCCTTGAGAGATGGCATGAACCTG  
401 V A K E F V A C Q I N E P P G V L I V S  
1201 GTGGCCAAGGAGTTCGTGGCGTGCCAGATCAACGAGCCGCCGGCGTGCTCATCGTGTGCG  
421 P F A G A G E M M H E A L I C N P Y E I  
1261 CCCTTCGCCGGCGGGGCGAGATGATGCACGAGGCGCTCATCTGCAACCCCTACGAGATC  
441 N A A A E V I H R A L T M P E D E R T L  
1321 AACGCCGCCGCCGAGGTCATCCACAGGGCTCTTACAATGCCAGAAGATGAAAAGGACCTTG  
461 R M N Y L R R R E K L H D V N Y W M R S  
1381 CGAATGAACTACCTGAGAAGGCGAGAGAAGCTTCATGATGTGAACTACTGGATGAGGTCT  
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1441 TTCCTCAAAGCGATGGGTTCTCTCATTGAGGAAGATGGAGACGAAGTCAGGCCACTACG  
501 M Q P V T M D D F D E Y L A K Y I G H T  
1501 ATGCAACCTGTCACAATGGATGACTTCGATGAGTACCTTGCTAAATACATTGGGCACACA  
521 H K L A L L L D Y D G T L A P I A P H P  
1561 CATAAATTAGCTCTACTGTTGGATTATGATGGCACCCCTGGCACCTATTGCCCCACATCCT  
541 D L A I M P E E T K N V L E R L S N M P  
1621 GATCTGGCTATCATGCCTGAAGAAACCAAGAATGTTCTTGAGAGATTGTCCAATATGCCA  
561 D V Y I A I I S G R N V N N V K S M V G  
1681 GATGTGTATATTGCTATCATATCTGGACGAAATGTCAACAATGTGAAATCCATGGTTGGG  
581 I E G I T Y A G S H G L E I L H P D G S  
1741 ATTGAAGGTATTACGTATGCTGGTAGTCATGGTTTGGAAATCCTTCATCCTGATGGAAGC  
601 K F V H P M P V E F E D K V T E L L K A  
1801 AAGTTTGTACATCCCATGCCTGTGGAATTTGAAGACAAAGTTACTGAATTACTAAAAGCT  
621 L Q D Q V C K D G A W V E N K G A L L T  
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641 F H Y R E T P V Q K R E A L V S K A Q Q  
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661 M I V A A G F K A G A A H C A L E A K P  
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721 M E A L K G M A A T F R V T S S H I V K

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 2281 GAACGACACTTTTCTAGGCGTGAAGTACGGCGGGACCC TGAGTCAAATCTCCTGTACAGA  
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 801 A V T P E P L \*  
 2401 GCTGTGACACCAGAACCATTGTAG

C

1 TTGACATTTTAAAGTGCCTCCACTCAGCATTCTCGGAACTTGTGCGAAAGCGTTCCGGGAAA  
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 18 N T E K L P P P C E S E I Y C H G D L L  
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 181 CATTACGTGCAGTTGGGAAGTGAATACGAGATTCTAAAGAATTTGTAGACATGAAAAATG  
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 118 Q D P D F R R F A E H L N S L W K S L G  
 421 CAAGATCCAGACTTCCGTGCGATTTCGAGAACATTTAAATAGTCTCTGGAAATCATTGGGC  
 138 R K M K D A V K Q S P E F Y S I I Y V P  
 481 CGTAAAATGAAAGATGCAGTTAAACAAAGTCTGAATTCTATTCCATTATCTATGTGCCT  
 158 H P V I V P G G R F R E F Y Y W D S F W  
 541 CATCCTGTAATTGTACCTGGAGGTCGGTTT TAGAGAATTTTACTATTGGGATTCATTTTGG  
 178 I I R G L L L S E M Q S T V K G M L E N  
 601 ATTATCAGAGGTTTACTTTTTGTGAGAAATGCAGTCAACTGTCAAGGGAATGCTAGAGAAC  
 198 F L S I V E R F G F I P N G G R I Y Y A  
 661 TTTTATCCATTGTTGAAAGGTTTGGATTTATACCCAATGGAGGTCGTATATATTACGCA  
 218 M R S Q P P F L I P M V Q T Y V D T T Q  
 721 ATGCGTTCACAACCACCATTCTCATACCGATGGTTCAA ACTTATGTAGATACCACTCAG  
 238 D Y E F I E K N I D T L D K E F Q H W M  
 781 GATTATGAGTTTATTGAAAAGAATATAGATACTCTGGATAAAGAATTTCAACATTGGATG  
 258 V N H T V N I K K D G K V Y T L A R Y K  
 841 GTGAATCACACAGTTAACATTA AAAAGATGGAAAAGTGTACACTCTAGCAAGATACAAG  
 278 D N S N G P R P E S Y K E D Y E S A H F  
 901 GACAATTCTAATGGCCCAAGACCCGAGTCATACAAA GAAGATTATGAGAGTGCGCACCTTC

298 F H T E E D K E N Y Y S E L K A A A E S  
 961 TTTCACACTGAAGAAGACAAAGAAAATTACTACTCTGAACTTAAAGCAGCTGCTGAATCT  
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 378 D I A E K L K E A V T A V L W H K E V G  
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 398 A W L D Y D M I N D K R R D Y F Y P T N  
 1261 GCTTGGTTAGATTATGATATGATAAATGATAAACGACGTGATTATTTCTATCCAACCAAT  
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 1321 ATTTCTCCTTTATGGACTGGCTGCTATGATACAACACACAAAAGAGCAATATGTAGGACGT  
 438 V L K Y L E H S R A T V L L G G I P T T  
 1381 GTACTCAAATACTTGGAGCATAGTCGTGCCACAGTACTCCTAGGAGGAATCCCAACAACA  
 458 M E L S N E Q W D Y P N A W A P L Q H L  
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 1681 GGATTTGGATGGTCCAATGGGGTTGTCATGGATTTCTTGGATAAATATGGTCACAACATA  
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 578 T K E S Q I L T A I Y A L L A T L A A G  
 1801 ACCAAAGAAAGTCAGATCCTGACAGCTATTTATGCTCTACTTGCCACACTTGCGGCAGGA  
 598 S I G \*

D

1 Q G E L L S V V Q N A H L Y P D S K K F  
 1 CAAGGCGAGCTCCTCAGTGTCTGTCAGAACGCTCACTTGTATCCCGACTCGAAGAAATTC  
 21 V D K R L I Y S P D E T L E Q F D A L M  
 61 GTCGACAAGCGGCTCATCTACTCGCCGACGAGACGCTGGAGCAGTTTCGACGCGTTGATG  
 41 K Q T G G N P S K E Q I Q K F V D D Y F  
 121 AAGCAAACCGGAGGCAACCCGTCCTCAAGGAGCAAATCCAAAAGTTCGTAGACGATTACTTC  
 61 E D G N E L E G A D L P D W V A E P A L  
 181 GAGGACGGCAACGAGCTGGAGGGGGCGGACCTGCCCCGACTGGGTGGCCGAGCCGGCGCTG  
 81 L R R I A D P K L R A W A S E L N G I W

