

**Preliminary Report
of
The Hakuho Maru Cruise
KH-09-5 Leg 5**

Jan. 29, 2009 - Feb. 10, 2009
(Eel Cruise Indian Ocean II)

Atmosphere and Ocean Research Institute
The University of Tokyo
2012

**Preliminary Report
of
The Hakuho Maru Cruise
KH-09-5 Leg 5
(EEL Indian Ocean II)**

Jan. 29, 2009 - Feb. 10, 2009

Atmosphere and Ocean Research Institute
The University of Tokyo
2012

By
The Scientific Members of the Expeditions

Edited by
Shun Watanabe, Kazuki Yokouchi, Jun Aoyama,
Tsuguo Otake and Katsumi Tsukamoto

Contents

01 Preface	1
02 List of participants	2
03 Track chart	3
04 Anguillid Leptocephali Collected during Leg 5 of KH-09-5	5
	All Scientists onboard	
05 Migration pathways of Anguilliform Leptocephali in the Southwest Indian Ocean	9
	Raymonde LECOMTE-FINIGER, Eric FEUNTEUN, Mari KUOKI, Michael J. MILLER, Elodie REVEILLC, Katsumi TSUKAMOTO and Tsuguo OTAKE	
06 Leptocephali Collected west of the Nazareth Bank of the Mascarene Ridge during Leg 5 of KH-09-5	13
	All Scientists onboard	
07 Taxonomical Identification of Muraenid Leptocephali Collected during the KH-09-5 Leg 5 Cruise using Morphology and DNA Barcoding	17
	Atsushi TAWA	
08 Leptocephalus food web	19
	Alexandre CARPENTIER, Christine DUPUY, Celine ELLIEN, Eric FEUNTEUN, Mari KUROKI, Raymonde LECOMTE-FINIGER, Michael MILLER, Tsuguo OTAKE, Stéphane POUS, Elodie REVEILLAC, Tony ROBINET, and Katsumi TSUKAMOTO	
09 Gobiidae larvae dispersion	30
	Celine ELLIEN, Shun WATANABE, Stephane POUS, and Katsumi TSUKAMOTO	

10	Report on the Parasitofauna of Mesopelagic Fishes	33
	Betty FALIEX and Elsa AMILHAT	
11	Diversity of fish larvae west of the Mascarene Ridge in the Southwestern Indian Ocean	35
	Tony ROBINET, Mari KUROKI, Alex CARPENTIER, Michael J. MILLER, Katsumi TSUKAMOTO and Tsuguo OTAKE	
12	Larval distribution and population structure of Thunnus in the Indian Ocean	38
	Takashi KITAGAWA	
13	Studies on feeding ecology of phyllosoma larvae of palinurid and scyllarid lobsters	39
	Masamichi MACHIDA and Shuhei NISHIDA	
14	Geographic Variation in the Assemblage Structure and Life Histories of Leptocephali across the Indian Ocean Basin	41
	Michael J. MILLER, Jun AOYAMA, Shun WATANABE, Eric FEUNTEUN, Sam WOUTHUYZEN, Augy SYAHAILATUA, Tsuguo OTAKE and Katsumi Tsukamoto	
16	Net records	42
17	CTD data	45
18	Working log	62

Preface

The KH-09-5 (Leg 5) research cruise was the second expedition of the R/V Hakuho Maru that focused on the search for the spawning areas of freshwater eels (genus *Anguilla*) that live in the western side of the Indian Ocean since the historical Carlsberg Foundation's Oceanographic Expedition Around the World from 1928-1930 by the Danish scientist Johannes Schmidt. During the 2010 cruise we also aimed to understand 1) Anguilliformes leptocephalus distribution and feeding ecology, 2) distribution and migration of amphidromous Sicydiinae gobies, and 3) the biological diversity in the western Indian Ocean near the Mascarene Ridge. A total of 28 scientists from France, Madagascar, USA and Japan, and one observation scientist from Mauritius participated in this cruise. This cruise represented actual international collaboration for studies on eels and other organisms in the western Indian Ocean.

During the cruise two *Anguilla bicolor bicolor* and a total of 579 Anguilliformes leptocephali were collected by 40 tows of the IKMT. A large number of fish larvae including a few Sicydiinae gobies were also collected. CTD observations at many of the 40 stations of IKMT towing will advance the understanding of details about the distribution and migration of fish larvae as well as eel leptocephali in relation to the oceanographic structure and currents of this interesting area. The analyses of water samples taken from several depth strata at 15 stations will provide vital information for revealing the food sources of leptocephali. The two *A. bicolor bicolor* leptocephali collected during this cruise are not enough to determine the spawning area. However, all the specimens of other leptocephali and the oceanographic data will certainly give us new insights about fish biology and ecology in the Indian Ocean.

An onboard seminar was held almost every day and a total of 20 titles were presented. A variety of interesting topics on eel biology and marine ecosystems presented by French, Madagascar, Mauritius and Japanese scientists were useful for understanding the research activities of each other. Several seeds of collaborative study on eel biology between French and Japanese scientists also came out during this cruise, which are really expected to develop into fruitful research progress in the future.

Since scientists of five different nations participated in this cruise, the official language used in the meeting and the seminar was English. Communication was therefore sometimes a little difficult, but never affected the scientific routine work. There was always a general understanding among all cruise members and an enjoyable atmosphere. There was no time loss caused by bad weather during this cruise and every observation scheduled was completed without any trouble. On behalf of all scientists aboard, I would like to express our sincere thanks to the Captain, the officers and the crew members of the R/V Hakuho Maru for their support during this cruise.

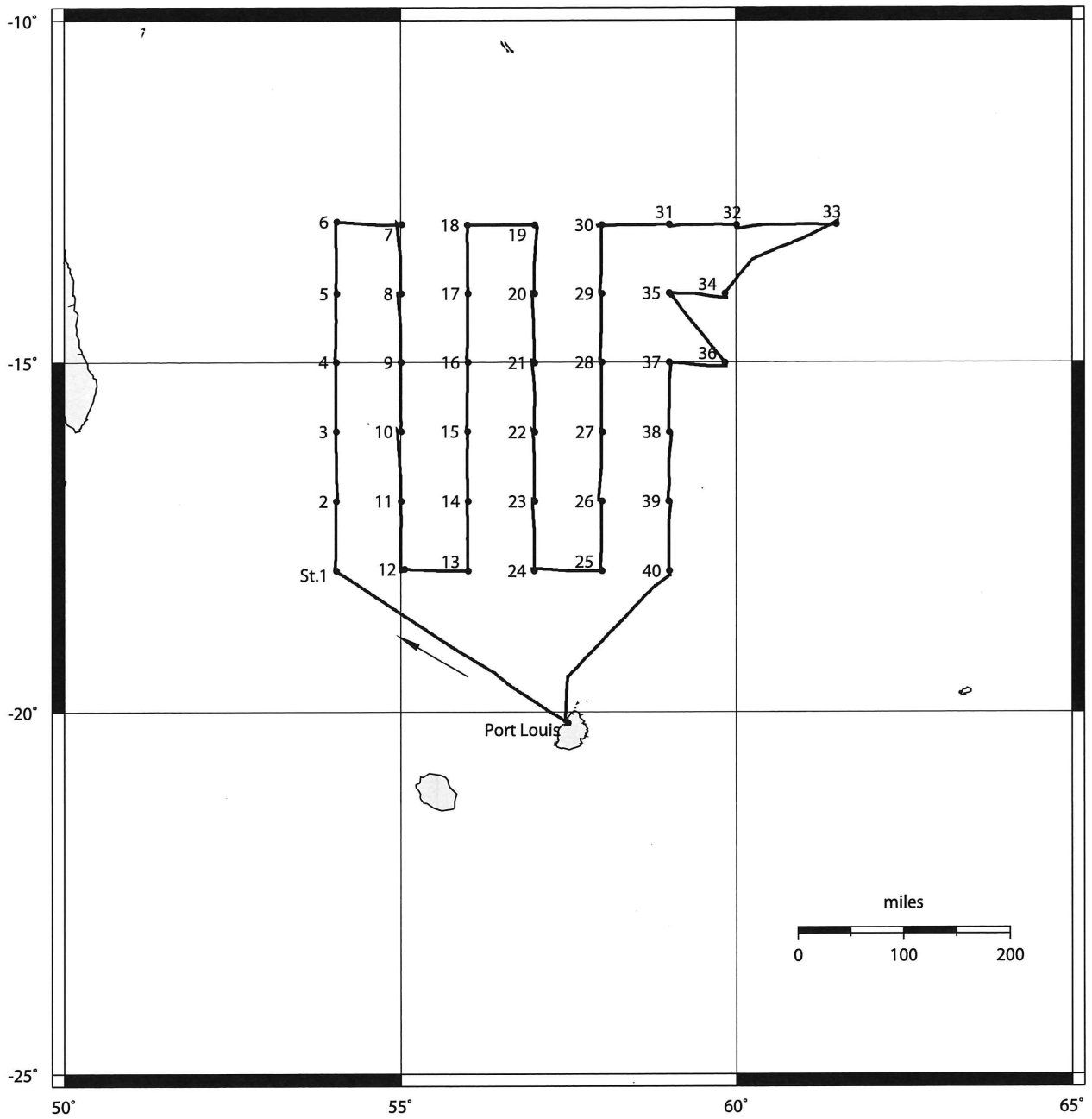
09 February 2010
Chief Scientist: Tsuguo Otake

Scientists on board HAKUHO-MARU (KH-09-5 Leg 5)

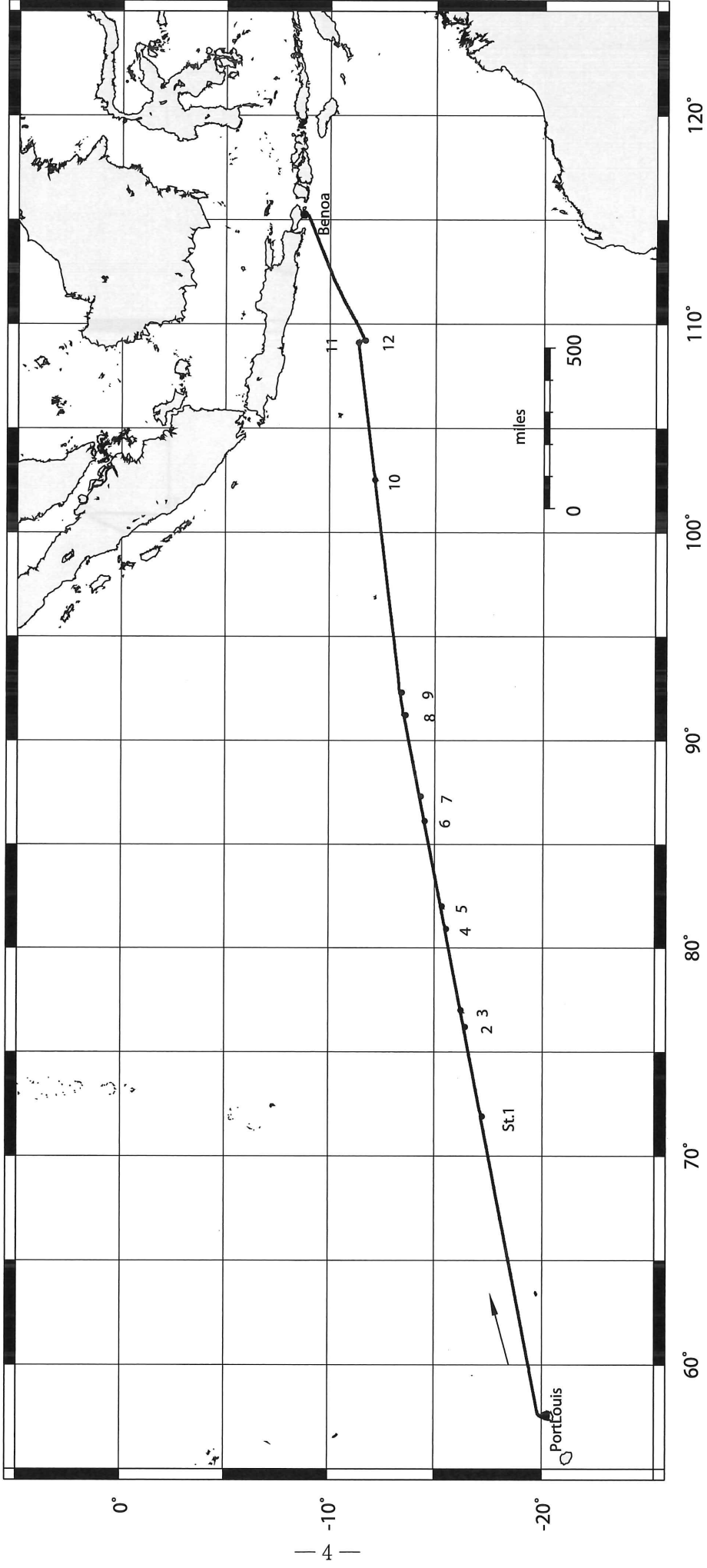
Name	Affiliation
OTAKE Tsuguo	Ocean Research Institute, The University of Tokyo
TSUKAMOTO Katsumi*	Ocean Research Institute, The University of Tokyo
AOYAMA Jun* ^o	Ocean Research Institute, The University of Tokyo
KITAGAWA Takashi*	Ocean Research Institute, The University of Tokyo
MILLER Michael J.*	Ocean Research Institute, The University of Tokyo
WATANABE Shun*	Ocean Research Institute, The University of Tokyo
OGUMA Kenji*	Ocean Research Institute, The University of Tokyo
KAMEO Katsura	Ocean Research Institute, The University of Tokyo
HAGIHARA Seishi*	Ocean Research Institute, The University of Tokyo
MANABE Ryotaro	Ocean Research Institute, The University of Tokyo
AMANO Yosuke	Ocean Research Institute, The University of Tokyo
HATA Masayoshi	Ocean Research Institute, The University of Tokyo
MACHIDA Masamichi*	Ocean Research Institute, The University of Tokyo
KUROKI Mari	The University Museum, The University of Tokyo
TAWA Atsushi*	Graduate School of Bioresource and Bioenvironmental Science, Kyushu University
AI Bunpei*	NPO Nihon Sakana no Kai
TAKAMORI Tomoyuki*	Marine Work Japan LTD
FEUNTEUN Eric	Museum National d'Histoire Naturelle, France
ROBINET Tony	Museum National d'Histoire Naturelle, France
REVEILLAC Elodie	Museum National d'Histoire Naturelle, France
Roger	Museum National d'Histoire Naturelle, France
POUS Stephan	Museum National d'Histoire Naturelle, France
ELLIEN Celine	Universite Paris 6, France
LECOMTE-FINIGER Raymonde	EPHE-CNRS Universite de Perpignan, France
AMILHAT Elsa	CNRS-Universite de Perpignan, France
FALIEUX Elisabeth	CNRS-Universite de Perpignan, France
DUPY Christine	Universite La Rochelle
CARPENTIER Alexandre	URU 420 Universite de Rennes 1
MUSSAI Prakash	Mauritius Oceanography Institute

*Onboard Leg6; ^oChief Scientist of Leg 6.

KH-09-5_Leg5



KH-09-5_Leg6



Anguillid Leptocephali Collected during Leg 5 of KH-09-5

All Scientists onboard

Anguillid eels inhabit the continental waters on both sides of the Indian Ocean basin and they also live in the rivers and streams of Madagascar and the other smaller islands of the western Indian Ocean (Robinet et al. 2003, 2008). However, except for a general spawning region in the eastern Indian Ocean indicated by collections of various sizes of leptocephali (Jespersen 1942; Aoyama et al. 2007), where the 5 or 6 species or subspecies of Indian Ocean anguillid eels go to spawn is unknown.

Therefore the collaborative cruise between scientists from France and Japan had the objective to learn more about the spawning areas of anguillid eels in the western Indian Ocean. However, due to constraints of limited ship time being available, and concerns of potential pirate activity north of 13°S, the sampling survey of Leg 5 of KH-09-5 was concentrated in a specific area to the west of the southern Mascarene Ridge (Fig. 1). Sampling was conducted at 40 stations using an Isaacs Kidd Midwater trawl (IKMT) with an 8.7 m² mouth opening and 0.5 mm mesh during both day and night. Step tows that sampled 5 different layers were conducted at night (120, 90, 70, 50, 30 m for 10 min at each depth layer), and oblique tows to a depth of about 500 m were made during the day.