241 CTCCGCCGTATCGCCGACCCCAAGCTGCGCGCCTGGGCGTCCGAGCTGAACGGCATATGG  
101 K N L S R R V S D E V R D H T D R Y S L  
301 AAGAATCTCAGCAGAAGGGTTTCCGACGAAGTCAGGGACCACACGGACAGATACTCGCTC  
121 I Y V P H H F I V P G G R F R E L Y Y W  
361 ATCTACGTTCCCTCACCCTTCATCGTGCCAGGCGGGCGCTTTAGGGAGCTGTACTATTGG  
141 D T Y W I V R G L L L S D M K D T V K G  
421 GATACCTACTGGATTGTTTCGAGGGCTTCTGCTTAGCGATATGAAGGACTGTAAAGGGT  
161 I I E N F L F L L D T H G I I P N G A R  
481 ATAATAGAAAATTTCTTGTTCCTGCTGGACACCCATGGAATAATTCCTAATGGAGCAAGA  
181 I Y Y L E R S Q P P L L T P M V Y N Y F  
541 ATTTACTACTTGGAACGCTCGCAACCCCCGCTTCTGACTCCCATGGTCTACAATTACTTT  
201 L A T K D E E F V K K N I H L L E R E L  
601 CTGGCTACCAAGGATGAAGAATTTGTGAAGAAAAACATCCACTTGCTTGAAAGAGAATTA  
221 D F W M K N R T V N V H K D G K T Y T L  
661 GATTTCTGGATGAAAAATCGTACAGTAAACGTGCACAAGGATGGAAAAACCTTACTCTT  
241 L R Y Y S P S Q G P R P E S Y R E D Y M  
721 CTCAGATACTATTCTCCATCGCAGGGCCCTAGACCAGAATCATAAGGGAGGATTACATG  
261 N G Q V F S S D Q R K E E F Y I D L K S  
781 AACGGCCAAGTATTCTTCTGATCAAAGAAAAAGAATTTTACATAGACTTGAAATCT  
281 A A E S G W D F S S R W F I K D G S N K  
841 GCTGCAGAATCTGGTTGGGATTTCTCCAGTCGATGGTTCATCAAAGATGGTAGCAACAAA  
301 G N L T D I H T K Y I I P V D V N A F V  
901 GGAAACTTGACAGACATTCATACAAAATATATAAATTCAGTAGATGTCAATGCTTTTGTG  
321 Y G N A R L L S T L H T E V V G N L D K  
961 TATGGTAATGCTCGTTTGCTTTCTACATTGCACACGGAAGTTGTAGGAACTTGGATAAA  
341 A K K Y S Q I A H E L K E A V T A V L W  
1021 GCCAAAAAATATTTCCCAAATGCCCATGAACTGAAGGAGGCAGTCACAGCAGTATTATGG  
361 N G T T G S W F D Y D L L N A K Q R T Y  
1081 AATGGTACAACCTGGTTCTTGGTTTGATTATGATCTTCTGAATGCAAAACAGCGCACCTAC  
381 F Y P S N L A P L W T K C Y N E K E T P  
1141 TTTTACCCATCAAACCTTGCTCCTTTATGGACAAAAGTGTACAATGAGAAAAGAGACTCCC  
401 I I A Q K V V K Y L Q S I E I L Q H Y L  
1201 ATCATTGCACAAAAAGTAGTTAAATACTTACAAAAGTATTGAAAATTTTACAACACTATCTT  
421 G G I P T S L N N T G E Q W D L P N A W  
1261 GGTGGAATTTCCACATCACTGAATAACACAGGAGAACAATGGGATCTTCCAATGCCTGG  
441 P P L Q N I I I Q G L Q S T K E P S A Q  
1321 CCACCTCTCCAAAACATTATTATTCAAGGCTTGCAGTCCACTAAGGAACCTAGTGCCAG  
461 K L A Y E F A E Q W I R S N Y K G F V E  
1381 AAATTAGCTTATGAATTCGCTGAACAATGGATTTCGATCAAACCTACAAAGGATTTGTGGAA  
481 H H D M Y E K Y D A E V P G N S G G G G  
1441 CACCATGATATGTATGAAAAGTATGATGCGGAGGTTCCAGGAAATTTCTGGAGGAGGGGGA  
501 E Y I P Q S G F G W T N G V V L E L L D  
1501 GAATACATTCCACAATCAGGATTTGGATGGACAAATGGAGTGGTTCTTGAACCTATTGGAC

521 T Y G T R L T A D N T M R R E V \*

E

1 M S V Q S D  
1 ATGTCGGTTCAGAGCGAT  
7 I E K R K Q I S V R G I A Q V E N V S T  
19 ATCGAGAAGCGCAAGCAGATCAGCGTCCGGGGCATAGCCCAGGTGGAAAACGTCTCCACC  
27 I K K T F N R H L H Y T L V K D R N V A  
79 ATAAAGAAGACGTTCAACCGACACCTGCATTACACGCTCGTCAAGGACCGGAACGTTGCC  
47 T P R D Y Y F A L A H S V K D N L V S R  
139 ACCCCGCGAGACTACTACTTCGCGTTGGCCCACTCCGTCAAAGACAACCTGGTGTGCGAGA  
67 W I R T Q Q Y Y Y E K D P K R V Y Y L S  
169 TGGATCCGCACACAGCAGTACTACTACGAGAAGGACCCCAAGAGAGTTTACTACCTATCC  
87 L E Y Y M G R T L Q N T M I N L G I Q G  
229 TTGGAGTACTACATGGGCCGTACCCTACAGAACACCATGATCAACCTGGGCATACAAGGA  
107 A C D E A M Y Q L G L D I E E L E D L E  
289 GCCTGTGATGAAGCTATGTACCAGCTGGGCTTGGACATTGAGGAGTTGGAGGACCTGGAG  
127 E D A G L G N G G L G R L A A C F L D S  
349 GAGGATGCTGGTTTTGGGAAATGGAGGCCCTTGGTTCGTTTAGCAGCCTGCTTCCCTGGACTCC  
147 M A T L G L A A Y G Y G I R Y E Y G I F  
409 ATGGCCACACTCGGTCTGGCAGCGTATGGATACGGTATCAGATATGAGTATGGCATCTTC  
167 T Q K I R N G E Q V E E P D D W L R F G  
469 ACGCAGAAGATTCGTAATGGAGAACAGGTAGAGGAGCCGGATGACTGGCTGAGATTTGGA  
187 N P W E K A R P E Y M L P V N F Y G R V  
529 AACCCCTGGGAGAAGGCAAGACCAGAATATATGTTACCAGTTAATTTCTACGGTCGAGTG  
207 E Q T P Q G V K W V D T Q V I F A M P Y  
589 GAACAGACTCCTCAAGGCGTCAAATGGGTGGATACACAGGTGATATTTGCGATGCCCTAT  
227 D S P V P G Y G N N V V N T L R L W S A  
649 GACAGCCCTGTTCCAGGTTATGGTAATAATGTTGTGAACACATTGAGGCTGTGGTCTGCT  
247 K S P V E F N L K F F N D G D Y I Q A V  
709 AAATCCCCTGTGGAATTTAACCTCAAATTTTTTAATGATGGTGACTACATCCAAGCAGTA  
267 I D R N L A E N I S R V L Y P N D N F F  
769 ATAGATCGAAACTTGGCAGAGAACATCTCAAGAGTGCTGTATCCAAATGATAATTTCTTT  
287 E G K E L R L K Q E Y F M C A A T L Q D  
829 GAAGGAAAAGAACTTCGTCTTAAACAAGAGTATTTTCATGTGTGCTGCTACTTTGCAGGAT  
307 L I R R F K A S K F G V R T A V R T N F  
869 CTCATACGCCGTTTTAAGGCTTCAAATTTGGTGTCCGACTGCAGTGAGAACAAATTTT  
327 E H F P D K V A I Q L N D T H P S L A I  
929 GAACACTTCCCTGACAAGGTTGCCATTGAGCTGAATGACACACATCCATCACTGGCAATC

347 P E L M R I L I D V E G L T W E Q A W P  
989 CCAGAGCTTATGCGGATCCTCATTGACGTGGAAGGCCTGACATGGGAACAGGCATGGCCC  
367 I V V R T C A Y T N H T V L P E A L E R  
1049 ATTGTGGTTTCGCACATGTGCCTATACAAATCACACTGTGCTACCTGAAGCTTTGGAGCGA  
387 W P V H L L E T I L P R H L Q I I Y H I  
1109 TGGCCTGTTTACTTACTGGAGACCATCTTGCCACGCCATCTGCAGATCATCTATCACATT  
407 N F L H L Q E V G K K Y P G D L D R L R  
1169 AATTTTCCTTCACCTGCAGGAAGTGGGAAAAAATACCCCTGGAGATCTTGATCGTCTTCGG  
427 R M S L V E E G G E K R I N M A H L C I  
1229 AGAATGTCCCTTGTGGAGGAAGCGGAGAGAAAAGAATTAATATGGCTCATCTCTGCATT  
447 V G S H A V N G V A R I H S D I I K S D  
1289 GTGGGTTCTCATGCAGTCAATGGAGTTGCAAGAATTCATTCTGACATCATTAAGAGTGAC  
467 I F K D F Y E M T P E K F Q N K T N G I  
1349 ATCTTCAAAGATTTCTATGAAATGACTCCAGAGAAATTTCAAATAAGACAAATGGTATC  
487 T P R R W L L L C N P G L S D L I A E K  
1409 ACTCCTCGTCGCTGGTTGCTGCTATGTAATCCTGGATTATCTGATTTGATTGCAGAGAAA  
507 I G E E W I T H L E Q L Q Q L K E F A K  
1469 ATTGGCGAGGAATGGATTACTCACCTGGAGCAACTGCAGCAATTGAAAGAATTTGCAAAG  
527 D P G F Q R A V Q K V K Q E N K L R L A  
1529 GATCCAGGCTTTTCAGAGAGCTGTCCAAAAAGTGAAGCAAGAGAACAAATTGAGGCTGGCT  
547 Q I L E K D Y G V K V N P A S M F D I Q  
1589 CAAATTCTGGAGAAGGACTATGGTGTCAAGGTGAATCCAGCATCCATGTTTGACATTCAG  
567 V K R I H E Y K R Q L L N C L H I I T L  
1649 GTTAAACGTATTCATGAATACAAACGTCAGTTGCTGAATTGTCTGCATATAATAACACTG  
587 Y N R I K R D P S A N I T A R T V M I G  
1709 TACAATCGCATAAAGAGAGATCCATCTGCAAAATATCACTGCAAGAACTGTGATGATTGGA  
607 G K A A P G Y Y M A K K I I K L I N A V  
1769 GGCAAGGCTGCTCCTGGCTATTATATGGCAAAGAAGATTATTAAGCTGATCAATGCTGTT  
627 G N I V N N D P I V G D K L K V I F L E  
1829 GGCAACATTGTGAACAATGACCCCATTTGTTGGAGATAAATTGAAAGTAATTTTCCCTAGAA  
647 N Y R V T L A E K I M P A A D L S E Q I  
1889 AACTACAGAGTGACACTTGCTGAAAAGATAATGCCAGCTGCAGATTTGAGTGAACAGATC  
667 S T A G T E A S G T G N M K F M L N G A  
1949 TCCACTGCTGGTACAGAAGCATCTGGAACCTGGAAATATGAAGTTTATGTTGAATGGAGCC  
687 L T I G T L D G A N V E M A E E M G R E  
2009 CTCACAATTGGAACCTTGGATGGTGCCAATGTAGAAAATGGCTGAGGAAATGGGTCCGGAA  
707 N I F I F G M T V D E V E A L K K R G Y  
2069 AACATCTTCATCTTTGGAATGACTGTGGACGAAGTAGAGGCACTGAAGAAACGAGGATAC  
727 N A H D Y Y A A N P E I Q Q C V D Q I R  
2129 AATGCCCATGACTACTATGCTGCAAAATCCTGAGATCCAACAGTGTGTGGACCAGATCCGT  
747 N G F F S P E N P S E F A D V A D V L M  
2169 AATGGATTTTTTCAGTCCTGAAAACCCATCAGAGTTTGTGATGTGGCAGATGTGCTGATG  
767 K Y D R F L T L A D F D A Y I K C Q D T

2229 AAGTATGATCGCTTCCTCACTCTTGCAGACTTCGATGCATACATCAAGTGCCAAGATACA  
 787 V S A V Y Q D Q A K W S E M A I N N I A  
 2289 GTATCAGCTGTATACCAGGATCAAGCAAAATGGTCAGAGATGGCTATCAACAACATTGCT  
 807 S S G K F S S D R T I A E Y A R E I W S  
 2349 TCATCGGGCAAGTTCTCAAGTGACCGCACAATTGCTGAATATGCTCGTAAAATTTGGAGT  
 827 V E P S W E K L P A P H E P R D D D T A  
 2409 GTGGAACCATCATGGGAAAAGTTACCTGCTCCGCATGAACCACGTGACGATGACACAGCT  
 847 A N G L A A G G K \*  
 2469 GCAAATGGGCTAGCAGCAGGTGGAAAGTAG