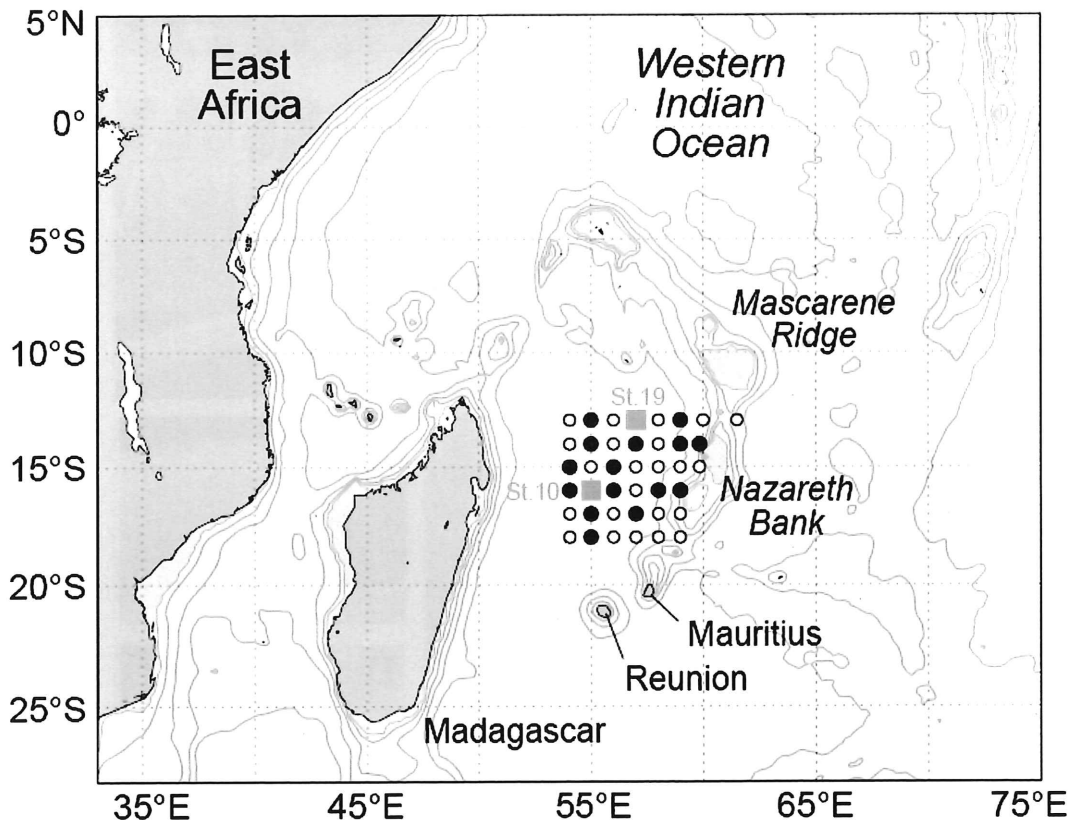


Figure 1. Map showing the stations where *Anguilla bicolor bicolor* leptocephali were collected (red squares) to the west of the Mascarene Ridge during Leg 5 of the KH-09-5 cruise. Black circles show night stations, white circles show day stations.

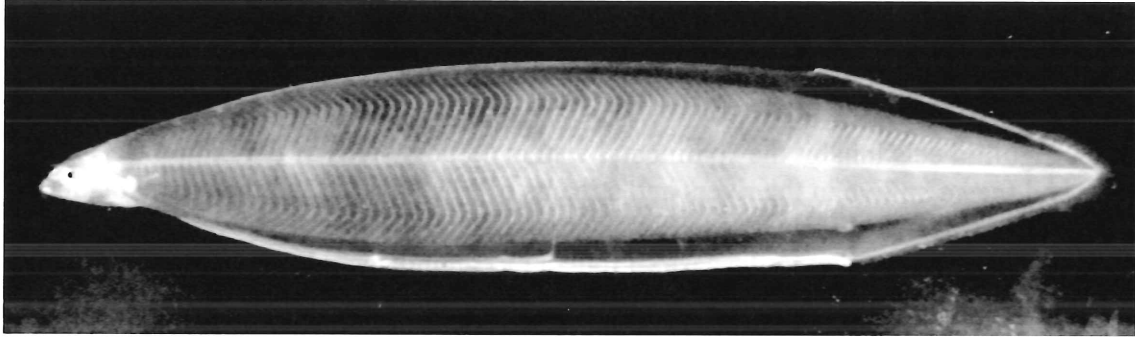


Figure 2. Photograph of a 54.1 mm TL *Anguilla bicolor bicolor* leptocephalus collected at Stn. 10 to the west of the Mascarene Ridge on 1 February 2010 during Leg 5 of the KH-09-5 cruise.

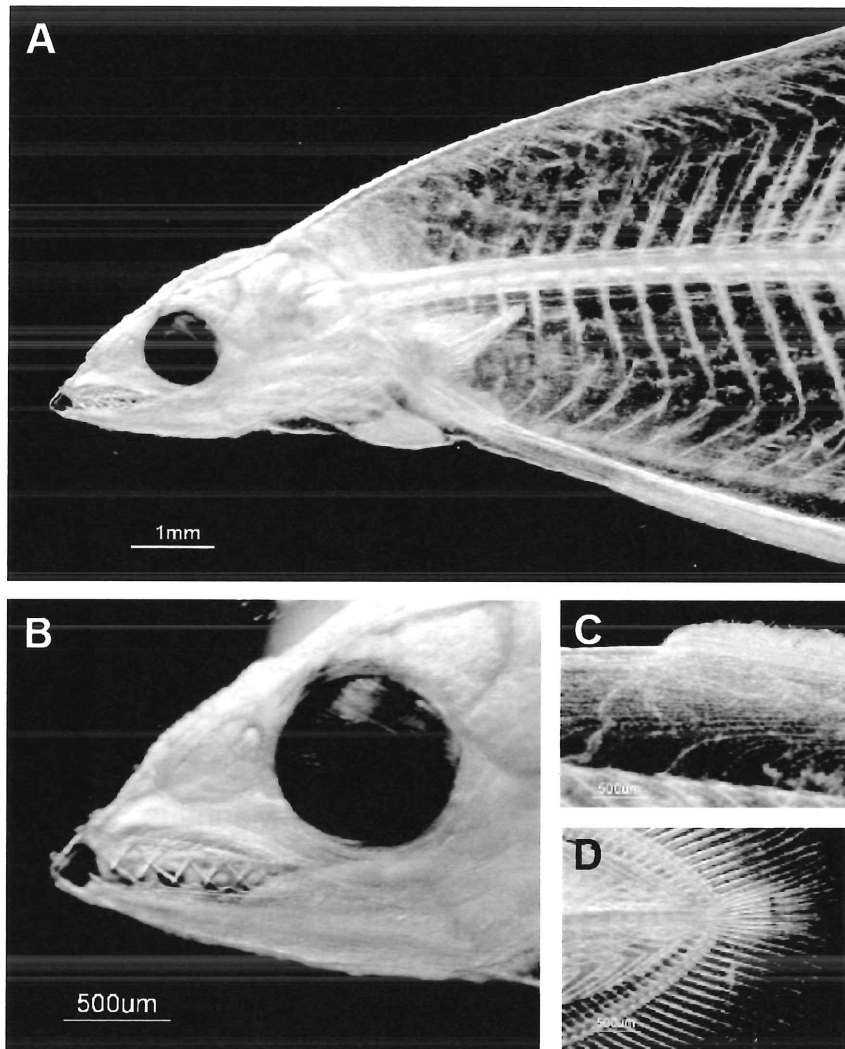


Figure 3. Photographs of a 54.1 mm TL *Anguilla bicolor bicolor* leptocephalus collected at Stn. 10 during Leg 5 of the KH-09-5 cruise showing the detailed morphology of the head region (A, B), the anterior edge of the dorsal fin (C), and the caudal fin region (D).

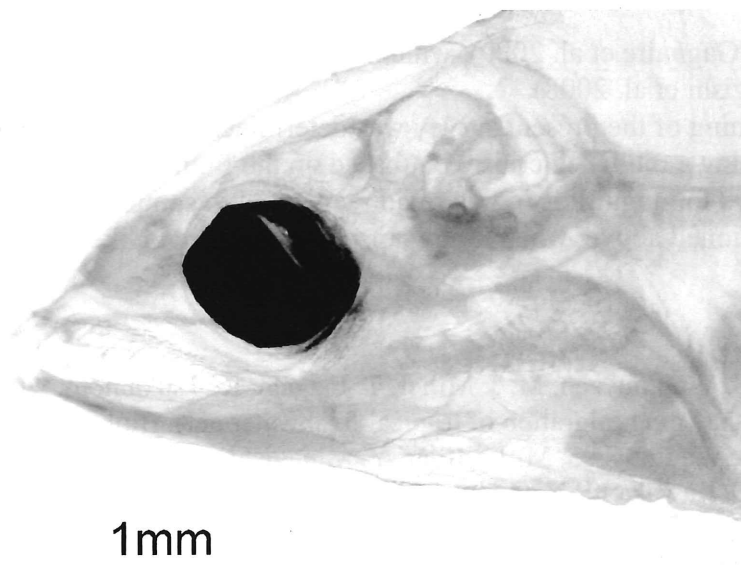


Figure 4. Head region of a 51.2 mm TL *Anguilla bicolor bicolor* leptocephalus collected at Stn. 19 on 3 February 2010 during Leg 5 of the KH-09-5 cruise showing the head features including the early gills, two otoliths, olfactory organ, and heart.

During this sampling effort, despite 579 marine eel leptocephali being collected (see leptocephali report), only 2 anguillid leptocephali were collected, indicating that anguillid leptocephali were only present in low densities in this area during this particular season. The anodorsal myomere count of these larvae (ADM: number of myomeres from the start of the dorsal fin to the start of the anal fin), and the distinctive slender body shape (Fig. 2), indicated that these larvae could be identified as *Anguilla bicolor bicolor*. The first specimen was collected at Stn. 10 (Fig. 1), was 54.1 mm TL, with 109 total myomeres (TM), and had an ADM count of 5, which is well below the separating point between shortfin and longfin leptocephali of about 8 or 9 (Jespersen 1942). The second specimen caught at Stn. 19 was 51.2 mm TL, with 110 TM, and had an ADM count of about 7 or 8. Since *A. bicolor bicolor* is the only shortfin species in this region, these specimens could be identified morphologically. This will be confirmed later using molecular genetic techniques (e.g. Aoyama et al. 2003, 2007; Kuroki et al. 2006, 2007, 2009; Gagnaire et al. 2007).

The low density and large size of the anguillid leptocephali during this survey indicate that this time of year is at least several months after much spawning activity had taken place near the sampling area. Anguillid eels of several species including *A. bicolor bicolor*, *A. mossambica*, and *A. marmorata* are known to recruit to the various islands of the region including Mauritius, Reunion, the Seychelles, and Madagascar (Robinet et al. 2003, 2007, 2008; Réveillac et al. 2008, 2009). Therefore to recruit to some of these areas after spawning in the westward flowing South Equatorial Current (SEC), at least some of the leptocephali must likely pass through the sampling grid of Leg 5 of KH-09-5. Based on this, and the similar result obtained in 2006 during late Nov. and early Dec., the primary spawning season of anguillids in this region can be inferred to sometime within the 6 month period of about Mar. to Sept..

Analyses of the otolith microstructure of the *A. bicolor bicolor* leptocephali collected during this survey and those from the 2006 survey can determine the spawning time of the larvae. Their age can also help evaluate their possible geographic area of origin, since the SEC has the potential to transport leptocephali long distances across the Indian Ocean basin, as has been recently suggested for *A.*

marmorata (Gagnaire et al. 2009), which has a metapopulation structure within the basin (Minegishi et al. 2008).

The timing of the present survey was determined by the many factors associated with scheduling a multi-leg cruise, but based on the findings of the present study, future surveys for anguillids in this region will need to sample at a specific time of year as mentioned above, when it now appears spawning is likely to occur.

References

- Aoyama, J., S. Wouthuyzen, M. J. Miller, T. Inagaki, K. Tsukamoto. 2003. Short-distance spawning migration of tropical freshwater eels. *Biol. Bull.* 204: 104–108.
- Aoyama J., S. Wouthuyzen, M. J. Miller, Y. Minegishi, G. Minagawa, M. Kuroki, S. R. Suharti, T. Kawakami, K. O. Sumardiharga, K. Tsukamoto. 2007. Distribution of leptocephali of the freshwater eels, genus *Anguilla*, in the waters off west Sumatra in the Indian Ocean. *Envir. Biol. Fish.* 80: 445–452.
- Gagnaire, P. A., K. Tsukamoto, J. Aoyama, Y. Minegishi, P. Valade, P. Berrebi, 2007. RFLP and semi-multiplex PCR-based identification of four eel species from the south-western Indian Ocean region. *J. Fish Biol.* 71: 279–287.
- Gagnaire, P. A., Y. Minegishi, J. Aoyama, E. Réveillac, T. Robinet, P. Bosc, K. Tsukamoto, E. Feunteun, P. Berrebi. 2009. Ocean currents drive secondary contact between *Anguilla marmorata* populations in the Indian Ocean. *Mar. Ecol. Progr. Ser.* 379: 267–278.
- Jespersen P. 1942. Indopacific leptocephalids of the genus *Anguilla*: Systematic and biological studies. Dana Report No 22
- Kuroki, M., J. Aoyama, M. J. Miller, S. Wouthuyzen, T. Arai, K. Tsukamoto. 2006. Contrasting patterns of growth and migration of tropical anguillid leptocephali in the western Pacific and Indonesian Seas. *Mar. Ecol. Progr. Ser.* 309: 233–246.
- Kuroki, M., J. Aoyama, M. J. Miller, S. Watanabe, A. Shinoda, D. J. Jellyman, E. Feunteun, K. Tsukamoto. 2008. Distribution and early life history characteristics of anguillid leptocephali in the western South Pacific. *Mar. Freshw. Res.* 59: 1035–1047.
- Kuroki, M., J. Aoyama, M. J. Miller, T. Yoshinaga, S. Shinoda, S. Hagihara, K. Tsukamoto. 2009. Sympatric spawning of *Anguilla marmorata* and *Anguilla japonica* in the western North Pacific Ocean. *J. Fish Biol.* 74: 1853–1865.
- Minegishi, Y., J. Aoyama, K. Tsukamoto. 2008. Multiple population structure of the giant mottled eel, *Anguilla marmorata*. *Molecular Ecology* 17: 3109–3122.
- Réveillac, E., E. Feunteun, P. Berrebi, P. A. Gagnaire, R. Lecomte-Finiger, P. Bosc, T. Robinet. 2008. *Anguilla marmorata* larval migration plasticity revealed by otolith microstructural analysis. *Can. J. Fish. Aquat. Sci.* 65: 2127–2137.
- Réveillac, E., P. A. Gagnaire, R. Lecomte-Finiger, P. Berrebi, T. Robinet, P. Valade, E. Feunteun. 2009. Development of a key using morphological characters to distinguish south-western Indian Ocean anguillid glass eels. *J. Fish Biol.* 74: 2171–2177.
- Robinet, T., R. Lecomte-Finiger, K. Escoubeyrou, E. Feunteun. 2003. Tropical eels *Anguilla* spp. recruiting to Reunion Island in the Indian Ocean: taxonomy, patterns of recruitment and early life histories. *Mar. Ecol. Progr. Ser.* 259: 263–272.
- Robinet, T., E. Feunteun, P. Keith, G. Marquet, J. M. Olivier, E. Réveillac, P. Valade. 2007. Eel community structure, fluvial recruitment of *Anguilla marmorata* and indication for a weak local production of spawners from rivers of Réunion and Mauritius islands. *Envir. Biol. Fish.* 78, 93–105.
- Robinet, T., E. Réveillac, M. Kuroki, J. Aoyama, K. Tsukamoto, M. W. Rabenevanana, P. Valade, P. A. Gagnaire, P. Berrebi, E. Feunteun. 2008. New clues for freshwater eels (*Anguilla* spp.) migration routes to eastern Madagascar and surrounding islands. *Mar. Biol.* 154: 453–463.

Migration pathways of Anguilliform Leptocephali in the Southwest Indian Ocean

Raymonde LECOMTE-FINIGER, Eric FEUNTEUN, Mari KUROKI,
Michael J. MILLER, Elodie REVEILLAC,
Katsumi TSUKAMOTO, and Tsuguo OTAKE

The small size and long development time of leptocephali have made studies on their dispersal difficult. Fish larvae usually cannot be burdened with tags, and high mortality and potentially widespread dispersal make the recovery of tags problematic. Fortunately, fish carry their own internal environmental recorders in the form of otoliths used for hearing, balance, and orientation. Otoliths are composed of calcium carbonate crystals (aragonite) within a protein matrix. They are formed at birth and grow daily as new crystal layers are deposited around the existing core by the precipitation of calcium.

Because the otoliths are metabolically inert, the elemental signature embedded in them can be an indicator of the characteristic of surrounding water masses and can be considered as proxies of environmental conditions (Campana 1999). The use of stable isotopes in otoliths offers an alternative approach to infer patterns of migration because they are generally formed in or near the O-isotope equilibrium with the mean water temperature. The oxygen isotopic composition is a reliable estimator of sea water temperature while carbon isotopic data can provide information relating to metabolic processes and the source of carbon involved in calcification.

This study using otoliths from leptocephali collected during Leg 5 of the KH-09-5 cruise in the Indian Ocean will utilize both anguillid and marine eel species of leptocephali as outlined below.

I - Anguillid leptocephali

Our aim is to clarify the routes of oceanic larval migration of anguillid eels by examining the stable oxygen (and in a minor way carbon) isotopic composition of leptocephalus otoliths. Two central questions will be addressed to learn about temperature of the water masses in which anguillid leptocephali migrate:

1. At what average depth (or t°) do the larval anguillid eels of the Indian Ocean migrate?

To address this first question, we will use the left sagittal otolith to measure the Oxygen and Carbon isotope ratios in the whole otolith. The results will be compared to the isotope ratio of water masses measured at various depths and locations. Route discrimination is based on the observation that otoliths of leptocephali inhabiting different water bodies will reflect the environmental conditions to form a unique isotopic signature in relation with the environmental conditions. This signature will reflect the proportion of time that the larvae occupied the particular water mass.

2. What are the changes of water masses used by leptocephali during their migration?

Little is known about the migration pathways of anguillid eels. Available studies on *A. japonica* and on Atlantic eels suggest that there is a shift in depth preference according to age. Recently hatched leptocephali have been mainly caught at range depths from 50 to 350 m, but they shift to being present only in the upper 100 m at night as they grow and are deeper during the day (Castonguay and McCleave 1987). This diel vertical migration pattern has been described for eel leptocephali only in a few areas of the ocean (Miller 2009), and there is no information on their vertical migration patterns in the southwest Indian Ocean.

For this analysis we will use the right sagitta to obtain ablation core samples (Nano sims or Ion Probe), from the core to the edge of the otolith. This should be a collaborative work between Japanese & French teams and the place where the analysis will be conducted needs to be decided. The otoliths of anguillid leptocephali caught in 2006 and in 2010 will hopefully be analysed.

Materials and methods

For the anguillid leptocephali, the front half of the body, including the head was preserved in ethanol, and will go to Japan (M. Kuroki). The otoliths will be extracted and the left one will be sent to R. Lecomte in Perpignan. The left one will be polished to the core for age reading (M. Kuroki) and then used for microchemical analysis (T. Otake and E. Reveillac).

II – Other anguilliform leptocephalus larvae

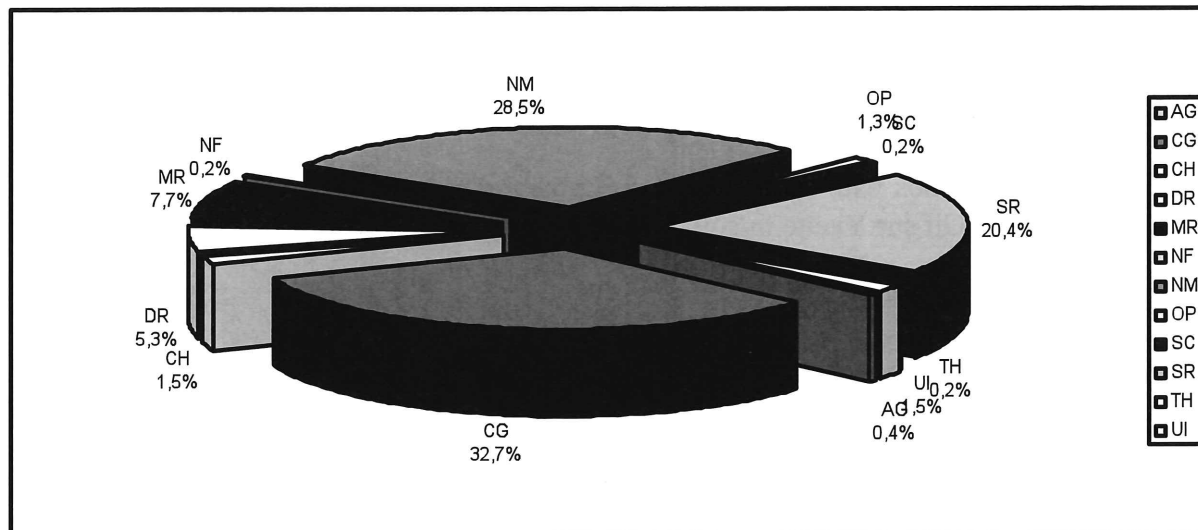
As is true for freshwater anguillid eels, little is known about the migration pathways of other anguilliform leptocephalus larvae. The few available studies on their depth distributions describe similar patterns as for anguillid eels, with an occurrence in the upper 100 m at night and sometimes deeper during the day (Miller 2009). There is evidence for differences in depth preference according to species and season. Mesopelagic eel leptocephali in the Atlantic, such as *Serrivomer* were caught in the upper 50 m layer, while other species were mainly found at deeper depths (Castonguay and McCleave 1997). The variation of depth preference according to age is not well known though due to a lack of studies using appropriate sampling gear. It may be hypothesized that the age/depth relationship varies among taxa, especially according to different migration loops or possible feeding preferences. Indeed, different patterns may be expected between freshwater eels, coastal eels (*i.e.* Muraenidae, Ophichthidae), and mesopelagic eels (*i.e.* Nemichthyidae) that have different spawning and recruitment areas. Between species or within species, age related differences of water depth preferences may be detected using the Oxygen isotope ratio of their otoliths.

Methods and work on board

All the leptocephali were sorted and identified to the lowest possible taxonomic level using morphological and meristic analysis. A total of 579 leptocephalus larvae were caught and the most abundant families were Congridae (32.7%), Nemichthyidae (28.5%) and Serrivomeridae (20.4%) as shown in Figure 1 (see leptocephali report).

We designed a size stratified sampling plan in the three main families to test variation of depth preference according to age/size. We collected a total of 107 leptocephali including 48 Congridae, 27 Nemichthyidae, and 18 Serrivomeridae, and from 1 to 3 specimens per family were collected of six other families (Fig. 1, bottom panel).

(replace figure to have no “NF” category)



	AG	CG	CH	DR	MR	NM	OP	SC	SR	TH	UI	
N =	2	48	2	3	3	27	3	0	18	1	0	107

Figure 1. Leptocephalus assemblage composition in % of the total number of leptocephali collected (top panel). The number of specimens of each family that was preserved for analysis is shown in the lower panel. AG: Anguillidae; CG: Congridae, CH: Chlopsidae, DR: Derichthyidae, MR: Muraenidae, NM: Nemichthyidae, OP: Ophichthidae, SC: Saccopharyngidae, SR: Serrivomeridae; TH: *Thalassenchelys*, UI: unidentified.

When specimens were over ca. 15 mm, heads were removed and placed in a vial that will be taken to Japan, where M. Kuroki will extract the otoliths and estimate the larval age using SEM. Then she will send both otoliths (left one whole and right one ground) to France for oxygen isotope analysis following the same procedure as that described for anguillid eels.

When specimens were smaller than ca. 15 mm they were preserved directly in ethanol vials. These specimens will be taken to Perpignan University and otoliths will be extracted. Age will tentatively be analysed without either imbedding nor grinding the otoliths. Then oxygen isotope composition will be analysed by R. Lecomte in pooled samples.

III – Water sampling.

In order to compare the relation between Oxygen isotope ratios in the water and in the otoliths, and to compare them to 2006 data, we collected 50 ml samples of water at various depths (surface and at the fluorescence max).

$\delta^{18}\text{O}$ ratio will be measured in France in collaboration with D. Blamart.

Water samples were collected at Stn. 1, 3, 6, 10, 13, 15, 18, 21, 22, 25, 27, 30, 33, 38.

Samples of gelatinous plankton were collected in order to determine their oxygen ratios:

- Hydrozoa (3 specimens)
- Siphonophora (3 specimens)
- Chaetognathe (3 specimens)
- Pteropoda (3 specimens)

The $\delta^{18}\text{O}$ isotope ratios with spectrometry in Laboratoire du Climat et des Sciences de l'Environnement-Gif sur Yvette in collaboration with D. Blamart.

References

Campana SE. (1999) Chemistry and composition of fish otoliths: pathways, mechanisms and applications. *Mar. Ecol. Progr. Ser.* 188: 263–297.

Castonguay M, McCleave JD. (1987) Vertical distributions, diel and ontogenetic vertical migrations and net avoidance of leptocephali of *Anguilla* and other common species in the Sargasso Sea. *J. Plankt. Res.* 9: 195–214.

Miller, M. J. (2009). Ecology of anguilliform leptocephali: remarkable transparent fish larvae of the ocean surface layer. *Aqua-BioSci Monographs* 2(4): 1–94.

Leptocephali Collected west of the Nazareth Bank of the Mascarene Ridge during Leg 5 of KH-09-5

All Scientists onboard

The sampling survey of Leg 5 of the KH-09-5 collaborative cruise between scientists from France and Japan collected a total of 578 leptocephali of 11 anguilliform families that included at least 50 species (Table 1). An Isaacs Kidd Midwater trawl (IKMT) with an 8.7 m² mouth opening and 0.5 mm mesh was used to conduct an intensive grid survey at 40 stations to the west of the Nazareth Bank. This large shallow bank is the southernmost bank of the Mascarene Ridge and likely provides habitats to a variety of marine eels of the families that live at depths less than a few hundred meters, such as the Congridae, Ophichthidae, and Muraenidae.

Stations were sampled during both day and night, with step tows that fished to a maximum depth of 150 m and then horizontally at depths of about 120, 90, 70, 50, 30 m for 10 min at each depth layer at night. Oblique tows that fished to a maximum depth of about 500 m were conducted during the day. This resulted in 16 nighttime step tows that collected 469 leptocephali and 24 daytime oblique tows that collected 109 leptocephali. See the *Anguilla* report for a map of day and night stations.

Table 1. Catches of various taxa of leptocephali during the Leg 5 collaborative cruise of KH-09-5 conducted to the west of the Nazareth Bank of the Mascarene Ridge. Estimated number of species is in parentheses.

Anguillidae (1)	N	TL range	Ophichthidae (~7)	N	TL range
<i>Anguilla bicolor bicolor</i>	2	51-54	Ophichthinae	10	7-136
Congridae (15)			Myrophinae	1	80
<i>Ariosoma</i> sp. 2	1	202	<i>Neenchelys</i>	2	27-60
<i>Ariosoma</i> sp. 3	1	96	sp.	3	6-116
<i>Ariosoma</i> sp. 6	1	81	Juvenile/adult	1	118
<i>Ariosoma</i> sp. 7	39	7-156	Synphobranchidae (1)		
<i>Ariosoma</i> sp. 8	3	11-17.0	Synphobranchinae	1	21
<i>Ariosoma</i> sp.	29	9-251	Derichthyidae (2)		
<i>Conger</i>	65	8-97.0	<i>Derichthys</i>	18	9.0-35
<i>Gnathophis</i>	32	10-77.0	<i>Nessorhamphus</i>	1	67
<i>Bathycongrus</i>	3	34-130	sp.	8	9-15.0
Congrinae	2	24-88	Nemichthyidae (1)		
<i>Gorgasia</i>	7	39-51	<i>Nemichthys</i>	108	10-243
sp.	8	8-31.0	<i>Avocettina</i>	34	6-203
Chlopsidae (6)			sp.	4	8-120
<i>Kaupichthys</i>	1	20	Serrivomeridae (2)		
<i>Robinsia caterinae</i>	1	50	sp.	106	7-63.0
sp. 5	2	46-55	Juvenile	1	99
sp.	3	17-19	Cyematidae (1)		
Muraenidae (12)			<i>Cyema atrum</i>	1	18
Muraeninae	25	14-67	Eurypharyngidae (1)		
Uropterigyinae	19	10-54.0	<i>Eurypharynx pelecanoides</i>	1	26
sp.	7	5-10.0	Thalassenchelys (1)		
			<i>Thalassenchelys</i>	1	49
			Unidentified	28	4-18.0
Total				580	

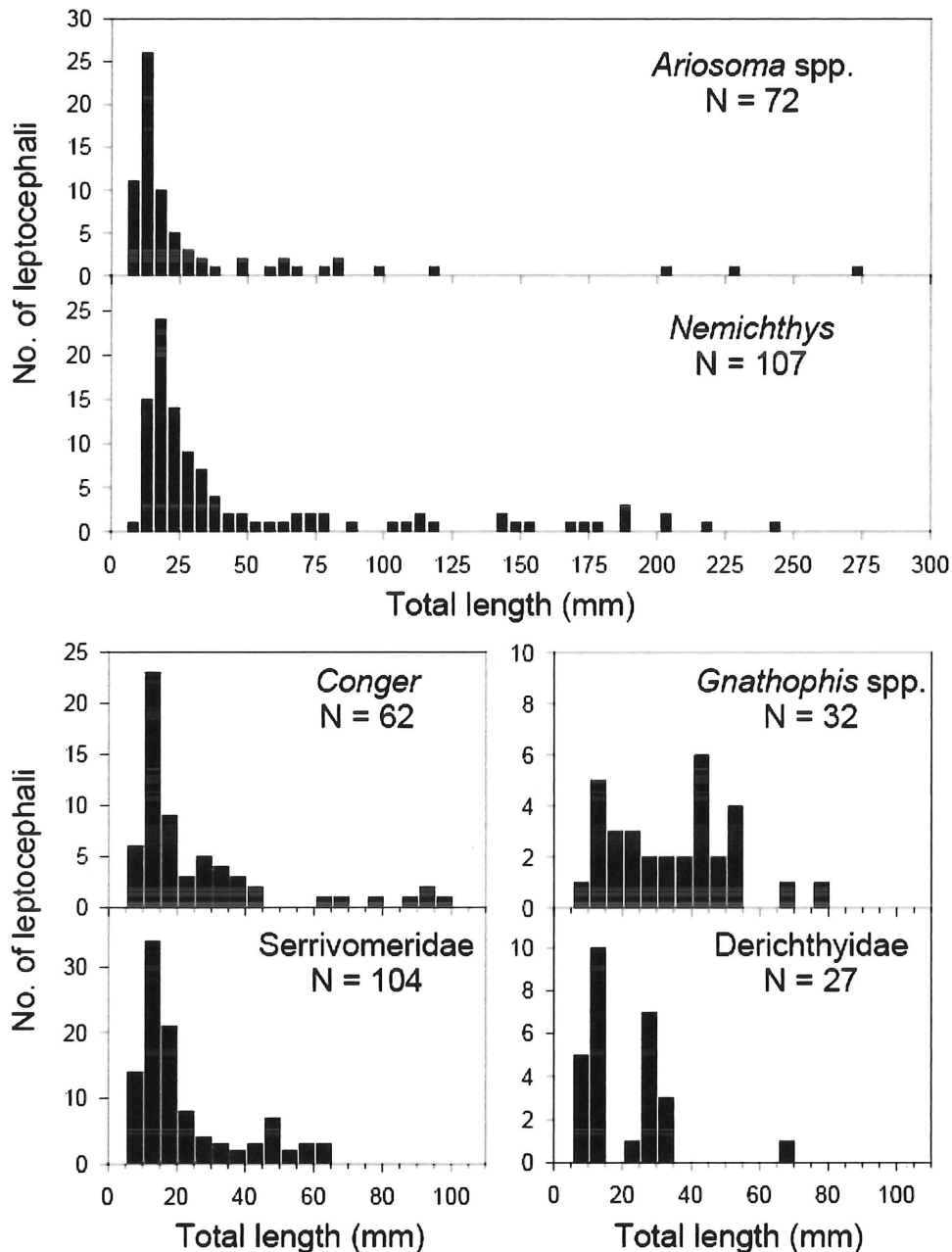


Figure 1. Length frequency distributions of several of the most abundant taxa of leptocephali collected during Leg 5 of KH-09-5 conducted to the west of the Nazareth Bank of the Mascarene Ridge.

Overall, the leptocephali of the shelf and slope marine eels of the family Congridae were the most abundant with 191 larvae being collected, and 15 species being distinguished (Table 1). This family was abundant partly due to many small *Ariosoma* and *Conger* leptocephali being collected (Figure 1). Some larger specimens of these two taxa were collected, but the dominant size of both was the 10-15 mm size class. Some *Gnathophis*, *Bathycongrus*, and *Gorgasia* leptocephali (Figure 2) were also collected, but the latter two taxa were only collected at larger sizes > 30 mm. During the cruise various specimens of the abundant taxa of congrid and other taxa were photographed with a digital imaging system that recorded the details of their external morphology (Figure 3).

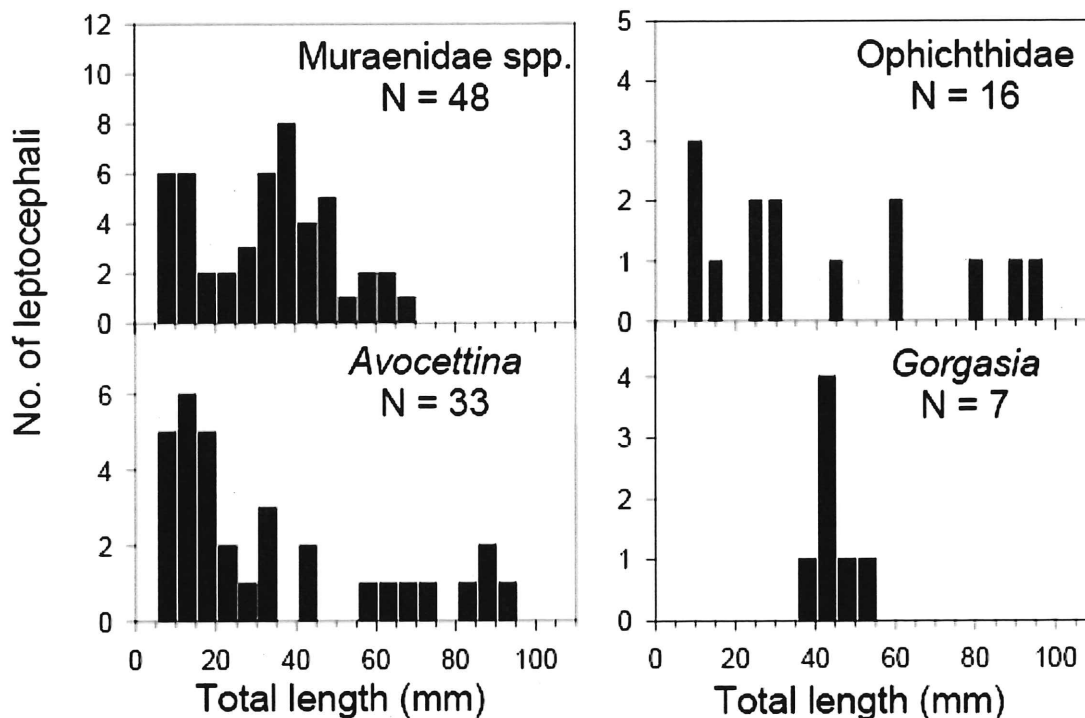


Figure 2. Length frequency distributions of taxa of leptocephali collected during Leg 5 of KH-09-5 conducted to the west of the Nazareth Bank of the Mascarene Ridge.