F

1 G L T F F L S F A I L A A V L G M L Q F  
 1 GGGCTTGACGTTTTTCTATCTTTTGCAATACTGGCAGCAGTGTGGGGATGCTACAGTT  
 21 G Y N T G V I N A P E A N I E N F M K D  
 61 TGGTTATAATACTGGGGTCATAAACGCTCCCGAGGCGAACATTGAAAATTTTATGAAAGA  
 41 V Y K H R Y G E D M T D D T V K L L Y S  
 121 TGTGTACAAGCATCGCTATGGTGAGGATATGACTGACGATACCGTGAAGTTGCTATATTC  
 61 L A V S I F A I G G M F G G F C G G M I  
 181 CTTGGCAGTATCGATATTTGCCATTGGAGGCATGTTCCGGTGGCTTTTGTGGAGGAATGAT  
 81 A N K F G R K G G L L L N N I L G V L G  
 241 TGCCAACAAATTTGGACGAAAAGGTGGTTTTGTACTAAATAATATTTCTCGGAGTTCTTGG  
 101 A V L M G C T K E T Q W Y E V L F L G R  
 301 TGCTGTATTAATGGGTTGTACGAAAGAAACACAATGGTATGAAGTACTGTTTCTCGGGCG  
 121 L I I G V N C G L N T S L V P M Y I S E  
 361 TCTTATCATTGGTGTCAATTGTGGTCTAAATACATCATTGGTTCCAATGTACATCTCAGA  
 141 I A P L N L R G G L G T V N Q L A V T V  
 421 AATTGCTCCTCTCAATTTAAGAGGGGGTCTGGGTACCGTTAACCAATTGGCTGTGACTGT  
 161 G L L L S Q V L G I E Q I L G T D E G W  
 481 TGGCTTGCTGCTTTTACAAGTACTTGGCATCGAGCAAATTTCTTGGCACAGACGAAGGATG  
 181 P I L L G L A V C P A L L Q L T L L P F  
 541 GCCAATCCTTCTTGGTCTAGCAGTTTGTCCAGCACTTCTACAGCTGACTTTGTTGCCCTTT  
 201 C P E S P R Y L L I T K Q W E E Q A R Q  
 601 TTGCCCTGAGTCACCAAGATACTTGTCTATTACGAAGCAGTGGGAAGAACAAGCGAGGCA  
 221 A L R R L R A S N Q V E E D I E E M R A  
 661 AGCGCTACGGAGGTTGCGTGCTTCAAATCAAGTGGAGGAAGACATCGAAGAAATGCGAGC  
 241 E E R A Q Q A E A T I S M M E L I C S P  
 721 AGAAGAAAGAGCCCAGCAAGCGGAAGCCACGATTTCCATGATGGAATTAATTTGCTCGCC  
 261 T L R S P L I I G V V M Q L S Q Q L S G  
 781 GACACTTAGATCGCCTCTTATCATTGGGGTGGTAATGCAGCTTTCGCAACAACATATCAGG  
 281 I N A V F Y Y S T G L F V S S G L S E E  
 841 GATTAATGCTGTTTTCTATTATTCGACAGGTCTGTTTGTGAGCTCTGGACTGTCCGGAAGA



301 Q A K F A T I G I G S I M V V M T L V S  
 901 GCAAGCAAAATTTGCCACAATTGGTATAGGTTTCGATAATGGTGGTGATGACCCTTGTGTC  
 321 I F L M D R M G R R S L H L Y G L G G M  
 961 CATATTCTTGATGGATCGCATGGGTTCGGAGATCACTGCACCTGTACGGCCTGGGTGGCAT  
 341 F I F S I F I T I S F L I K E F F G Y V  
 1021 GTTTATCTTCTCAATATTTATCACCATCTCTTTTCCTTATAAAAGGAGTTTTTTGGTTACGT  
 361 Q D M I D W M S Y W S V V S T L S F V V  
 1081 CCAGGACATGATTGACTGGATGTCATACTGGTCTGTGGTATCTACGTTGAGCTTTGTGGT  
 381 F F A V G P G S I P W M I T A E L F S Q  
 1141 GTTCTTCGCTGTGGGTCCCTGGATCCATCCCGTGGATGATAACTGCGGAGCTCTTTTCCCA  
 401 G P R P A A M S I A V L V N W M A N F L  
 1201 AGGACCACGACCAGCAGCTATGTCCATTGCTGTATTGGTCAACTGGATGGCGAACTTCCT  
 421 V G I G F P S M N T A L E N Y T F L P F  
 1261 TGTTGGAATTGGATTTCCAGTATGAATACTGCCCTGGAAAACTACACATTTCTTCCCTT  
 441 S L F L A I F W I F T Y K K V P E T K N  
 1321 CAGCTTGTTTCCTTGCAATCTTTTGGATTTTACGTACAAGAAAAGTGCCAGAGACCAAGAA  
 461 K T F E E I L A L F R Q G N G R G S L R  
 1381 TAAAACATTCGAAGAAATTCTAGCACTCTTTCGACAGGGAAATGGCAGGGGCAGTCTTCG  
 481 D S R L Y  
 1441 GGACAGTAGGTTGTATGG

Figure S1 cDNAs and deduced amino acid sequences of Trehalose transporter (Tret 1) (A),  
 Trehalose phosphate synthase (B), Trehalase-1 (C), Trehalase-2 (D), Glycogen polyphosphate (E)  
 and Glucose transporter (Glut 1) (F).

## Materials and methods.

### Insects

The two-spotted crickets *G. bimaculatus* were reared in plastic containers (55×39×31cm) in the temperature conditioned room at 27±1°C, with the long-day photoperiod by 16h:8h as described (Tsukamoto et al., 2014).

### Bibliometric network

The literature data for Chapter 1 were collected from the publications dating from 1990 to 2018 via PubMed (<https://www.ncbi.nlm.nih.gov/pubmed/>). All titles and abstracts were extracted to select the items by the binary counting method over the limitation by the number of 10 as the limitation. Finally, screened keywords for nodes were selected manually to generate the bibliometric mapping.

### RNA-sequencing

Total RNA was extracted using TRI reagent® as described in the commercially available protocol (Molecular Research Center, Inc., Cincinnati, OH, USA). The extracted total RNA was further purified by RNeasy® Mini Kit (QIAGEN, Venlo, Netherlands). RNA sequencing was subjected to HiSeq® 2000 system (Illumina, Inc., San Diego, CA, USA). Contigs were de novo assembled using the software CLC genomic workbench (<https://www.qiagenbioinformatics.com/products/clc-main-workbench/>) and comprehensively annotated using blastx.

### Chemically synthesis of neuropeptides

The mature neuropeptides were manually synthesized according to the Fmoc method C-terminally primed by a Rink-amide resin (Merck Millipore, Darmstadt, Germany). Fmoc amino acids were purchased from Kokusan Chemical (Tokyo, Japan). Fmoc moieties from the chemically extended peptide on the resin were de-protected by incubation with 20% piperidine in dimethylformamide at room temperature for 30 min. Fmoc amino acids were then linked to the first amine group on the synthesizing peptide on the resin in the presence of 1-hydroxybenzotriazole and N, N-dicyclohexylcarbodiimide in

N-methylpyrrolidone. The resulting peptidyl resin was incubated to deprotect all protecting groups and release the peptide from the resin with a cleavage cocktail composed of trifluoroacetic acid (TFA)/ dichloromethane (DCM)/ anisole/ trimethylsilyl bromide (TMSBr)/ 3,4-ethoxylene dioxy thiophene (EDT) of the ration 10/5/2/2/1, v/v/v/v/v on ice for 30 min. The resulting crude peptide was applied to reversed-phase HPLC (Jasco SC-802, PU-880, UV-875; Jasco INT., Tokyo, Japan) on a Senshu Pak PEGASIL-300 ODS column (4.6 mm i.d. ×250 nm; Senshu Kagaku, Tokyo, Japan). The adsorbed peptide was eluted over the program of a linear gradient of 10-60% acetonitrile containing 0.1% TFA over 30 min, at a flow rate of 1 ml/min monitored by absorbance at 225 nm. The highly purified synthetic peptides were confirmed by measurement the molecular mass using MALDI-TOF MS as described below.

#### MALDI-TOF MS analysis

MALDI-TOF MS and MSMS analyses were performed by TOF/TOF 5800 System (AB SCIEX, Framingham, MA, USA). Peptide solutions were subjected to the target of sample stage after mixing with a semi-saturated solution of  $\alpha$ -cyano-4-hydroxycinnamic acid (CHCA; 1:1 v/v) and followingly dried. Amino acid sequencing MSMS analyses were analyzed using the resulting spectra acquired by accumulation of 250–1000 laser shots in total.