The leptocephali other families of shelf eels were much less abundant than the congrid, but there were 51 muraenid leptocephali collected (see Muraenidae report) at a wide size range up to almost 70 mm (Figure 1). Chlopsid leptocephali were rare (N = 7, 6 species) and only 16 ophichthid leptocephali of about 7 species were collected (Table 1). There were no Moringuidae or Nettastomatidae collected during the survey.

The next most abundant families after the congrid were the mesopelagic eel leptocephali of the Nemichthyidae and the Serrivomeridae (Table 1). They were collected at almost their complete size ranges, but both *Nemichthys* (N = 108) and the serrivomerids (N = 106) were most abundant at sizes of 10-20 mm (Figure 1). Several very large *Nemichthys* were collected at sizes of 200-250 mm long. There were also 34 *Avocettina* leptocephali (Nemichthyidae) collected at a wide range of sizes, and at least 27 leptocephali of the mesopelagic eel family Derichthyidae were also collected at various sizes (Table 1, Figure 1, 2).

This survey was conducted along the southern end of the Mascarene Ridge where the sampling survey with the IKMT in 2006 indicated that the diversity of leptocephali of shelf eels is lower. The 2010 survey showed however, that several species of congrid (*Ariosoma* and *Conger*) use these latitudes for larval development areas, as also do the leptocephali of the mesopelagic eels of the Nemichthyidae and Serrivomeridae. Future studies will analyze the assemblage structure and life history characteristics of leptocephali collected in this area in 2010 and in 2006 to learn about the early life history of marine eels that live on and around the banks of the Mascarene Ridge. These analyses can then be compared to the more than 1,100 leptocephali collected during Leg 6 of KH-09-5 at about 12 stations in the eastern 2/3 of the Indian Ocean, and the other 2006 stations that were sampled in a transect across the Indian Ocean.

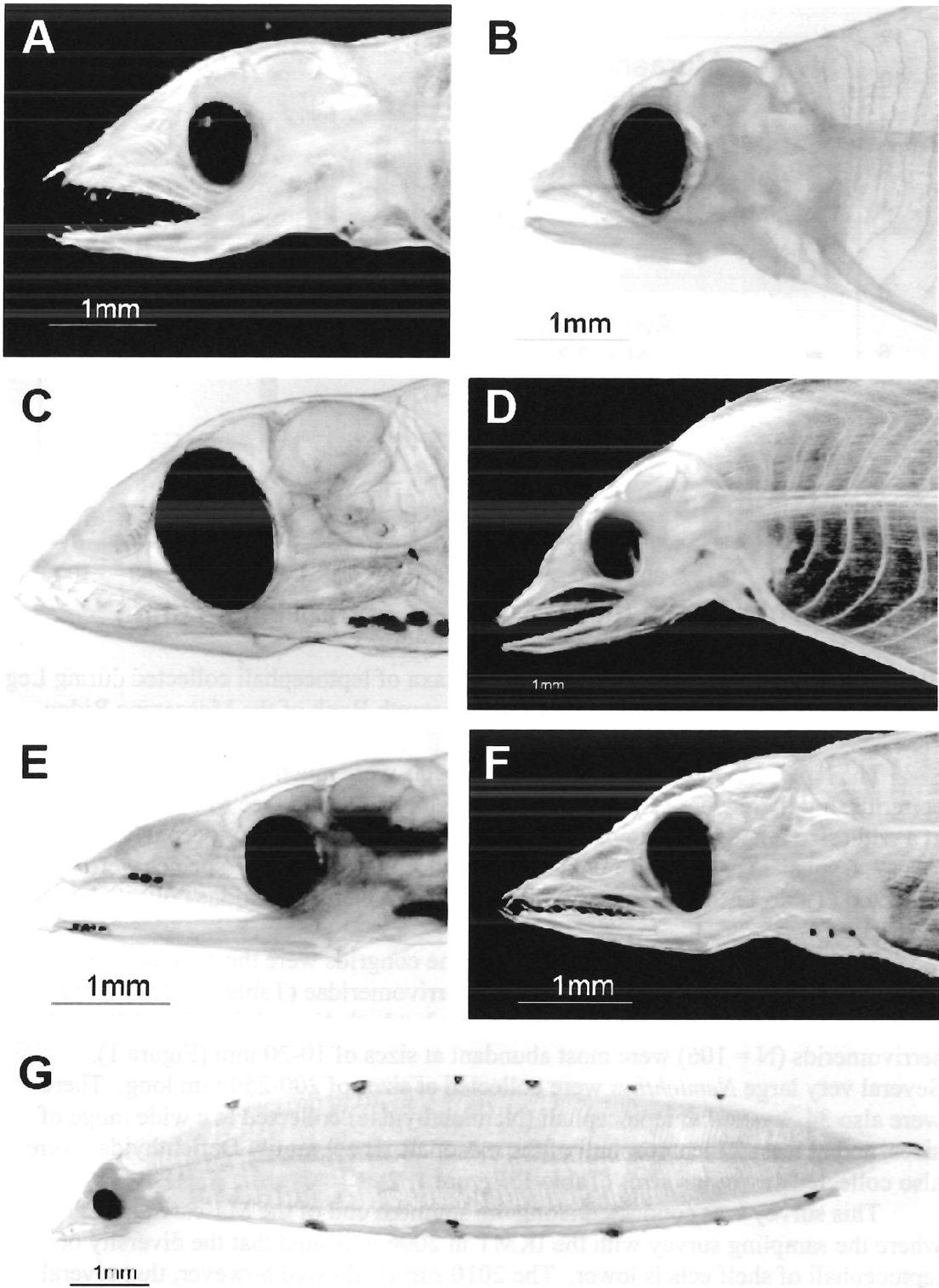


Figure 3. Photographs of various taxa of leptocephali collected during the cruise that include (A) *Gorgasia*, (B) *Derichthys*, (C) *Conger*, (D) *Ariosoma*, (E) Ophichthidae, (F) *Gnathophis* type, and (G) a small *Ariosoma* with dorsal spots that disappear in larger sized leptocephali of that species.

Taxonomical Identification of Muraenid Leptocephali Collected During the KH-09-5 Leg 5 Cruise using Morphology and DNA Barcoding.

Atsushi Tawa

Moray eels (family: Muraenidae) are one of the most diverse families of eels and more than at least 40 species have been reported in the Red Sea and western Indian Ocean (Smith 1962, Randall et al. 1995). However, only a few species or types of leptocephali have been reported (D'Ancona 1928, Nair et al. 1949,1960, Jones and Pantalu 1952). The Muraenidae is a fairly homogeneous group whose leptocephali show little fundamental variation. Genera and species groups are characterized mainly by pigmentation and fin position (Smith 1989).

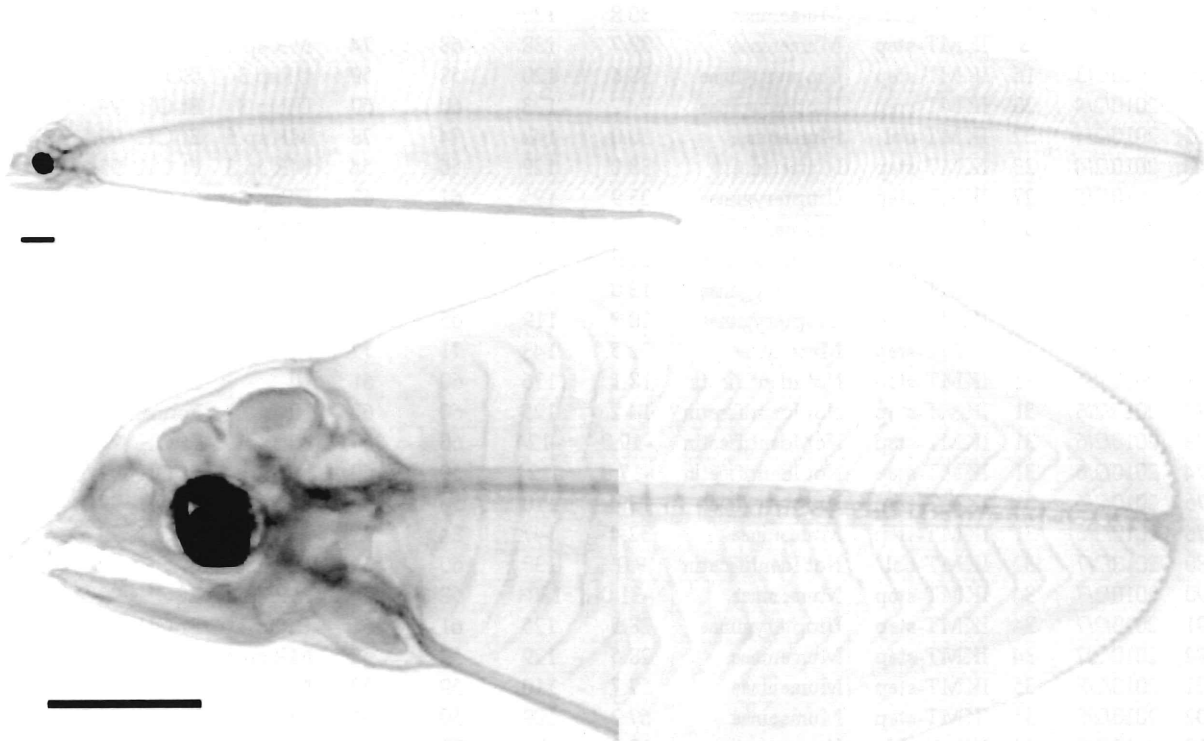


Figure 1. Lateral view of whole body, head and tail tip of UR sp. 3 type (No. 138). Bars 1 mm.

A total of 51 specimens of muraenid leptocephali were collected during the KH-09-5 Leg 5 cruise (Table 1). These specimens could be easily distinguished into 7 types of Muraeninae and 5 types of Uropterygiinae by morphology and pigmentation patterns onboard. The most abundant species was UR sp. 3 characterized by no pigment (Figure 1) and total myomere 119-128, which nearly corresponds with *Anarchias seychellensis* Smith, 1962 (Total vertebra VN: 121-132, Böhlke et al. 2001). Some VN overlap in adults, so it is very difficult to identify muraenid leptocephali to the species level morphologically, so DNA will be used.

Table 1 Sampling stations and body features of KH-09-5 Leg 5 muraenid leptocephali.

No.	Date	St.	Net	Subfamily	TL	TM	LVBV	PAM	Type	Fix.
20	2010/1/30	3	IKMT-step	Uropterygiinae	64.1	137	66	79	URsp.5	Et-OH 99.5%
26	2010/1/30	3	IKMT-step	Muraeninae	42.5	108	55	54	MR sp.1	Et-OH 99.5%
27	2010/1/30	3	IKMT-step	Muraeninae	44.2	126	57	54	MR sp.3	Et-OH 99.5%
90	2010/1/31	5	IKMT-step	Muraeninae	40.0	120	54	54	MR sp.3	Et-OH 99.5%
91	2010/1/31	7	IKMT-step	Muraeninae	46.5	124	72	81	MR sp.2	Et-OH 99.5%
92	2010/1/31	7	IKMT-step	Muraeninae	58.4	142	69	76	MR sp.6	Et-OH 99.5%
114	2010/1/31	7	IKMT-step	Muraeninae	40.2	145	83	99	MR sp.7	Et-OH 99.5%
136	2010/2/1	8	IKMT-step	Uropterygiinae	36.5	122	61	62	URsp.3	Et-OH 99.5%
138	2010/2/1	8	IKMT-step	Uropterygiinae	41.3	125	59	59	URsp.3	Et-OH 99.5%
147	2010/2/1	8	IKMT-step	Uropterygiinae	37.2	124	62	62	URsp.3	Formalin 10%
148	2010/2/1	8	IKMT-step	Uropterygiinae	39.7	120	59	59	URsp.3	Et-OH 99.5%
149	2010/2/1	8	IKMT-step	Uropterygiinae	35.8	120	59	61	URsp.3	ISO Et-OH 99.5%
150	2010/2/1	8	IKMT-step	Uropterygiinae	45.5	128	61	60	URsp.3	Et-OH 99.5%
151	2010/2/1	8	IKMT-step	Uropterygiinae	34.3	123	58	59	URsp.3	ISO Et-OH 99.5%
152	2010/2/1	8	IKMT-step	Uropterygiinae	37.5	121	59	61	URsp.3	Et-OH 99.5%
153	2010/2/1	8	IKMT-step	Muraeninae	61.1	111	57	55	MR sp.1	Et-OH 99.5%
159	2010/2/1	9	IKMT-obl	Muraeninae	30.8	130	64	69	MR sp.4	Et-OH 99.5%
160	2010/2/1	8	IKMT-step	Muraeninae	39.7	138	68	74	MR sp.5	Et-OH 99.5%
270	2010/2/3	16	IKMT-step	Uropterygiinae	38.6	120	58	59	URsp.3	ISO Et-OH 99.5%
302	2010/2/4	20	IKMT-step	Uropterygiinae	34.2	123	60	60	URsp.3	Et-OH 99.5%
312	2010/2/4	22	IKMT-obl	Muraeninae	50.0	142	74	78	MR sp.6	Et-OH 99.5%
344	2010/2/4	23	IKMT-step	Muraeninae	33.0	129	56	54	MR sp.3	Et-OH 99.5%
447	2010/2/5	27	IKMT-step	Uropterygiinae	23.9	125	61	62	URsp.3	Et-OH 99.5%
459	2010/2/6	31	IKMT-step	Uropterygiinae	15.7	132	62	57	URsp.4	Et-OH 99.5%
460	2010/2/6	31	IKMT-step	Uropterygiinae	13.9	116	64	59	URsp.2	Et-OH 99.5%
461	2010/2/6	31	IKMT-step	Uropterygiinae	13.0	121	56	57	URsp.3	Et-OH 99.5%
462	2010/2/6	31	IKMT-step	Uropterygiinae	10.9	119	65	72	URsp.3	Et-OH 99.5%
470	2010/2/6	31	IKMT-step	Muraeninae	22.5	143	71	77	MR sp.6	Et-OH 99.5%
471	2010/2/6	31	IKMT-step	Not Identificatin	18.1	126	60	61	NI	Et-OH 99.5%
472	2010/2/6	31	IKMT-step	Not Identificatin	14.2	125	60	62	NI	Formalin 10%
473	2010/2/6	31	IKMT-step	Not Identificatin	~19.0	~124	60	57	NI	Et-OH 99.5%
474	2010/2/6	31	IKMT-step	Not Identificatin	~17.8	~122	59	60	NI	Et-OH 99.5%
475	2010/2/6	31	IKMT-step	Uropterygiinae	13.4	110	56	59	URsp.1	Et-OH 99.5%
476	2010/2/6	31	IKMT-step	Muraeninae	32.4	147	84	98	MR sp.7	Et-OH 99.5%
480	2010/2/7	32	IKMT-obl	Not Identificatin	9.0	130	60	62	NI	Et-OH 99.5%
490	2010/2/7	34	IKMT-step	Muraeninae	~31.0	130+	58	54	MR sp.3	Et-OH 99.5%
491	2010/2/7	34	IKMT-step	Uropterygiinae	28.5	125	61	61	URsp.3	Et-OH 99.5%
492	2010/2/7	34	IKMT-step	Muraeninae	28.5	129	59	56	MR sp.3	Et-OH 99.5%
501	2010/2/8	35	IKMT-step	Muraeninae	57.7	110	59	57	MR sp.1	Formalin 5%
502	2010/2/8	35	IKMT-step	Muraeninae	67.2	109	59	56	MR sp.1	Et-OH 99.5%
509	2010/2/8	36	IKMT-obl	Uropterygiinae	18.9	124	57	60	URsp.3	Et-OH 99.5%
510	2010/2/8	36	IKMT-obl	Muraeninae	47.3	140	70	76	MR sp.6	Et-OH 99.5%
511	2010/2/8	36	IKMT-obl	Muraeninae	50.0	142	70	77	MR sp.6	Et-OH 99.5%
524	2010/2/9	38	IKMT-step	Not Identificatin	9.6	134+	-	-	NI	Et-OH 99.5%
561	2010/2/9	38	IKMT-step	Not Identificatin	11.0+	150+	98	119	NI	Et-OH 99.5%
565	2010/2/9	38	IKMT-step	Not Identificatin	8.0	-	-	-	NI	Et-OH 99.5%
566	2010/2/9	38	IKMT-step	Not Identificatin	6.7	-	-	-	NI	Et-OH 99.5%
569	2010/2/9	38	IKMT-step	Not Identificatin	6.1	-	-	-	NI	Et-OH 99.5%
573	2010/2/9	38	IKMT-step	Not Identificatin	Egg 3.0	-	-	-	NI	E-vial Et-OH 99.5%
574	2010/2/9	38	IKMT-step	Muraeninae	52.1	139	68	76	MR sp.6	Et-OH 99.5%
575	2010/2/9	38	IKMT-step	Muraeninae	47.9	144	84	95	MR sp.7	Et-OH 99.5%

Leptocephalus food web

Alexandre CARPENTIER, Christine DUPUY, Celine ELLIEN, Eric FEUNTEUN, Mari KUROKI, Raymonde LECOMTE-FINIGER, Michael MILLER, Tsuguo OTAKE, Stéphane POUS, Elodie REVEILLAC, Tony ROBINET, & Katsumi TSUKAMOTO

Leptocephalus diet remains poorly documented and the subject of much debate. This is mainly due to the fact that no zooplankton has been seen in the digestive tract. This observation made that the sources of food remained for a long time enigmatic. Some investigators have suggested that the digestive tract would not be functional that larvae fed on plankton, or on gelatinous plankton (Jellyfishes, Siphonophora, Scyphomedusae, Ctenophores and Appendicularians) or that larvae fed essentially on dissolved organic matter (particulate organic matter, POM and dissolved organic matter, DOM) and on detrital particles. The direct absorption of Dissolved Organic Carbon was also hypothesized but this theory was finally shown to be wrong.

Studies based on Isotope Ratios of N & C showed that the trophic level of several leptocephalus taxa were very low (Otake et al. 1993). Other studies pointed out the presence of ciliates and faecal pellets in gut contents suggesting that these food items could represent key food items for leptocephalus.

The aim of the present study is therefore to clarify the respective roles of particulate organic matter and small organisms as ciliates, copepods, larvae, etc. in the diet of leptocephalus larvae's diets. We also aimed at comparing the diet of fish larvae which to understand if they compete with leptocephalus or if they feed on different food sources. Therefore, we will study C & N stable isotope ratio among various taxa and potential food resources and leptocephali taxa.

To achieve this:

1. We sampled leptocephali larvae which were selected among the families caught during the survey.
2. We filtered water from various depths to select total Particulate Organic Matter at various depths
3. We selected a range of organisms (chaetognath, Copepods and small crustaceans, gelatinous, larvacean houses,...) that may be food items for leptocephali

4. We also selected a range of fish larvae with comparable sizes as leptocephali.

I- Sampling design for leptocephali

A total of 110 leptocephalus larvae were sampled. They were selected among ca. 520 leptocephali larvae sampled during night and day tows. The samples were all preserved in ethanol but processed in different ways according to size. All the details of the preservation are given in the lepto datasheet and summarized below in table 1.

1. Small leptos (*ca.* 15 mm) have been preserved in ethanol vials. They will be taken and processed in France. Otoliths will be extracted for oxygen isotope analysis by Raymonde Lecomte for migration pathways study (see relevant section for details). The bodies will be used for C & N isotope analysis by the French Museum and Perpignan University
2. Larger specimens were cut into two parts.
 - The head was preserved in vials, kept on board Hakuho Mary and shipped to Tokyo University for Ageing (Mari Kuroki). For each specimen, one of the otoliths will be ground to core for age reading at the University of Tokyo. The second one will be kept in a vial. Both otoliths will then be sent to France to Eric Feunteun at the French Museum for the study on the migration pathways of leptocephali (see relevant section for details). Molecular studies (Jun Aoyama).
 - The bodies were preserved in ethanol and taken to France at the French Museum by Eric Feunteun for C & N isotope analysis. Work will be done in collaborative work between University of Perpignan and the French Museum.

Table 1. Hakuho Maru Cruise 2010 Leg HK-09-5 sample design. Explanation of the database with samples preservation per family

Family	Preservation	Body part	Aim	Destination	NB of samples
	F	Gut	The gut was preserved in formalin for analysis of gut content in Japan (Prof. Otake).	JAPAN	2
	Ethanol	Rear half	The rear half of the body was preserved in ethanol and will go to France (Elodie Réveillac & Eric Feunteun) for stable isotopes $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ analysis.	FRANCE (Dinard)	2
	Ethanol	Frontal half including head	The frontal half of the body including the head was preserved in ethanol and will go first to Japan. There, otolith ageing will be performed (Mari Kuroki & Jun Aoyama) on one otolith that will thereafter be used by the French (Elodie Réveillac, Eric Feunteun in coll. with Raymonde Lecomte & Dominique Blamart) for oxygen stable isotopes ($\delta^{18}\text{O}$) analysis by ablation with ion probe/nanoSIMS. The second otolith will also be sent to France for in toto $\delta^{18}\text{O}$ analysis by mass spectrometry (Raymonde Lecomte & Dominique Blamart in coll. with Elodie Réveillac, Eric Feunteun).	JAPAN and then FRANCE (Dinard & Perpignan)	2
Other Anguilliform leptocephali	F	Whole larvae or part of	Formalin preserved larvae were not cut, they will go to Museum collections in Japan and France.	JAPAN & FRANCE (Paris)	42
	E-vial	Whole larvae	Larvae entirely preserved in E-vial with ethanol will go to Japan for genetic analyses.	JAPAN	47
	DNA	Whole larvae or part of	Body or piece of body preserved in ethanol will go to Japan or to France (Tony Robinet).	JAPAN or FRANCE (Paris)	4
	FR	Whole larvae or part of		JAPAN	270

						58	JAPAN and then FRANCE (Dinard & Perpignan)
	Head		ISO	Heads of larvae were cut, preserved in E-vial with ethanol and will be sent to Japan for otolith ageing (Mari Kuroki and Jun Aoyama) then both otoliths will be sent to France for $\delta^{18}\text{O}$ analyses by ablation with ion probe/nanoSIMS on the right Sagitta (Elodie Réveillac, Eric Feunteun in coll. with Raymonde Lecomte & Dominique Blamart) and for in toto $\delta^{18}\text{O}$ and $\delta^{13}\text{C}$ analyses by mass spectrometry on the left Sagitta (Raymonde Lecomte & Dominique Blamart in coll. with Elodie Réveillac, Eric Feunteun).		58	FRANCE (Dinard, Perpignan & Paris)
	Body			Bodies of larvae were preserved in plastic bags in ethanol and will be sent to France for analyses of $\delta^{13}\text{C}$, $\delta^{15}\text{N}$ and $\delta^{18}\text{O}$ composition by mass spectrometry (Eric Feunteun, Elodie Réveillac, Raymonde Lecomte & Dominique Blamart). A small piece has been preserved in ethanol for genetic barcoding in French National Museum (Tony Robinet).		39	FRANCE (Perpignan)
	Whole larvae		ISO-E-vial	Entire larvae were preserved in E-vial with ethanol (small specimens) and will be sent to France for stable isotopes analyses of $\delta^{13}\text{C}$ (otoliths and body), $\delta^{15}\text{N}$ (body) and $\delta^{18}\text{O}$ (otoliths and body) by Raymonde Lecomte & Dominique Blamart in coll. with Elodie Réveillac, Eric Feunteun.		4	JAPAN and then FRANCE (Dinard & Perpignan)
	Head		ISO-Lipid	Heads of larvae were cut, preserved in E-vial with ethanol and will be sent to Japan for otolith ageing (Mari Kuroki and Jun Aoyama) then both otoliths will be sent to France for $\delta^{18}\text{O}$ analyses by ablation with ion probe/nanoSIMS on the right Sagitta (Elodie Réveillac, Eric Feunteun in coll. with Raymonde Lecomte & Dominique Blamart) and for in toto $\delta^{18}\text{O}$ and $\delta^{13}\text{C}$ analyses by mass spectrometry on the left Sagitta (Raymonde Lecomte & Dominique Blamart in coll. with Elodie Réveillac, Eric Feunteun).		4	France (Paris)
	Body			One part of the body was preserved in ethanol. It will be processed as in ISO labels. In addition another part of the bodies were frozen and will be taken to France (Céline Ellien) for fatty-acid analyses in comparison with gobies.		4	

As for the migration pathway study, the specimens were chosen in order to address three main questions.

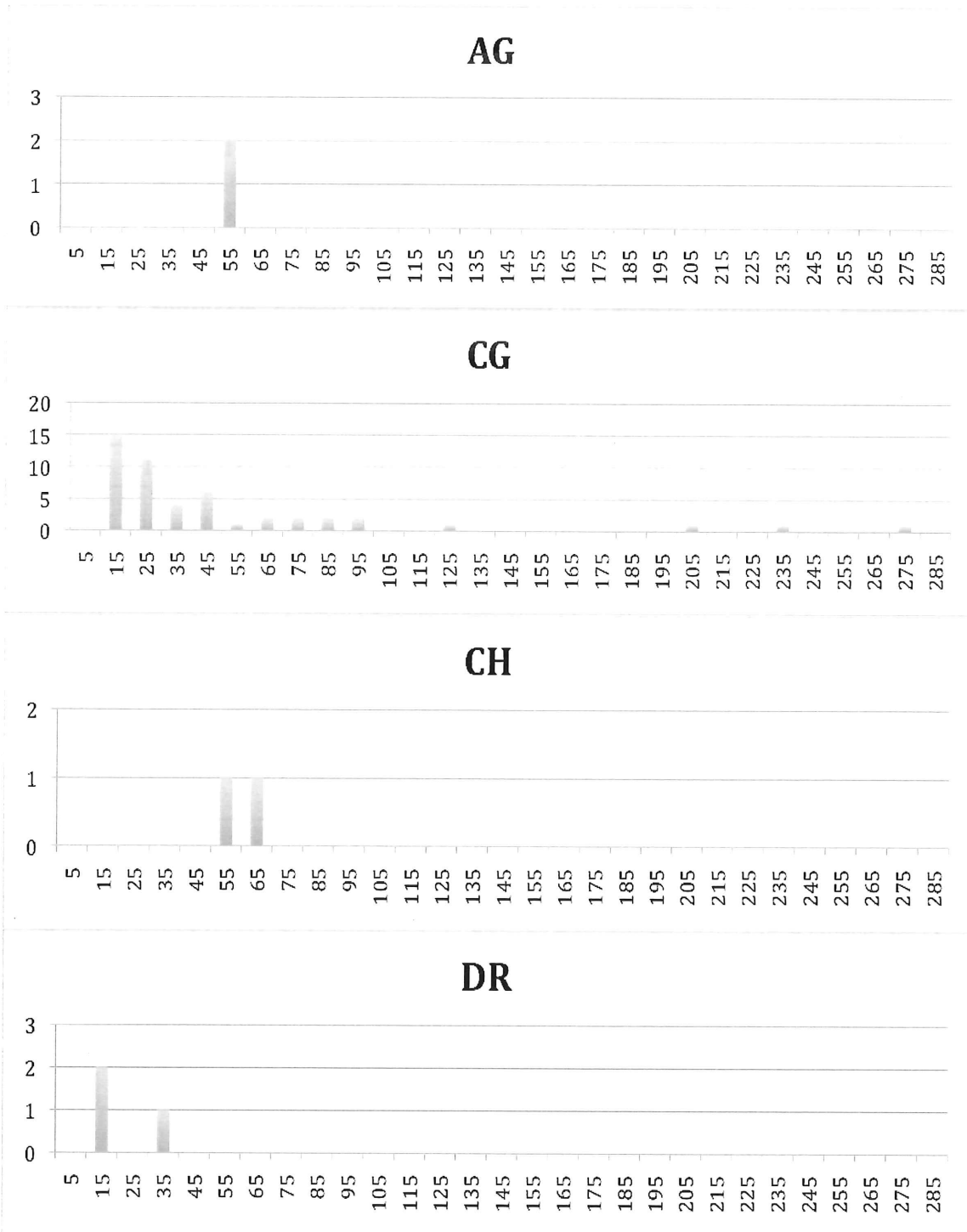
1. Are there differences of trophic level between taxa? To study this question, we selected one to three or more specimens of similar sizes from 9 families (table 2 and figure 1).
2. Does the diet shift according to size and age of leptocephalus and are there different of size / diet patterns among the three main families of anguilliform leptocephalus assemblages (Congridae, Nemichthyidae and Serrivomeridae) ? In this case, we selected 49 serrivomeridae, 29 nemichthyidae and 18 serrivomeridae. There size range of selected specimens was similar to those of the whole sample (table 2 and figure 1).
3. Are there spatial differences of leptocephalus diets in the south west Indian Ocean? This question will be addressed by comparing results from different stations.

The sample design

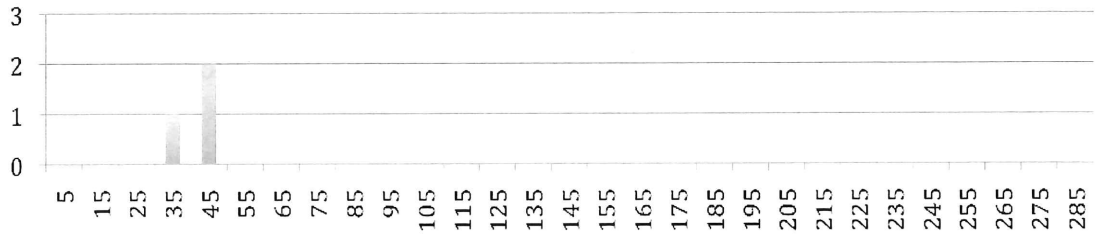
Table 2. Number of individuals caught per family in each station that will be sent to France for stable isotopes composition analyses in otoliths and body tissues, according to Table 1. AG = Anguillidae; CG = Congridae; CH = Chaupychthidae; DR = Derrichthidae; NM = Nemichthyidae; OP = Ophichthyidae; SR = Serrivomeridae; TH = Thallassonchelys.

Station number	AG	CG	CH	DR	MR	NM	OP	SR	TH	Total
3		14	2			7		5	1	29
7		6				5		3		14
8					2					
10	1									1
11		3		1		6		4		14
13		6								6
15		2				5		2		9
16					1					
19	1	1				3		3		8
23		10		2		1	1			14
25							1			1
27		3								3
31		1								1
34		2					1	1		4
38		1				2				
Total	2	49	2	3	3	29	3	18	1	110

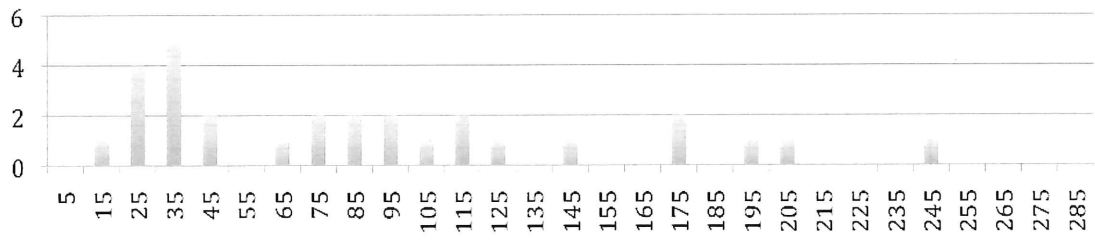
Figure 1. Histograms of size class frequencies (number of individuals) per family that will be sent to France for stable isotopes analyses. AG = Anguillidae; CG = Congridae; CH = Chaupythidae; DR = Derrichthidae; NM = Nemichthyidae; OP = Ophichthyidae; SR = Serrivomeridae; TH = Thallasonychelys.



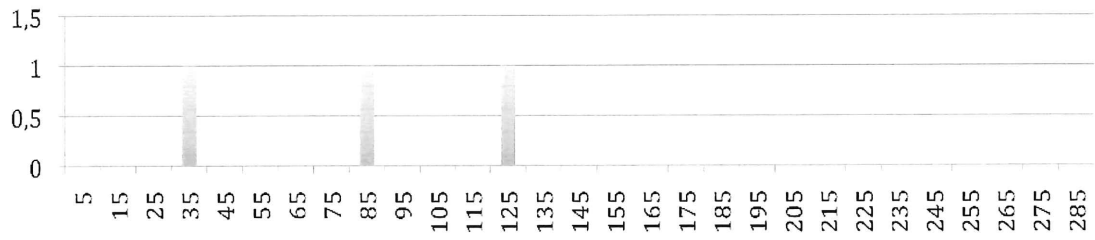
MR



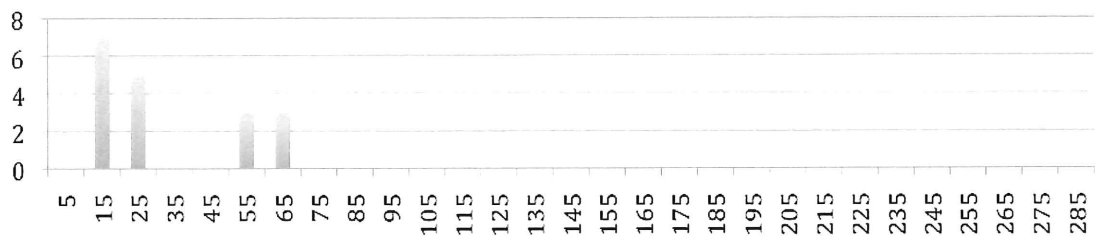
NM



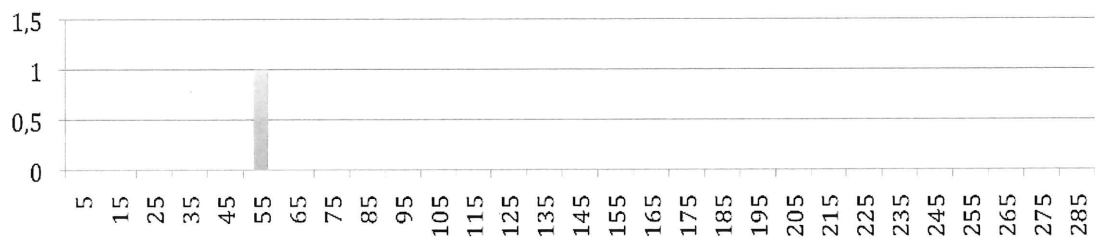
OP



SR



TH



3. SWIO FOOD WEB study

Report of the cruise Hakuho-Maru-09-5: Alexandre Carpentier, Christine Dupuy, Eric Feunteun, Mari Kuroki, Michael Miller, Tsuguo Otake, Stephane Pous, Elodie Reveillac, Tony Robinet, Katsumi Tsukamoto, Shun Watanabe

The Table 3 is a summary of the action, number of the samples during the cruise.