#### RNA interference (RNAi)

Double-stranded RNA (dsRNA) was prepared using template plasmid DNA carrying the fragment DNAs obtained from tissue distribution analysis. DsRed-dsRNA was used as an experimental control of dsRNA, corresponding to 14 to 420 nt of DsRed2 nucleotide sequence. The used primers were listed below. RNA was synthesized using T7 RNA polymerase according to manufacturer's protocol using 500 ng of PCR products as template DNA primed by T7 sequence overhanged on the 5' portion of primers. The resulting RNA was purified by phenol/chloroform extraction and ethanol precipitation. The purified RNA was dissolved in diethylpyrocarbonate-treated water to a final concentration of 0.5  $\mu\text{g}/\mu\text{l}$ . dsRNA was generated by gradually cooling the denatured synthesized RNA in 100°C for 5 min and then cooled down to room temperature. Resulting dsRNA was then stored in -20°C until experimental use.

## Expression analysis

cDNAs which were reverse transcribed from total RNA derived from various tissues of adult crickets were prepared as described above. Reverse transcription was primed by an oligo dT18 primer using ReverTra Ace® (Toyobo Co. Ltd., Osaka, Japan). The cDNAs were used as template DNAs for RT-PCR using Gotaq Green Master Mix (Promega Corporation, Madison, WI, USA). The used primers were listed below. PCR products were electrophoresed on a 1.0% agarose gel and visualized by staining by ethidium bromide. DNA bands were extracted and purified using Wizard SV Gel and PCR Clean-up system (Promega Corporation, Madison, WI, USA) and subsequently perform TA-cloning into pGEM-T Vector (Promega Corporation, Madison, WI, USA) according to the manufacturer's protocol. The extraction of the resulting plasmid DNA was primed by the universal M13 primer as listed below, and the inserted cDNAs were sequenced for confirmation using 3500 Genetic Analyzer (Applied Biosystems, Waltham, MA, USA). Real-time quantitative RT-PCR was performed by Thermal Cycler Dice with SYBR-premix Ex Taq-II (TaKaRa, Bio, Japan) with the primers as listed below. The transcriptional levels were normalized to Elongation factor by  $\Delta\Delta C_t$  method to calculate the relative transcriptional level. Primers were designed according to <http://www.primer3plus.com/cgi-bin/dev/primer3plus.cgi>.

## Lipid and carbohydrate extraction from hemolymph

Lipid and carbohydrate were extracted according to Lorenz's method (Lorenz, 2003). Collected 5  $\mu$ l of the crickets' hemolymph from the abdomen was transferred in a tube containing 20 mg of solid sodium sulfate and 200  $\mu$ l of 75% methanol. The mixture was sonicated for 5 min. After adding 600  $\mu$ l of chloroform/methanol (1:1), the sonicated solution was then vortexed and centrifuged (15,000 rpm, 4°C, 10 min). The resulting supernatant was transferred into a new centrifuging tube. Next, 300  $\mu$ l of chloroform/methanol (1:1) was then added. After vortexing and centrifugation (15,000 rpm, 4°C, 10 min), the supernatant was mixed to the former tube containing aqueous layer. The collected aqueous layer was vortexed and centrifuged again (15,000 rpm, 4°C, 10 min) after addition of 500  $\mu$ l chloroform and 300  $\mu$ l 1 M NaCl. The organic lower layer and the aqueous upper layer, which were then dried under vacuum with centrifugation, were used as lipid fraction and carbohydrate fraction for quantification of lipid and trehalose (insect blood sugar), respectively.

### Measurement of lipid

Lipid extracted from cricket hemolymph as described above was quantified according to Sulfo-phospho-vanillin method (Zöllner and Kirsch, 1962). Lipid fractions were dissolved in 40  $\mu\text{l}$  of chloroform/ methanol (1:1). 5  $\mu\text{l}$  of the lipid solution was mixed with 100  $\mu\text{l}$  of sulfuric acid, followed by heated at 100°C for 10 min. After cooling to room temperature, the heated lipid solution was mixed with 500  $\mu\text{l}$  of vanillin reagent (0.2% vanillin in 67% o-phosphoric acid). Absorbance at 595 nm was measured using a spectrophotometer. Quantification of lipids were performed compared with the standard curve generated by Cholesterol (Sigma-Aldrich Japan, Tokyo, Japan); a standard curve of 100, 20, 4, 0.8, 0.16  $\mu\text{g}/\mu\text{l}$  and blank.

### Measurement of carbohydrate

Carbohydrate fraction was dissolved in 100  $\mu\text{l}$  distilled water. 20  $\mu\text{l}$  of the carbohydrate fraction was incubated with 1  $\mu\text{l}$  trehalase (Sigma-Aldrich Japan, Tokyo, Japan) and 5  $\mu\text{l}$  of 0.27 M Citric acid (pH 5.7) at 37°C for 4 hours. After cooling to room temperature, solution is measured by Glucose assay kit (Wako, Tokyo, Japan) according to the manufacture's protocol. The glucose concentration was compared with standard samples 100, 20, 4, 0.8, 0.16  $\mu\text{g}/\mu\text{l}$  of glucose, and then calculated as carbohydrate levels as trehalose level.

### Food intake assay

On third day after adult emergence, 5  $\mu\text{g}$  of dsRNA or 100 pmol of chemically synthesized neuropeptides were injected. These treated crickets were kept isolated in a plastic container. Diet consumption was quantified the difference of diet before and after experiments (24 hours). Before weighing the tablets, the diet tablets were dried by baking at 80 °C for 2 hours. For preparation of dietary tablets, indigestible cellulose was mixed with three different nutrients, which were isocaloric (4 kJ/g). All examined diets contained 33% casein, 33% dextrin and 33% soybean oil for total energy.

### Excretion assay

Excretion activity was measured by counting the number of fecal pellets, which has been used

previously as an index of food intake (Konuma et al., 2012). After treatment of dsRNA and peptides, I counted on the next day of injection. In the experiment of RNAi, I counted the third day after treatment.

#### First-bite time assay

Crickets were kept isolated in a plastic container with water only for 24 hours. At 1 hour after treated with 100 pmol of chemically synthesized neuropeptides, normal diets were replaced. Time were measured until crickets firstly accessed to the food.

#### Crop storage assay

Crickets were dissected for intestine 1 hour after application of diet and injection with 100 pmol chemically synthesized GbCCAP. Width of crop was measured by a scale.

#### RNA probe synthesis

DNA templates were amplified by Ex Taq with 5 ng of vector and gene specific primer with T7 sequence. After purifying PCR products by phenol/chloroform/isopropanol and 70% ethanol, products are incubated with transcription buffer, 0.1 M DTT, BSA, DIG-labeling mixture and T7 polymerase at 37°C for 2 hours. After DNase treatment and purification. The quality of riboprobe was confirmed by electrophoresis with 1.5% TAE agarose gel.