Table 3: Summary of the number of samples during the cruise

N° Station	Chlorophyll <i>a</i> by size class ($\geq 20\mu\text{m}$, 5-20 μm , $\leq 5\mu\text{m}$)	Microplankton (40-300 μm) analysis with flowcam	Stable isotopes of water by size class ($\leq 200\mu\text{m}$ and $\leq 5\mu\text{m}$)	Stable isotopes of mesozooplankton	Stable isotopes of fish larvae	Stable isotopes of small fishes	Prokaryote diversity
1	12	4	15	4	74	20	
2				8			
3	12	4	16	6	74	20	
6	12	4	16	6	74	20	
7				6			
10	12	4	16	6	74	20	4
12				2			
13	12	4	16	7	74	20	
15	9	3	12	10	74	20	3
18	12	4	16	2			
19				1	74	20	
21					74	20	
25	12	4	16	4	74	20	
27	12	4	16	5	74	20	5
29				1			
30	12	4	16	7	74	20	
33	12	4	16		74	20	
35	12	4	16	10	74	20	
38	12	4	16	7	74	20	
40	12	4	16	8	74	20	
Total	165	55	219	100	1110	300	12
Analysis	University of La Rochelle	University of La Rochelle	University of La Rochelle	CRESCO Dinard and University of La Rochelle	CRESCO Dinard and University of La Rochelle	CRESCO Dinard and University of La Rochelle	University of La Rochelle

DETERMINATION OF THE SIZE STRUCTURE OF THE PRIMARY PRODUCTION USING ANALYSIS OF CHLOROPHYLL A BY SIZE CLASS

Total chlorophyll: for each depth (100m, max fluo, 50m and 5m)

- 1) Filtration of 2L of seawater on nylon net 200 μm . Recuperation of filtrated water.
- 2) Filtration on GF/F (water pump). Fins were used to recuperate the filter which does not be manipulated by fingers to prevent any contamination.
- 3) Filter was placed in an Eppendorf and kept frozen at $-20\text{ }^{\circ}\text{C}$.

Chlorophyll per size class: fraction 5-20 μm / fraction $< 5\mu\text{m}$, for each depth

- 1) Filtration of 2L of water on nylon net 20 μm . Recuperation of filtrated water.
- 2) Filtration on 5 μm filter (Erlen + water pump). Fins were used to recuperate the filter which does not be touched by fingers. Recuperation of filtrated water.
- 3) Filtration on GF/F (water pump). Use fins to recuperate the filter.
- 4) Filter was placed in an Eppendorf and kept frozen at $-20\text{ }^{\circ}\text{C}$.

The frozen filters will be analyzed at the University of La Rochelle. The concentration of chlorophyll by size class ($> 20 \mu\text{m}$, $5-20 \mu\text{m}$, $< 5 \mu\text{m}$) will be expressed by μg chlorophyll *a*/L. It will be possible to convert the chlorophyll *a* in carbon biomass (by a conversion factor found in the literature).

DESCRIPTION AND QUANTIFICATION THE MICROPLANKTONIC COMMUNITY (40-300 μm) USING FLOWCAM TECHNOLOGY

- 1) For each depth, a NISKIN bottle (12L) has been filtrated on a nylon net of $63 \mu\text{m}$.
- 2) Recuperation of filtrated materials on the net with $0.2 \mu\text{m}$ filtrated seawater. It has been placed in a vial with 25 ml of $0.2 \mu\text{m}$ filtered seawater. At the beginning of the cruise, until station 18, the flowcam was not operational. Then, samples were fixed by formalin (5% final concentration). The fixed samples will be analyzed in the University of La Rochelle. From the station 25, the samples were immediately analyzed *in vivo* by flowcam. In addition, samples were recuperated and fixed by formalin (5% final concentration) for conservation in the laboratory.

The determination, concentration, biomass, size spectral of organisms and detritic particles between 40 and $300 \mu\text{m}$ will be determined in the University of La Rochelle.

DETERMINATION OF THE STRUCTURE OF FOOD WEB WITH SAMPLES OF WATERS (2 SIZE CLASSES) FROM NISKIN BOTTLES, MESOZOOPLANKTON AND FISHES FROM IKMT TRAWLS

Estimation of isotopic ratios of water for each depth

This action is complementary to the work of Raymonde Lecomte.

- 1) Filtration of 400 ml water on nylon net $200 \mu\text{m}$. Recuperation of filtrated water.
- 2) Filtration of 180 ml of water on burned GF/F (3 x 60 ml with a syringe). Fins were used to recuperate the filter which does not be touched by fingers to prevent any contamination.
- 3) Filter was put in an Eppendorf and kept frozen at -20°C . It was been twice conducted for a replicate.
- 4) Filtration of 400ml on $5 \mu\text{m}$ filter (Erlen + water pump).
- 5) Filtration of 180 ml of water on burned GF/F (3 x 60 ml with a syringe). Fins were used to recuperate the filter which does not be touched by fingers.
- 6) Filter was put in an Eppendorf and kept frozen at -20°C . It has been twice conducted for a replicate.

The isotopic ratio (C and N) will be determined for these size classes of particles in the water. The work will be made in the University of La Rochelle.

Estimation of isotopic ratios of mesozooplankton, larvae fishes and small fishes

Mesozooplankton was collected from IKMT trawls and sampled in order to perform stable isotopes (C and N) analysis. This action is complementary to the work of Raymonde Lecomte. Fishes were selected by Tony Robinet and Alexandre Carpentier. Organisms were selected in different group for the mesozooplankton and 5 common groups for the fishes and preserved in absolute ethanol (see Table 4).

The samples are taken by Alexandre Carpentier and Eric Feunteun to prepare the analysis of stable isotopes. These analyses will be made in the CRESCO Dinard and in the University of La Roche Ile.

Table 4: Number of mesozooplankton organisms and fish specimens sampled by IKMT trawls for analysis of stable isotopes

Type	Number per station where sampling water
Copepods	30 specimens
Isopodes	50 specimens
Ostracodes	50 specimens
Small and large shrimps	depend of the size, 3 to 10 specimens
Mysis of crustacean	30 specimens
Larvae of crustacean	30 specimens
Chaetognathes	depend of the size, 3 to 20 specimens
Siphonophora	10 specimens
Jellyfish	10 specimens
Larvae fishes	74 specimens
Small fishes	20 specimens

PROKARYOTE DIVERSITY

Prokaryote diversity: for each depth

1) Filtration of 1 to 3L of water on 0.2µm filter (water pump). Fins were used to recuperate the filter which does not be touched by fingers to prevent ant contamination.

2) filter has been stored in an Eppendorf and kept frozen at -20°C.

These analyses will be made at the University of La Rochelle.

DATA OF THE CRUI SE NECESSARY :

- Profile of CTD and Chelsea fluorimeter
- Depth of the Niskin bottles water sampling
- Diversity, distribution and abundance of Leptocephale larvae and other groups of mesozooplankton (copepods, jellyfishes, chaetognathes, crustacean larvae,

fishes) of IKMT trawls.

TOTAL DOC Analysis.

In addition to the previous, total DOM was sampled in the following way. Samples are processed by Raymonde Lecomte on board, and taken to Perpignan University for isotope analysis.

- collect 2 litres of water at various depths (surface and at the fluorescence max)

- filter this water on GF-F Whatman filters

- dry the filters in oven during 12 hours

- preserve the filters (at obscurity in Aluminium paper) at 4°C

and then, back to laboratory to measure the C and N isotope ratios with spectrometry (GV Isoprime spectrometer at University of Perpignan in col. with P. Kerherve)

Each sample was done at station

1, 3, 6, 10, 13, 15, 18, 21, 22, 25, 27, 30, 33, 38

Gobiidae larvae dispersion

Celine Ellien, Shun Watanabe, Stephane Pous, & Katsumi Tsukamoto

The main purpose of our participation in the KH-09-5 Leg 5 cruise on the R/V Hakuho Maru, was to improve our knowledge about the marine larval stage of amphidromous Gobiidae, and particularly on the Gobiidae larvae of the family Sicydiinae, for which little is known.

Two aspects will be particularly studied, based on the samples obtained during this cruise:

- 1) The location and abundance of the larvae in the southwestern Indian Ocean (SWIO)
- 2) The larval life history traits, with a particular attention given to their feeding behaviour

I. Location and abundance of Sicydiinae larvae in the SWIO

During the sorting activity on board, a particular attention was given to the Sicydiinae larvae, and we attempted to identify them based on morphological criteria such as pigmentation, shape of the eye, and shape of the dorsal fin. The shape of the pelvic fin was more difficult to assess at this preliminary step of observation, even if it is the most determinant characteristic at the adult stage for the Gobiidae.

According to these criteria, some larvae have been identified as Sicydiinae larvae, but this has to be confirmed by genetics, once back in the laboratory. Note that sequence primers of Clara Lord (MNHN, Paris) will allow us to confirm or reject the identification of the sampled individuals as being of the Sicydiinae.

All the sampled individuals have been frozen (at a temperature of -80°C) for further analysis.

Table 1. Locations, abundances and total lengths (TL) of the Sicydiinae larvae sampled during the cruise, associated with their reference numbers. An indication of the corresponding photograph number is given when it exists.

Station number	Abundance	Reference number	TL (mm)	Photo number
12	17			
		KH 09-5 St12_1	9	
		KH 09-5 St12_2	9	
		KH 09-5 St12_3	23	
		KH 09-5 St12_4	10	St12-001
		KH 09-5 St12_5	9	St12-004 →St12-005
		KH 09-5 St12_6	9	
		KH 09-5 St12_7	13	St12-006 →St12-007
		KH 09-5 St12_8	9.5	
		KH 09-5 St12_9	11	

		KH 09-5 St12_10	19	St12-008 →St12-009
		KH 09-5 St12_11	11	St12-010 →St12-011
		KH 09-5 St12_12	17	St12-012 →St12-013
		KH 09-5 St12_13	14	
		KH 09-5 St12_14	20.5	
		KH 09-5 St12_15	18	
		KH 09-5 St12_16	17	St12-014 →St12-015
		KH 09-5 St12_17	27	St12-016 →St12-019
15	1			
		KH 09-5 St15_1	13.5	St15-001 →St12-007
19	1			
		KH 09-5 St19_1	10	St19-045 →St19-113
23	7			
		KH 09-5 St23_1	14	
		KH 09-5 St23_2	13	
		KH 09-5 St23_3	15	
		KH 09-5 St23_4	14.5	
		KH 09-5 St23_5	16	
		KH 09-5 St23_6	18	
		KH 09-5 St23_7	15	
34	3			
		KH 09-5 St34_1	10	
		KH 09-5 St34_2	11	St34-001 →St34-002
		KH 09-5 St34_3	9.5	St34-003 →St34-006
35	2			
		KH 09-5 St35_1	17	
		KH 09-5 St35_2	15	St35-001 →St34-006
37	5			
		KH 09-5 St37_1	25	St37-061 →St37-064
		KH 09-5 St37_2	20.5	
		KH 09-5 St37_3	10	
		KH 09-5 St37_4	11	
		KH 09-5 St37_5	8	
39	2			
		KH 09-5 St39_1	16	
		KH 09-5 St39_2	12	

Note that for station 37, the individuals referred to as KH 09-5 St37_1 and KH 09-5 St37_2 have a pelvic fin characteristics that look like those of Eleotridae.

A map will summarise the obtained data of the locations and abundances of the sampled larvae, once their identification is confirmed.

Photos will be recorded in the database of Fishbase, if the larvae are proved to belong to the Sicydiinae and the best looking individual will be used for the National Museum of Natural History (Paris) collections.

II. Study of the larval biology

Once in the lab in Paris, the otoliths will be extracted to assess the larvae age, and a correlation and a regression model between total length and age will be made, if the final number of individuals is sufficient (after confirmation of their identification by genetic analysis).

Most importantly, their feeding behaviour will be analysed through fatty acid analysis. For that purpose, individuals will be analysed through the standard protocol of fatty acid extraction and gas chromatography.

During the cruise, water was sampled at the depth of the maximum fluorescence, which is the depth where most of the planktonic organisms (both phyto- and zooplankton) can be found. The water has been filtered on 0.7 μm mesh size GF/F filters in order to retain the smaller size organisms such as phytoplankton, dinoflagellates etc.

The obtained filters have been frozen (at a temperature of -80°C), and C. Elliene will process fatty acids analysis on them, once back in Paris. The objective is to infer the fatty acids profiles of the larvae with the fatty acid profile of their environment, to have some indication of what the larvae feed on. The fatty acid analyses give information on the diet of the organisms for the 2 to 3 weeks before their capture, as fatty acids are metabolised and then stored in muscles or organs after assimilation.

As the filters didn't retain mesoscale organisms, a spoon of "soup" was also kept (i.e. what we collected in the IKMT net, after the sorting out of fish larvae, gelatinous plankton...) at some stations (i.e. St. 23, St. 37...) where Gobiidae larvae were found, and was frozen, for fatty acid analyses later. This may provide complementary fatty acid profiles, and complete the data on filtered water, to have full concordance with the fatty acid profiles of larvae.

As the feeding behaviour of the larvae depends on the size of their mouth, this parameter has been photographed in detail, to make inferences with plankton size. It is hoped that this and the other types of data collected during the cruise will allow the diet of Sicydiinae larvae to be determined.

Report on the Parasitofauna of Mesopelagic Fishes

Betty Faliex and Elsa Amilhat

The study of biodiversity and its preservation has been a growing concern in the scientific community for about fifteen years. In this context, the study of the biodiversity of parasites from mesopelagic fishes is of particular interest, as according to several authors, it is largely underestimated. Mesopelagic fish parasites have been often neglected, although they represent an integral part of the mesopelagic environment and form an essential part of marine biodiversity.

The objective of this work is to give a first overview of the biodiversity of mesopelagic fish parasites in the southwestern Indian Ocean.

During the 2010 Leg 5 cruise of KH-09-5 on Hakuho Maru, we dissected 72 fish (juveniles and adults) belonging to at least 11 groups (Table 1). For each fish, gills, digestive tract and body cavity were investigated for macroparasites. A total of 41 parasite specimens were recorded that belong to 4 groups: Nematoda (8), Monogenean (2), Digenean (7), Cestoda (11) and 6 types of parasites that are still unidentified (24). Nematodes were preserved in ethanol, Monogeneans and Digeneans were fixed in Bouin solution and Cestoda larvae preserved in 10% formalin. Photographs from both fish and the parasites have been taken.

All the otoliths of the dissected fish were collected. Although, because of the small number of parasitized fish, we may not be able to link otolith shape with parasitism levels. However, we will analyse the shapes of the otoliths collected and photos of the otoliths will be taken with SEM to document otolith shape biodiversity of mesopelagic fish of the southwestern Indian Ocean.

Even if leptocephali could not be dissected for parasite research, we consider that their observation under the stereomicroscope would have detected any encysted parasites larvae. Thus, three unidentified external parasites have been found on 3 different leptocephali belonging to the families Serrivomeridae, Congridae and Ophichthidae. The parasites were preserved in alcohol. The serrivomerid leptocephalus was preserved in formalin for morphological identification of the parasite.

In addition, we found digenean parasites in the body cavity of a chaetognath. These parasites were preserved both in ethanol and formalin for morpho-anatomical examination.

All the parasites will be brought back to Perpignan for further identification.

Table 1. Details of the fishes and chaetognaths investigated for the presence of parasites during KH-09-5 Leg 5 to the west of the Mascarene Ridge in the southwestern Indian Ocean.

Fish group	No. of fish studied (standard length in cm)	% of infected fish	Parasites	Stations
<i>Brama myersi</i>	2 (3.8-4.7)	0	0	14,22
Carangidae	8 (6 -8.5)	12.5	1 monogène Polyopisthocotylea adult 17 unidentified parasites	27,31
<i>Chauliodus</i> sp. (<i>macouni</i> ?)	2 (6.2-14.6)	0	0	20,37
Chiasmodontidae (<i>Pseudoscopelus scriptus sagamianus</i>)	1 (5.7)	0	0	28
Gonostomatidae	5 (6-18)	20	1 cestode larvae 1 nematode larvae	24,26
Melanostomidae	7 (5.3-24.7)	28.6	2 digenea adults 1 cestoda larva 1 cestoda adult diphyllidea?	3,19,23,30,37,38
Myctophiformes	38 (2.4-7)	39.5	4 digenea adults, 1 undet. Parasite, 7 cestoda larvae, 6 nematode larvae and 1 monogenea	4,7,8,11,12,16 17,19,23,24,28,31 32,34,35
Saccopharyngidae	1 (2.4)		0	
Undetermined fish N°1 and N°2	2		1 cestode larvae on N°1	21,35
Undet. fish larvae	3		3 undetermined external parasite 1 digenean metacercaria encysted in the flesh	23,34
Leptocephali (Serrivomeridae, Congridae, Ophichthidae)	3	0.005	1 undet. external parasites N°1 and 2 undet. external parasites N°2	3,37,38
TOTAL	72		41	
Chaetognathe	About 100	Around 6%	6 digenea adults	35,36,38

Diversity of fish larvae west of the Mascarene Ridge in the Southwestern Indian Ocean

Tony Robinet, Mari Kuroki, Alex Carpentier, Michael J. Miller,
Katsumi Tsukamoto, and Tsuguo OTAKE

During the KH-06-4 sampling survey in the same region as the present cruise, the diversity of leptocephali larvae of Anguilliformes was studied in the western Indian Ocean by describing the structure of the species assemblage of more than 500 leptocephalus larvae, but this analysis was restricted to anguilliform larvae.

The species identification of most of the other fish larvae in the ocean offshore of Madagascar and to the west of the Mascarene Ridge is difficult due to the lack of taxonomic characters in the early life stages of many fishes. However, species identification is now possible as a routine technology by DNA sequencing (barcoding: COI etc.) in the laboratory.

During the present cruise all the other fish larvae (non-anguilliforms) were therefore systematically sorted out of the plankton of each tow (step and oblique tows). High resolution digital pictures were taken of some specimens with stereo microscopes aboard and these were kept individually in 99% ethanol (for future DNA sequencing). When more than one of a species was collected, specimens were preserved in 10% formalin (for morphological description). In other cases specimens were presorted and preserved in ethanol.

The number of larvae collected (excluding anguilliform leptocephali) is estimated to be up to 5,000, among which 500 were pictured and preserved individually. All of the larvae will be brought back to the University Museum of the University of Tokyo (Dr. Kuroki) and then shared with the National Museum of Natural History in Paris (Dr. Robinet), to be identified to the lowest taxonomic level possible.

The objectives of this study are:

- 1) To supply collection material of fish larvae from this particular region to the museums in France and Japan.
- 2) To analyze the variability of species assemblage according to sampling depth, latitude, and night/day time.
- 3) To provide information on unidentified small fish larvae using DNA sequences and to link the successive development stages in the poorly known groups of mesopelagic fishes.

Moreover, the expected species composition of the fish larvae collected during KH-09-5 will provide specimens to other ongoing research in fish science (tunas, swordfish, Gonostomatidae etc.). The photographs shared between France and Japan from this cruise can also eventually be used to update photographic databases at the museums or other organizations.

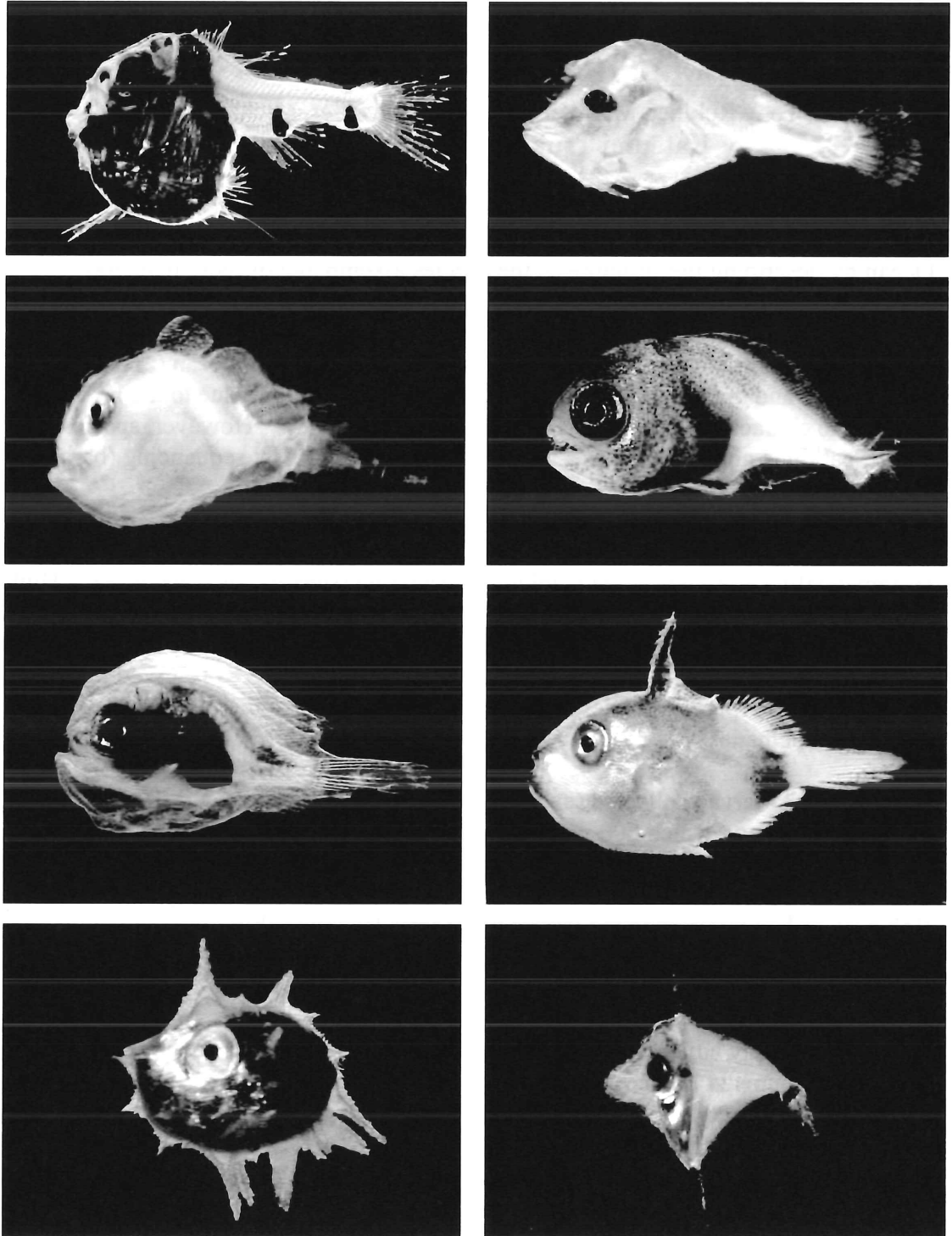


Figure 1. Various fish larvae collected with the IKMT during the KH-09-5 sampling survey west of the Mascarene Ridge, including a hatchetfish, anglerfish, triggerfish, and an ocean sunfish/moonfish/mambo (e.g. *Mola mola*) in the bottom left panel.

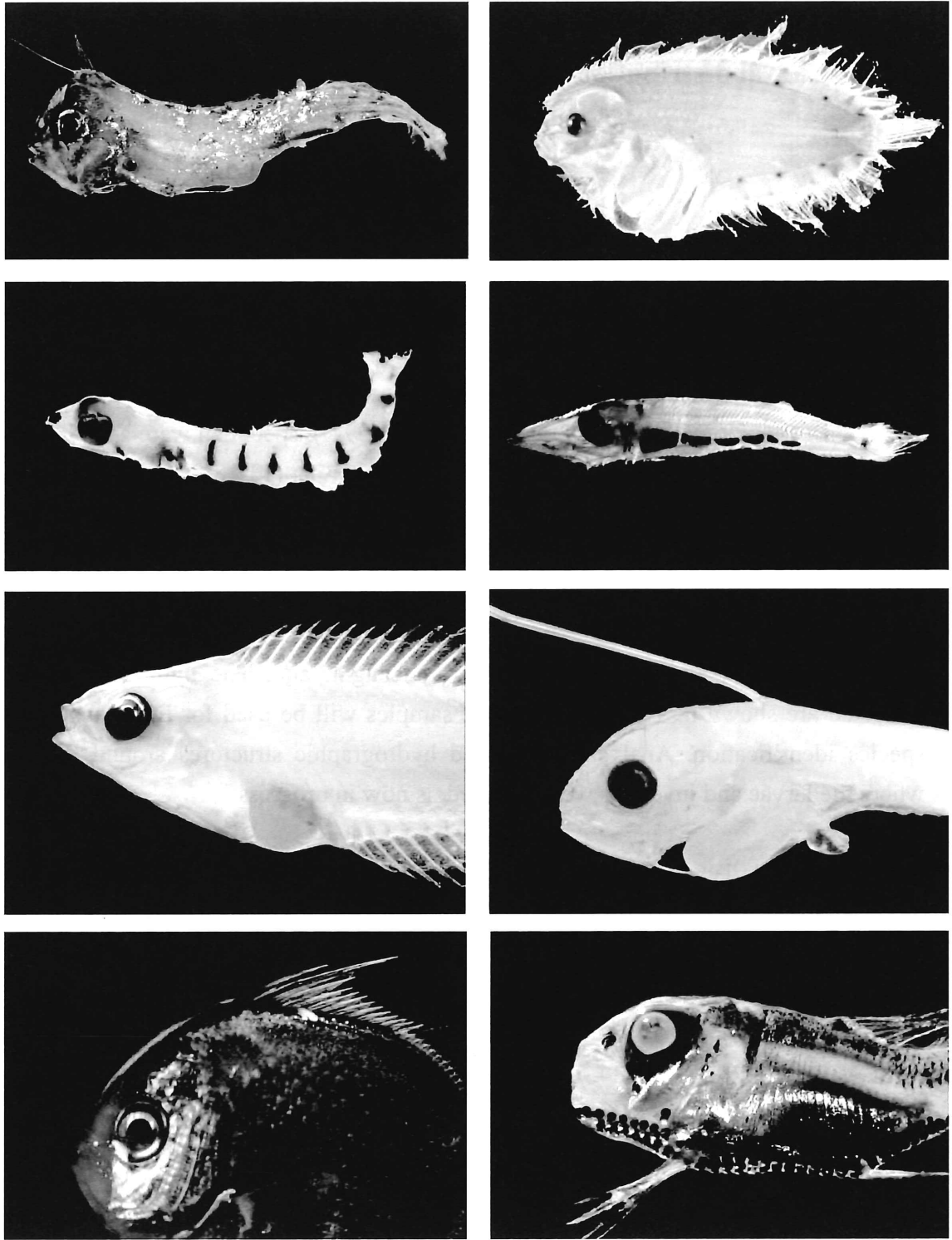


Figure 2. Various other fish larvae collected with the IKMT during the KH-09-5 sampling survey west of the Mascarene Ridge, including a labrid, flatfish, etc.

Larval distribution and population structure of *Thunnus* in the Indian Ocean

Takashi Kitagawa

Scombridae contains 15 genera including *Thunnus*. The genus *Thunnus* contains eight species. In the Indian Ocean, yellowfin (*T. albacares*), bigeye (*T. obesus*), southern bluefin tuna (*T. maccoyii*) and albacore (*T. alalunga*) are distributed, but larval distributions and population structures of those species are not still unknown.

During 29 January- 9 February 2010 (KH-09-5), specimen were sampled at 40 stations in the area northern part of the EEZ of Mauritius (13-18°S, 54-60°E) with the Issacs Kidd Midwater Trawl (IKMT) with an 8.7 m² mouth opening and 0.5 mm mesh. Oblique and step tows were carried out during the day and night, respectively. CTD (+CMS) observation was also conducted. Details of the net tows and CTD casting are shown somewhere in this cruise report.

Tuna like larvae and juveniles were caught, and. The number of samples collected are shown in Table 1. All of the samples will be used for DNA analysis for species identification. Analysis of detailed hydrographic structures around the areas where the larvae and juveniles were sampled, is now in progress.

Table 1: Tuna samples collected during 29 Jan- 9 Feb 2010 (KH-09-5),

St.1	1	St.2	12	St.3	6	St.7	10
St.8	4	St.11	4	St.13	2	St.15	9
St.17	2	St.19	16	St.23	2	St.24	3
St.25	2	St.27	12	St.28	11	St.30	1
St.31	27	St.34	6	St.37	18		

Studies on feeding ecology of phyllosoma larvae of palinurid and scyllarid lobsters

Masamichi MACHIDA and Shuhei NISHIDA

Introduction

The larvae of palinurid and scyllarid lobsters are called “phyllosoma”, with their transparent and flattened bodies. Phyllosoma larvae are distributed in open ocean, with long planktonic period for over 6 months. Their prey items in natural environment have been mostly unknown because their low natural density has made wild specimens difficult to be captured for experimental observations. The main purpose of the present cruise is to collect phyllosoma larvae for analyses of stable isotope ratios, fatty acid composition and gut contents, and for species identification using genetic markers.

Phyllosoma, plankton and POM sampling

A total of 28 phyllosomas was collected using an Isaacs-Kidd Midwater Trawl (IKMT, 0.5mm mesh aperture) towed obliquely, horizontally or step-wise during Legs 5 and 6 (Table 1). Phyllosomas were sorted from the original zooplankton samples immediately after capture. These phyllosomas were fixed or frozen for the respective analyses as described below.

Sea-water samples were collected from 0, 50, 100, 200 m and the depths of chlorophyll maximum, and POM (particulate organic matter) was filtered on pre-combusted GF/F filters which were then preserved at -80°C.

Food habitats of phyllosoma

Studies of mouthpart and gut morphology of phyllosoma larvae have suggested that they are suited for feeding on soft foods such as gelatinous zooplankton (Nishida et al., 1990; Johnston and Ritar, 2001). DNA analyses of gut contents have identified Cnidaria and Urochordata (Suzuki et al., 2006) as potential food sources. On the basis of these bodies of information, we collected gelatinous zooplankton samples (Cnidaria, Ctenophore, Chaetognatha, Heteropoda, Thecosomata, Gymnosomata, Thaliacea, Salpida), net zooplankton mixture (random sampling) and POM for analyses of stable isotope ratios and fatty-acid composition to estimate trophic levels of phyllosoma. These samples and sorted phyllosomas were identified to genera, families, or higher taxonomic groups and frozen at -80°C for the analyses on land. Some other phyllosomas were frozen or fixed in 100% alcohol for genetic identification of gut-contents, or fixed

in 2% formaldehyde or in Karnovsky's fixative for light/electron microscopic observation on feeding structure and gut contents.

Species identification of phyllosoma with genetic markers

It is difficult to identify the species of scyllarid phyllosomas, due to the limited morphological information that are available only for the final stage larvae in most species. Identification at the genus level is also difficult, particularly between *Parribacus* and *Scyllarides*, by morphological characteristics. Genetic species identification of invertebrate is often possible on the basis of mitochondrial cytochrome oxidase subunit I (COI) and/or 16S rDNA genes, coupled with a GenBank homology search. However, COI and 16S rDNA references of scyllarid lobsters are still very incomplete. With this circumstance, we aimed at obtaining sequence references and morphological characteristics of the scyllarid lobsters. Some of the sorted scyllarid phyllosomas were identified to possible lowest levels and preserved in 100% alcohol for genetic analysis, while some other specimens were fixed and preserved in formaldehyde as references for morphological analysis.

Table 1. Number of phyllosoma larvae collected during Legs 5 and 6 of KH-09-5 cruise.

	KH-09-5, Leg5	Leg.6	Total
<i>Panulirus</i> spp.	4	15	19
Scyllarinae spp.	1	1	2
others	2	5	7
Total	7	21	28

Geographic Variation in the Assemblage Structure and Life Histories of Leptocephali across the Indian Ocean Basin

Michael J. Miller, Jun Aoyama, Shun Watanabe, Eric Feunteun, Sam Wouthuyzen, Augy Syahailatua, Tsuguo Otake, Katsumi Tsukamoto and various other cruise members

During Leg 6 of the KH-09-5 cruise in 2010, which was scheduled primarily as a transit cruise to cross the Indian Ocean from Mauritius to Benoa of Bali Indonesia, 12 IKMT stations were also made during nighttime (Figure 1). These tows collected more than 1,100 leptocephali and consisted primarily of mesopelagic eel leptocephali from Stn. 1 to Stn. 10, and then were dominated by the leptocephali of shallow water eels at Stn. 11 and 12. Multiple tows were made at Stn. 12 where many ophichthids, congrid and moringuids were collected.

These collections made during Leg 6 of KH-09-5 will be combined with the transect of 20 stations that was made in 2006 during KH-06-4 across the Indian Ocean (Figure 1) along with the sampling that was conducted to the west of the Mascarene Ridge in both years also, to gain understanding of the dynamics of marine eel life histories in this semi-enclosed ocean basin in relation to ocean current patterns and the temperature structure of the surface layer where leptocephali live and feed.

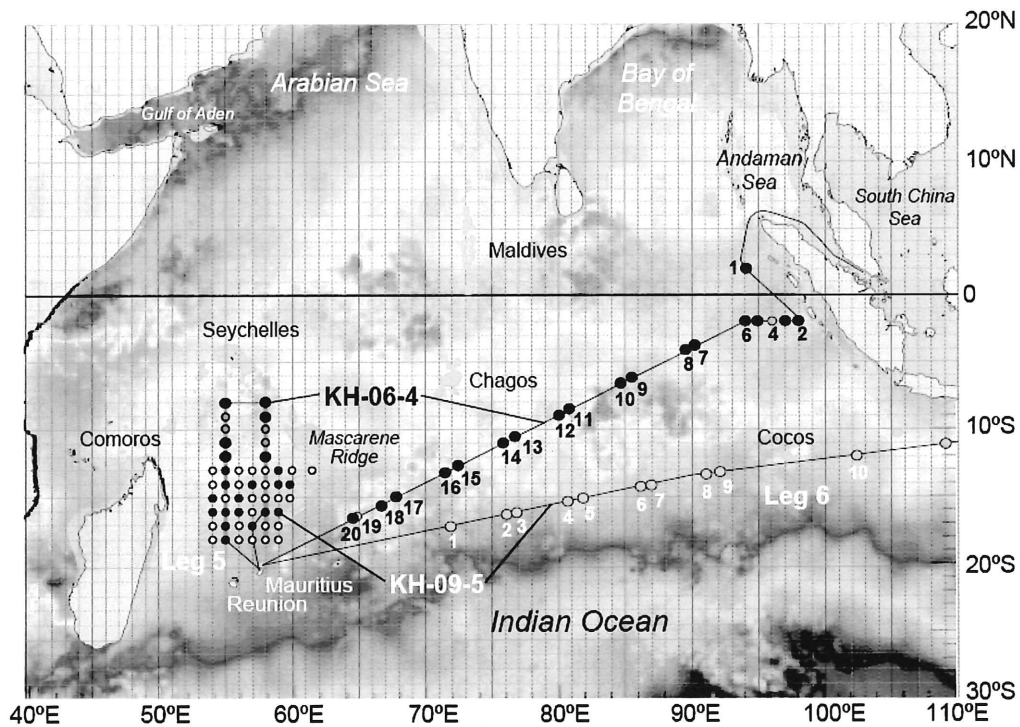


Figure 1. Map showing the sampling stations of the KH-06-4 and KH-09-5 sampling surveys for leptocephali. White and red circles in the Mascarene Ridge region show day tows, but most other stations are night tows in the rest of the map. Red lines show the transects sampled west of the Mascarene Ridge in 2006, with half of the stations overlapping with the 2010 sampling grid. Stn. 12 of KH-09-5 Leg 6 is just off the map. The station map is plotted over sea surface temperature for 14 January 2010 a few weeks before the start of Leg 5 of KH-09-5, obtained from the US Navy NRL.