#### *In situ* hybridization

Tissues were dissected in PBS (Wako, Tokyo, Japan) and fixed in 4% paraformaldehyde (PFA)/PBS for 12 hours at 4°C. After washing with PBST (PBS + 0.5% Triton X-100, Invitrogen) (3 times, 10 min each), tissues were digested by Proteinase K (2 µg/ml, Wako) for proper time. After re-fixing with 4% PFA solution and 0.2% glutaraldehyde, tissues are washed with PBST 3 times. RNA probe (0.1 ng/µl) is added to hybridization buffer (1.5% blocking reagent, 50% Formamide, 25% 20xSSC (pH 7.0), 0.1% TritonX-100, 0.1% CHAPS, 1% DNA from salmon sperm) and incubated in a moisture chamber for 48 h at 70°C after a 120 min pre-hybridization. Tissues were then blocked in 1.5% blocking buffer/KTBT (1.5 M NaCl, 0.1 M KCl, 1 M Tris-HCl (pH 7.5)) for 1h at room temperature. After washed by wash

solution (10% 20xSCC (pH=7.0), 0.1% CHAPS) (3 times, 60 min each), the tissues were incubated with secondary antibody (Anti-DIP AP, Fab fragments from sheep, 1:200, Sigma-Aldrich, Tokyo, Japan) for 24 h at 4°C and washed again with KTBT (5 times, 10 min each). CDP-star (Sigma-Aldrich Japan, Tokyo, Japan) is used for staining according to the manufacture's instruction.

#### Immunohistochemistry

Tissues were dissected in PBS and were fixed in 4% paraformaldehyde (PFA)/PBS for 12 hours at 4°C. After washing in PBST (PBS + 0.5% Triton X-100) (3 times, for 10 min each), tissues were blocked in 10% normal goat serum (Wako, Tokyo, Japan) in PBST for 1 h at room temperature, and then incubated with a primary antibody (1:1500) for 48 h at 4°C. After washing with PBST (3 times, for 10 min each), the tissues were incubated with a secondary antibody (Alexa Fluor 488 goat anti-rabbit IgG, 1:1000, Invitrogen, A27034) for 24 hours at 4°C and washed again with PBST (5 times, 10 min each). Control tissues are treated with secondary antibody only. For CCAP immunohistochemistry and MSR in situ hybridization, first antibody of immunohistochemistry is incubated together with secondary antibody in MSR in situ hybridization.

#### Gut contraction assay

After removal of head, crickets on the fifth days after adult emergence are spread out along the abdomen by showing foregut clearly with no harm on peripheral neurons in the 0.9% NaCl. Their foreguts were exposed to 100 pmol synthetic MS for the foregut was conducted observation of gut contraction for 40 s. Experimental control was treated with PBS in the same methods. The contractile movement was observed and recorded using a digital microscope USB2.0 DigiScope II v2TM (CHRONOS, Taiwan). The contractile movements were analyzed using the ImageJ software (<https://imagej.net/Fiji>) with the Optic flow plug-in (Gaussian window MSE). The output results were visualized as a kymograph.

#### Semi-quantification of neuropeptides by MALDI-TOF MS

Extracts of crickets' brains are supersonic treated with 300µl of 60% acetonitrile containing 0.1%

TFA after immersing for 30 minutes, following Zip-Tip treatment for isolation of peptides. Products are semi-quantification measured by MALDI-TOF MS and MSMS with internal standard of *B. mori* peptide GSRYamide-1.

#### Statistical analyses

Statistical analyses were performed using a software, Prism 7 (GraphPad). Each statistical method was described in the figure legends.



### Primer used in this study

*Elongation factor* RT-PCR-f: 5'- ATGCCTGTATCTTGACTGCTCA -3'  
*Elongation factor* RT-PCR-r: 5'- ATGGTTTGCTTCCAGTTCAGT -3'  
*Elongation factor* real time PCR-f: 5'- CCCTGCTGCTGTTGCTTT -3'  
*Elongation factor* real time PCR-r: 5'- CCCATTTTGTTCGGAGTGC -3'  
*DsRed*-f: 5'- GAACGTCATCACCGAGTTCA -3'  
*DsRed*-r: 5'- TGGTCTTCTTCTGCATCACG -3'  
M13 RT-PCR-f: 5'- GTTTTCCAGTCACGACGTT -3'  
M13 RT-PCR-r: 5'- GGAAACAGCTATGACCATGA -3'  
T7-Special-f: 5'-TAATACGACTCACTATAGGGAGACCGCGGAATTCGAT-3'  
T7-Special-r: 5'-TAATAGACTCACTATAGGGAGAGAATTCAGTAGTAT-3'  
*GbAKH* RT-PCR-f: 5'- TCCATGGAGGAATTGCACCC -3'  
*GbAKH* RT-PCR-r: 5'- CTTGGTTCTGGTTTTGGCGG -3'  
*GbAKH* real time PCR-f: 5'- CGCCAAAACCAGAACCAAGG -3'  
*GbAKH* real time PCR-r: 5'- TGGAGTTCAGACGTGTGCTC -3'  
*GbACP* RT-PCR-f: 5'- TCCGCAACTCATGGCTTCCT -3'  
*GbACP* RT-PCR-r: 5'- TGGAGAAGGTGATCTGCGCG -3'  
*GbACP* real time PCR-f: 5'- CCACCCATCTGCGGTAGAAG -3'  
*GbACP* real time PCR-r: 5'- CCATGAGTTGCGGAGTAGCT -3'  
*GbCRZ* RT-PCR-f: 5'- CGAAGGATCTCCACAGACG -3'  
*GbCRZ* RT-PCR-r: 5'- TGAGGAATACTCGTCGCTGC -3'  
*GbCRZ* real time PCR-f: 5'- GGTGCGTGCTCCTGCTGAGT -3'  
*GbCRZ* real time PCR-r: 5'- CCGCGGCTGTACTGGAAGGT -3'  
*GbCCAP* RT-PCR-f: 5'- CCACAAGAGGCAAGCGGAC -3'  
*GbCCAP* RT-PCR-r: 5'- CCGGTGAAGGCGTTGCAGAA -3'  
*GbCCAP* real time PCR-f: 5'- CCACAAGAGGCAAGCGGAC -3'  
*GbCCAP* real time PCR-r: 5'- CCGGTGAAGGCGTTGCAGAA -3'  
*GbMS* RT-PCR-f: 5'- CCCGTCATCAAGAGGCAAGA-3'  
*GbMS* RT-PCR-r: 5'- ACATGCTTTGTACAGGGCCG -3'  
*GbMS* real time PCR-f: 5'- AGAGGCAAGACGTGATCAC-3'  
*GbMS* real time PCR-r: 5'- ACATGCTTTGTACAGGGCCG -3'  
*GbAst-A* RT-PCR-f: 5'- AAGGCCGCATGTACTCCTTC -3'  
*GbAst-A* RT-PCR-r: 5'- GGTCCCCATGTACGACTTCG -3'  
*GbAst-A* real time PCR-f: 5'- TCCGAGTACAAGCGCCTGCC -3'  
*GbAst-A* real time PCR-r: 5'- GAAGCTGTACATGCCGGCGC -3'  
*GbMIP* RT-PCR-f: 5'-GGGGTGGGCAGTTACTCTTC -3'  
*GbMIP* RT-PCR-r: 5'- TCGTACTGGCCCTCGATCTC -3'  
*GbMIP* real time PCR-f: 5'- ACGTGGCCGGTTCCTCCTTC -3'  
*GbMIP* real time PCR-r: 5'- TGAGCGCTGTGTGGGGGAAG -3'  
*GbAst-C* RT-PCR-f: 5'- CAGGAGACGGCGTTGAAGG -3'  
*GbAst-C* RT-PCR-r: 5'- CTACCATCGTCGTCCACCAG -3'  
*GbAst-C* real time PCR-f: 5'- CAGTAGCTGCGCTTGCCTG -3'

*GbAst-C* real time PCR-r: 5'- CGTCGAGACCGCGCTCATCA-3'  
*GbAT* RT-PCR-f: 5'- AGCAAGCCCCGCACGATACG -3'  
*GbAT* RT-PCR-r: 5'- AGGGGCTCAGTACAGTGGCCT -3'  
*GbAT* real time PCR-f: 5'- TTCGCCAGCAGCAGACTT -3'  
*GbAT* real time PCR-r: 5'- ACAGCTCGCCGTCCTGGTTC -3'  
*GbsNPF* RT-PCR-f: 5'- CGCTGTCCCTGATGCTGCTACTC-3'  
*GbsNPF* RT-PCR-r: 5'- CCTCACAATAAATGGCGGTGCTT -3'  
*GbsNPF* real time PCR-f: 5'- TTTCCGGAATTGCTTTGCCG-3'  
*GbsNPF* real time PCR-r: 5'- CGCTTGCGAACATACTCAGC -3'  
*GbFMRFa* RT-PCR-f: 5'- TCGGAAGGGCCAACCA -3'  
*GbFMRFa* RT-PCR-r: 5'- TCCAGCGACACCTGTGC -3'  
*GbFMRFa* real time PCR-f: 5'- CGCGCGGGAGAGGAAATGGA -3'  
*GbFMRFa* real time PCR-r: 5'- AAGAAGTTGGATGCGGCGCG -3'  
*GbDH31* RT-PCR-f: 5'- CGTCGTTGGGCGTGATGAGTGA -3'  
*GbDH31* RT-PCR-r: 5'- CTAGCGGCGAGGGAGGAAGAA -3'  
*GbDH31* real time PCR-f: 5'- GCTGGAGATGCTGGCACGGA -3'  
*GbDH31* real time PCR-r: 5'- GGGCGCGCTTGTCTCAACG -3'  
*GbDH44* RT-PCR-f: 5'- GCAATCGCATCCAGCAGAACC -3'  
*GbDH44* RT-PCR-r: 5'- AAGGGAAACAGGGTGACAGAACAGA -3'  
*GbDH44* real time PCR-f: 5'- AGTCGCTGTCCATCGTGGCG -3'  
*GbDH44* real time PCR-r: 5'- GCTGGATGCGATTGCCTTGC -3'  
*GbAst-A* receptor RT-PCR-f: 5'- AGATCACGCAGAAGGTGGTG -3'  
*GbAst-A* receptor RT-PCR-r: 5'- CGGATGGACATCGACGTGAT -3'  
*GbAst-A* receptor real time PCR-f: 5'- TTTTGCGTGCCGTTACCCGC -3'  
*GbAst-A* receptor real time PCR-r: 5'- TTGCACCAGACGTCCCCGAA -3'  
*GbBur-a* receptor RT-PCR-f: 5'- CAAGCTGTTCCCTGTCCGACT -3'  
*GbBur-a* receptor RT-PCR-r: 5'- CGTTGTTGCCAGGTTCAAG -3'  
*GbBur-a* receptor real time PCR-f: 5'- ACCGCATTGGCAGCCTGGAA -3'  
*GbBur-a* receptor real time PCR-r: 5'- TGCTCTGGCAGGTGCGTGAT -3'  
*GbBur-β* receptor RT-PCR-f: 5'- CTCCGACCAGTACGCTTACC -3'  
*GbBur-β* receptor RT-PCR-r: 5'- GAGCTCGACAGGTTGGAGTC -3'  
*GbBur-β* receptor real time PCR-f: 5'- AGCAGAAGCAGCACAAGCGC -3'  
*GbBur-β* receptor real time PCR-r: 5'- AGCGCGACGAGCTGAAGTTCT -3'  
*GbCCHa-2* receptor RT-PCR-f: 5'- GCTGTGCATCATCGCGTGCT -3'  
*GbCCHa-2* receptor RT-PCR-r: 5'- CACCTTTTTGCGGGCGCGAA -3'  
*GbCCHa-2* receptor real time PCR-f: 5'- GCTGTGCATCATCGCGTGCT -3'  
*GbCCHa-2* receptor real time PCR-r: 5'- CACCTTTTTGCGGGCGCGAA -3'  
*GbMIP* receptor RT-PCR-f: 5'- ACTTCTTCGAGCAGTGCTACT -3'  
*GbMIP* receptor RT-PCR-r: 5'- TGACCTGCGTCACGTGGTT -3'  
*GbMIP* receptor real time PCR-f: 5'- AATGTTTCGGCCGCGCTTCC -3'  
*GbMIP* receptor real time PCR-r: 5'- ACCTGCGTCACGTGGTTGGT -3'  
*GbNPF* receptor RT-PCR-f: 5'- CAGTACGTGGTGCCCTTCAT -3'  
*GbNPF* receptor RT-PCR-r: 5'- CTGGAACCTCTTGCGGAAGT -3'

*GbNPF* receptor real time PCR-f: 5'- TACTCGCGCATCTGCCGCAA -3'  
*GbNPF* receptor real time PCR-r: 5'- TTCGTCTTGCGCCGAGTGGA -3'  
*GbRYa* receptor RT-PCR-f: 5'- CTGTGCTACGGCTCCATCTC -3'  
*GbRYa* receptor RT-PCR-r: 5'- GAAGATGCTCACGTTACGC -3'  
*GbRYa* receptor real time PCR-f: 5'- GCGCATGCAGAACGTCACCA -3'  
*GbRYa* receptor real time PCR-r: 5'- AAGGTTCCAGCGCTGCAGCA -3'  
*GbFMRFa* receptor RT-PCR-f: 5'- CGCCTCATGCTCTCCATCTT -3'  
*GbFMRFa* receptor RT-PCR-r: 5'- AGATGAAGGGGTTGACGACG -3'  
*GbFMRFa* receptor real time PCR-f: 5'- TGCTTCTGCCGCTCATGCT -3'  
*GbFMRFa* receptor real time PCR-r: 5'- AGATGAAGGGGTTGACGACGGC -3'  
*GbGlyco-A2* receptor RT-PCR-f: 5'- TGCTGGTAGCGAGTGCATTGTTG -3'  
*GbGlyco-A2* receptor RT-PCR-r: 5'- GGGTTGATACGCAGCGTGTGG -3'  
*GbGlyco-A2* receptor real time PCR-f: 5'- ACTCGCACAGTCAGCATACC -3'  
*GbGlyco-A2* receptor real time PCR-r: 5'- CCAGGATTCGCAGTAGCCTC -3'  
*GbGlyco-B5* receptor RT-PCR-f: 5'- CTACAAGGTGGTGAAGGCGGACTC -3'  
*GbGlyco-B5* receptor RT-PCR-r: 5'- TGTTTGGGAAGCGCCAGTCC -3'  
*GbGlyco-B5* receptor real time PCR-f: 5'- CTACAAGGTGGTGAAGGCGGACTC -3'  
*GbGlyco-B5* receptor real time PCR-r: 5'- TGTTTGGGAAGCGCCAGTCC -3'  
*GbInotocin* receptor RT-PCR-f: 5'- CCAATGAGGAATTGCGCGCG -3'  
*GbInotocin* receptor RT-PCR-r: 5'- TCTGTGAGGTGGCAAG-3'  
*GbInotocin* receptor real time PCR-f: 5'- TGTCTTGCCACACCGACCCC -3'  
*GbInotocin* receptor real time PCR-r: 5'- AGGTCCGAGGGTCCATCCGA -3'  
*GbOrcokinin* receptor RT-PCR-f: 5'- CGTGCCTCTACACCGATTCTCA -3'  
*GbOrcokinin* receptor RT-PCR-r: 5'- TATCTCGACGTTGGCTCAACTCATG -3'  
*GbOrcokinin* receptor real time PCR-f: 5'- ACCGGCTTCGACAACCTCAT -3'  
*GbOrcokinin* receptor real time PCR-r: 5'- CGTTGACGCCGACGAGGT -3'  
*GbPDF* receptor RT-PCR-f: 5'- CTGCTCGACAAGGAGGTAGC -3'  
*GbPDF* receptor RT-PCR-r: 5'- GCCCTGCATCATTACAGCAC-3'  
*GbPDF* receptor real time PCR-f: 5'- GAAGCCAATGCTGCTCCTTCTCC -3'  
*GbPDF* receptor real time PCR-r: 5'- GTGAGTTGATGATTCCGAGTTCCCTT -3'  
*GbCCAPR-1* RT-PCR-f: 5'- GGGAGGCCCGACTTGTATTC -3'  
*GbCCAPR-1* RT-PCR-r: 5'- TTGAAAGTGACGTTCTGCGC -3'  
*GbCCAPR-1* real time PCR-f: 5'- GCAAGTGGTGTGGATATTC -3'  
*GbCCAPR-1* real time PCR-r: 5'- ACCTCCAGTCAGTCATTCT -3'  
*GbCCAPR-2* RT-PCR-f: 5'- CGCGCGCCAAGATCAAGACGGT -3'  
*GbCCAPR-2* RT-PCR-r: 5'- AACGCAGCGCCGGGAAGCACA -3'  
*GbCCAPR-2* real time PCR-f: 5'- TCGGAAATCATCAACTCACTCTTAGGTC -3'  
*GbCCAPR-2* real time PCR-r: 5'- CCACGGCGATGTTGGTCTGC -3'  
*GbMSR* RT-PCR-f: 5'- CCACCAACTCCATCCTCACC -3'  
*GbMSR* RT-PCR-r: 5'- GTTGTGGTCGCTCTTGTG -3'  
*GbMSR* real time PCR-f: 5'- TGCTCAGCCCCTGCTCT -3'  
*GbMSR* real time PCR-r: 5'- TGTGGTCGCTCTTGTGTAGTCG -3'  
*GbAKHR* RT-PCR-f: 5'- ATGGACAGGTAATTCGCCATTTT -3'

*GbAKHR* RT-PCR-r: 5'- ATGAAGAAGAACAGCACGATGAC -3'  
*GbAKHR* RNAi-f: 5'- CCATTCCTGTCCCAGCCATT -3'  
*GbAKHR* RNAi-r: 5'- AACTCGTGCATGAACCCGAT -3'  
*GbAKHR* real time PCR-f: 5'- TCTGGGTACTCTCGCAGCGC -3'  
*GbAKHR* real time PCR-r: 5'- CGTGTACGGCGCGTTCAACG -3'  
*GbIRS* RT-PCR-f: 5'- CTACATGGAAGTGGGCAGGT -3'  
*GbIRS* RT-PCR-r: 5'- GGGGTAGAAGGTGACATGGA -3'  
*GbIRS* real time PCR-f: 5'- ACTTGCTCTGCCTTACGGAC -3'  
*GbIRS* real time PCR-r: 5'- CTGTTGGCCTTCTCCTCCTG -3'  
T7-*GbIRS*-f: 5'- GCTTCTAATACGACTCACTATAGCTACATGGAAGTGGGCAGGT -3'  
T7-*GbIRS*-r: 5'- GCTTCTAATACGACTCACTATAGGGGTAGAAGGTGACATGGA -3'  
*GbMSR*-probe-f: 5'- GCCTGCTTCGTGCTGTTCC -3'  
*GbMSR*-probe-r: 5'- TCCGTCTGGCGGCTCTTG -3'  
T7-*GbMSR*-probe-f: 5'- TAATACGACTCACTATAGGGAGAGCCTGCTTCGTGCTGTTCC -3'  
T7-*GbMSR*-probe-r: 5'- TAATACGACTCACTATAGGGAGATCCGTCTGGCGGCTCTTG -3'  
*GbTret1* RT-PCR-f: 5'- GCGGCGAAAGACTAAAGACG -3'  
*GbTret1* RT-PCR-r: 5'- TGCAGGTTGATGCTGGTGA -3'  
*GbTret1* real time PCR-f: 5'- AGACTACCATCCTCGCCACC -3'  
*GbTret1* real time PCR-r: 5'- TGACGCCACGCAGAAG -3'  
*GbGlut1* RT-PCR-f: 5'- GTCGGAGATCACTGCACCTGTAC -3'  
*GbGlut1* RT-PCR-r: 5'- CCCCTGCCATTTCCCTGT -3'  
*GbGlut1* real time PCR-f: 5'- CGCTACGGAGGTTGCGTGCTT-3'  
*GbGlut1* real time PCR-r: 5'- TTGCTGGGCTCTTTCTTCTGCTC-3'  
*GbGp* RT-PCR-f: 5'- GATGGCTATCAACAACATTGCTTC -3'  
*GbGp* RT-PCR-r: 5'- TTGTCTACGGCACTTCTGTTCA -3'  
*GbGp* real time PCR-f: 5'- TCTTCGGAGAATGTCCCTTGTGG -3'  
*GbGp* real time PCR-r: 5'- CTCCATTGACTGCATGAGAACCC-3'  
*GbTPS* RT-PCR-f: 5'- AGGCTAGGCTGCCGAGTG -3'  
*GbTPS* RT-PCR-r: 5'- GCCCATTGATGCGTCCC -3'  
*GbTPS* real time PCR-f: 5'- TGCGGATTGTGGGTGGG -3'  
*GbTPS* real time PCR-r: 5'- TGATTCAGGTATGGGCTCATTGG -3'  
*GbTPS* RT-PCR-f2: 5'- AGGCTAGGCTGCCGAGTG -3'  
*GbTPS* RT-PCR-r2: 5'- GCCCATTGATGCGTCCC -3'  
*GbTPS* real time PCR-f2: 5'- CGGCGTGCTCATCGTGTCG -3'  
*GbTPS* real time PCR-r2: 5'- GGCGTTGATCTCGTAGGGGTTG -3'  
*GbTre1* RT-PCR-f: 5'- TCTGCTTAGCGATATGAAGGACAC -3'  
*GbTre1* RT-PCR-r: 5'- CATCGACTGGAGAAATCCCAAC -3'  
*GbTre1* real time PCR-f: 5'- CGTATCGCCGACCCCAAGCT -3'  
*GbTre1* real time PCR-r: 5'- CCTGACTTCGTGCGAAACCCTTC -3'  
*GbTre2* RT-PCR-f: 5'- CTCCTCCGTGCGAAAGTGAA-3'  
*GbTre2* RT-PCR-r: 5'- CAGGATGAGGCACATAGATAATGG -3'  
*GbTre2* real time PCR-f: 5'- ATGGACTGGCTGCTATGATACAAC -3'  
*GbTre2* real time PCR-r: 5'- TCCAGGCTTTCCACAAACAGAT -3'

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