KH-09-5 Net Record Leg 5

St.	Location		Date	Time		Net Type	Mesh size (mm)	Towing Method	Wire out (m)	Sampl. layer (m)	Reel speed (m/s)	Ship speed (kt)	Sea Depth (m)
	Net in	Net out		Net in	Net out								
1	S 18-00.1 E 54-01.6	S 17-57.4 E 54-01.2	100130	07:12	08:37	IKMT	0.5	Obl.	1542	0-527	1.0-0.5	2.5-2.0	4654-4782
2	S 17-00.4 E 54-02.4	S 16-59.0 E 54-03.0	100130	13:50	14:55	IKMT	0.5	Obl.	1185	0-513	1.0-0.5	2.5-2.0	4708-4725
3	S 15-59.8 E 54-02.1	S 15-56.0 E 54-01.8	100130	20:02	21:33	IKMT	0.5	Step	534	0-155	1.0-0.5	2.5-2.0	4516-4566
4	S 14-59.8 E 54-01.8	S 14-56.7 E 54-01.2	100131	2:15	3:43	IKMT	0.5	Step	585	0-160	1.0-0.3	2.5-2.0	4500-4592
5	S 14-00.3 E 54-02.3	S 13-58.4 E 54-02.1	100131	8:39	9:47	IKMT	0.5	Obl.	1265	0-521	1.0-0.5	2.5-2.0	4216-4264
6	S 13-00.2 E 54-02.0	S 12-57.4 E 54-02.4	100131	14:52	16:13	IKMT	0.5	Obl.	1506	0-618	1.0-0.5	2.5-2.0	4605-4628
7	S 12-59.8 E 55-00.0	S 12-58.4 E 54-56.5	100131	20:16	21:47	IKMT	0.5	Step	344	0-153	1.0-0.3	3.0-1.5	3878-4215
8	S 13-59.9 E 54-59.9	S 13-58.2 E 54-57.3	100201	2:16	3:55	IKMT	0.5	Step	554	0-157	1.0-0.3	2.5-1.0	4481-4516
9	S 14-59.8 E 55-00.1	S 14-55.8 E 54-59.4	100201	8:25	9:59	IKMT	0.5	Obl.	1738	0-523	1.0-0.5	2.7-1.5	4266-4543
10	S 16-00.2 E 54-59.8	S 16-58.7 E 54-57.0	100202	15:20	16:40	IKMT	0.5	Obl.	1433	0-560	1.0-0.5	2.5-2.0	4617
11	S 17-00.3 E 54-59.9	S 17-04.7 E 54-59.9	100201	20:59	22:37	IKMT	0.5	Step	639	0-160	1.0-0.3	2.6-1.9	4218-4922
12	S 18-00.2 E 55-00.1	S 17-58.6 E 55-02.8	100202	2:32	4:06	IKMT	0.5	Step	680	0-155	1.0-0.3	2.5-2.0	4586-4590
13	S 17-59.8 E 55-59.4	S 17-56.9 E 56-00.3	100202	9:08	10:29	IKMT	0.5	Obl.	1500	-	1.0-0.5	2.3-1.5	4555-4574
14	S 16-59.8 E 56-00.1	S 16-56.3 E 56-00.4	100202	15:09	16:41	IKMT	0.5	Obl.	1500	-	1.0-0.5	2.5-2.0	4444-4499
15	S 15-59.9 E 55-59.4	S 15-56.6 E 55-59.4	100202	21:28	23:00	IKMT	0.5	Step	474	0-154	1.0-0.3	2.6-1.5	4365-4537
16	S 14-59.7 E 56-00.0	S 14-56.3 E 55-59.6	100203	3:03	4:30	IKMT	0.5	Step	550	0-158	1.0-0.3	2.5-2.0	4210-4335
17	S 14-00.3 E 56-00.1	S 13-57.7 E 56-00.5	100203	9:56	11:11	IKMT	0.5	Obl.	1440	0-599	1.0-0.5	2.5-1.5	4400-4425
18	S 13-00.3 E 55-59.5	S 12-59.7 E 55-57.1	100203	16:06	17:14	IKMT	0.5	Obl.	1275	0-503	1.0-0.5	2.5-2.0	4334-4357
19	S 13-00.1 E 57-00.1	S 13-02.4 E 57-02.9	100203	21:33	23:14	IKMT	0.5	Step	574	0-153	1.0-0.3	2.6-2.0	4343-4350
20	S 13-59.9 E 56-59.9	S 13-57.5 E 56-58.1	100204	3:20	4:45	IKMT	0.5	Step	505	0-155	1.0-0.3	2.5-1.5	4315-4323
21	S 15-00.4 E 56-59.9	S 14-58.0 E 56-57.7	100204	10:12	11:34	IKMT	0.5	Obl.	1534	0-545	1.0-0.5	2.6-1.5	4313-4334

KH-09-5 Net Record Leg 5

St.	Location		Date	Time		Net Type	Mesh size (mm)	Towing Method	Wire out (m)	Sampl. layer (m)	Reel. speed (m/s)	Ship speed (kt)	Sea Depth (m)
	Net in	Net out		Net in	Net out								
22	S 16-00.6 E 56-59.8	S 15-58.4 E 56-58.0	100204	16:49	18:04	IKMT	0.5	Obl.	1345	0-500	1.0-0.5	2.6-1.3	4332-4345
23	S 17-00.0 E 57-00.0	S 16-56.7 E 56-58.6	100204	22:27	23:55	IKMT	0.5	Step	461	0-153	1.0-0.3	2.7-1.1	3462-4247
24	S 18-00.0 E 56-59.8	S 17-57.9 E 56-58.0	100205	4:31	5:37	IKMT	0.5	Obl.	1205	0-503	1.0-0.5	2.5-1.3	4190
25	S 17-59.6 E 57-59.7	S 17-55.4 E 57-59.0	100205	10:57	12:25	IKMT	0.5	Obl.	1718	0-507	1.0-0.5	2.5-2.1	3986-4032
26	S 17-00.1 E 58-00.0	S 16-58.7 E 57-57.3	100205	17:05	18:23	IKMT	0.5	Obl.	1500	-	1.0-0.5	2.0-1.5	3493-3591
27	S 16-00.3 E 58-00.5	S 15-58.3 E 57-59.5	100205	23:29	0:51	IKMT	0.5	Step	288	0-151	1.0-0.3	2.6-2.0	4075-4088
28	S 14-59.6 E 57-59.8	S 14-56.6 E 57-58.6	100206	5:43	7:02	IKMT	0.5	Obl.	1541	0-534	1.0-0.5	2.3-1.2	4165-4186
29	S 13-59.9 E 58-00.2	S 13-58.3 E 57-58.6	100206	11:36	12:42	IKMT	0.5	Obl.	1289	0-507	1.0-0.5	2.9-2.0	4142-4166
30	S 13-00.1 E 57-59.6	S 13-00.9 E 57-56.3	100206	17:28	18:54	IKMT	0.5	Obl.	1706	0-578	1.0-0.5	2.3-1.5	4282-4285
31	S 12-59.9 E 58-59.8	S 13-01.5 E 59-00.7	100206	23:25	0:44	IKMT	0.5	Step	378	0-162	1.0-0.3	2.9-2.3	4019-4022
32	S 13-00.2 E 60-00.0	S 13-03.2 E 60-01.0	100207	5:03	6:20	IKMT	0.5	Obl.	1500	-	1.0-0.5	2.3-1.6	3620-3682
33	S 12-59.0 E 61-29.7	S 12-57.1 E 61-31.2	100207	13:23	14:44	IKMT	0.5	obl.	1500	-	1.0-0.5	2.0-2.5	2748-2852
34	S 14-00.2 E 59-50.0	S 14-03.8 E 59-51.8	100207	22:36	00:10	IKMT	0.5	Step	524	0-156	1.0-0.5	2.1-2.7	3924-3925
35	S 13-59.9 E 58-59.9	S 14-02.7 E 59-02.1	100208	03:59	05:26	IKMT	0.5	Step	620	0-156	1.0	1.5-2.5	4008-4021
36	S 15-00.4 E 59-50.0	S 15-03.2 E 59-51.1	100208	14:25	15:44	IKMT	0.5	Obl.	0-557	0-557	0.5-1.0	1.9-2.0	707-1053
37	S 15-00.1 E 58-59.9	S 15-00.8 E 59-01.8	100208	19:19	20:25	IKMT	0.5	Obl.	1189	0-503	1.0	1.5-2.5	3942-3949
38	S 16-00.5 E 58-59.6	S 16-01.8 E 59-02.2	100209	1:18	2:45	IKMT	0.5	Step	520	0-163	0.3-1.0	2.0-2.5	1792-2126
39	S 17-00.1 E 58-59.2	S 17-00.0 E 59-01.7	100209	6:48	7:58	IKMT	0.5	Obl.	1342	0-501	0.5-1.0	1.5-2.5	1076-1512
40	S 18-00.0 E 58-59.4	S 18-02.6 E 59-00.5	100209	12:58	14:24	IKMT	0.5	Obl.	1583	0-510	0.5-1.0	2.0-2.5	2470-2587

KH-09-5 Net Record Leg 6

St.	Location		Date	Time		Net Type	Mesh size (mm)	Towing Method	Wire out (m)	Sampl. layer (m)	Reel. speed (m/s)	Ship speed (kt)	Sea Depth (m)
	Net in	Net out		Net in	Net out								
1	S 17-10.2 E 71-57.4	S 17-12.6 E 71-57.0	100217	1:15	2:42	IKMT	0.5	Step	469	0-153		1.8-2.4	4546-4864
2	S 16-22.1 E 76-09.1	S 16-24.6 E 76-08.7	100217	20:08	21:36	IKMT	0.5	Step	486	0-161		1.7-2.7	4782-5007
3	S 16-13.5 E 76-57.7	S 16-16.6 E 76-56.1	100218	2:19	3:47	IKMT	0.5	Step	450	0-152		0.9-2.5	4738-4873
4	S 15-28.9 E 80-55.5	S 15-31.7 E 80-53.9	100218	20:16	21:46	IKMT	0.5	Step	491	0-160		2.0-2.6	4948-5116
5	S 15-17.5 E 81-57.1	S 15-20.6 E 81-55.3	100219	2:16	3:41	IKMT	0.5	Step	478	0-152	0.3-1.0	2.3-2.5	4823-5185
6	S 14-30.4 E 86-06.1	S 14-32.4 E 86-04.9	100219	20:06	21:27	IKMT	0.5	Step	337	0-149		1.7-2.7	2549-2683
7	S 14-17.3 E 87-15.7	S 14-18.5 E 87-17.0	100220	2:14	3:36	IKMT	0.5	Step	377	0-153	0.3-1.0	2.0-2.5	2564-2633
8	S 13-33.3 E 91-11.2	S 13-31.0 E 91-09.2	100220	20:22	21:46	IKMT	0.5	Step	434	0-150		1.7-2.7	4918-5266
9	S 13-21.8 E 92-17.0	S 13-21.0 E 92-14.6	100221	2:14	3:38	IKMT	0.5	Step	399	0-150	0.3-1.0	2.3-2.5	4822-5122
10	S 12-08.2 E 102-31.4	S 12-11.0 E 102-32.2	100222	19:31	20:58	IKMT	0.5	Step	460	0-159	0.2	1.6-2.7	3267-4531
11-H	S 11-20.1 E 109-07.8	S 11-21.1 E 109-07.9	100224	0:16	0:52	IKMT	0.5	Hori.	191	0-40	0.2	2.3-2.6	5197-5201
11-S	S 11-21.4 E 109-07.9	S 11-23.6 E 109-08.5	100224	1:06	2:30	IKMT	0.5	Step	462	0-150	0.3-0.5	1.9-2.8	5027-5162
12-H1	S 11-37.9 E 109-13.4	S 11-38.8 E 109-12.6	100224	3:54	4:26	IKMT	0.5	Hori.	127	0-17.9		1.3-2.5	4910-4918
12-H2	S 11-39.1 E 109-12.4	S 11-40.3 E 109-11.2	100224	4:37	5:18	IKMT	0.5	Hori.	173	0-27.4		1.4-2.8	4890-4902
12-H3	S 11-40.5 E 109-10.9	S 11-41.6 E 109-10.0	100224	5:30	6:02	IKMT	0.5	Hori.	169	0-27.8		1.5-2.5	4224-4930

KH-09-5_Leg5		St.1	Depth	4772 m		KH-09-5_Leg5		St.2	Depth	4702 m			
Date:	2010.1.30		Lat.	18 00.04S		Date:	2010.1.30		Lat.	17 00.02S			
Time:	2:13		Long.	54 01.88E		Time:	9:02		Long.	54 02.08E			
CTD data (LAY)	Pres.	Temp.	Sal	DO	FLC	CTD data (LAY)	Pres.	Temp.	Sal	DO	FLC		
	db	°C	(psu)	ml·l ⁻¹	μg/l		db	°C	(psu)	ml·l ⁻¹	μg/l		
	5	29.015	35.060	4.31	0.02		5	29.545	35.052	4.31	0.01		
	10	29.019	35.060	4.32	0.02		10	29.410	35.046	4.31	0.01		
	20	28.980	35.056	4.33	0.02		20	29.285	35.041	4.31	0.01		
	30	28.728	35.045	4.41	0.02		30	28.867	35.066	4.47	0.02		
	40	27.240	34.987	4.66	0.03		40	27.959	35.071	4.59	0.02		
	50	26.645	34.959	4.70	0.03		50	26.962	34.983	4.70	0.03		
	75	25.092	34.892	4.68	0.07		75	25.424	34.880	4.80	0.05		
	100	24.171	34.856	4.61	0.14		100	24.150	34.876	4.62	0.11		
	125	23.439	34.982	4.28	0.17		125	23.318	35.009	4.42	0.24		
	150	22.351	35.185	3.81	0.10		150	22.518	35.216	4.24	0.11		
	175	21.103	35.353	3.56	0.04		175	21.443	35.475	4.09	0.05		
	200	19.909	35.315	3.13	0.03		200	20.540	35.586	4.09	0.03		
	250	17.219	35.347	3.10	0.02		250	18.111	35.432	3.33	0.02		
	300	15.439	35.327	3.38	0.02		300	16.147	35.427	3.66	0.02		
	400	12.803	35.192	4.68	0.01		400	12.532	35.147	4.45	0.01		
500	10.751	34.912	4.87	0.01	500	10.243	34.837	5.00	0.01				
CTD data (BTL)						CTD data (BTL)							
BTL No.	Depth m	Pres. db	Temp. °C	Sal (psu)	DO ml·l ⁻¹	FLC μg/l	BTL No.	Depth m	Pres. db	Temp. °C	Sal (psu)	DO ml·l ⁻¹	FLC μg/l
Sur.	0	***	28.8	***	***	***	Sur.	0	***	29.8	***	***	***
1	198	199	20.001	35.316	3.15	0.03	1						
2	198	199	20.024	35.316	3.14	0.03	2						
3	197	199	20.031	35.318	3.15	0.03	3						
4	110	111	23.716	34.918	4.42	0.18	4						
5	110	111	23.904	34.867	4.50	0.19	5						
6	110	111	23.885	34.873	4.46	0.20	6						
7	109	110	23.821	34.892	4.46	0.19	7						
8	110	110	23.835	34.887	4.49	0.19	8						
9	110	111	23.836	34.886	4.47	0.20	9						
10	111	111	23.867	34.876	4.49	0.19	10						
11	100	101	24.098	34.863	4.54	0.16	11						
12	99	100	24.148	34.862	4.58	0.15	12						
13	99	100	24.171	34.862	4.58	0.15	13						
14	100	101	24.267	34.868	4.62	0.15	14						
15	100	100	24.312	34.860	4.64	0.13	15						
16	100	100	24.305	34.861	4.65	0.12	16						
17	49	49	26.791	34.966	4.70	0.04	17						
18	49	50	26.870	34.970	4.68	0.04	18						
19	49	50	26.803	34.967	4.69	0.03	19						
20	49	49	26.879	34.972	4.68	0.03	20						
21	49	49	26.859	34.971	4.68	0.03	21						
22	49	49	26.848	34.972	4.68	0.04	22						
23	5	5	29.022	35.060	4.32	0.02	23						
24	5	5	29.033	35.062	4.32	0.02	24						

KH-09-5_Leg5		St.5	Depth	4219 m		KH-09-5_Leg5		St.6	Depth	4614 m			
Date:	2010.1.31		Lat.	14 00.20S		Date:	2010.1.31		Lat.	13 00.26S			
Time:	3:56		Long.	54 02.32E		Time:	10:05		Long.	54 02.24E			
CTD data (LAY)	Pres.	Temp.	Sal	DO	FLC	CTD data (LAY)	Pres.	Temp.	Sal	DO	FLC		
	db	°C	(psu)	ml·l ⁻¹	μg/l		db	°C	(psu)	ml·l ⁻¹	μg/l		
	5	29.085	35.042	4.34	0.03		10	28.966	34.985	4.35	0.02		
	10	29.088	35.041	4.35	0.03		20	28.931	34.980	4.34	0.02		
	20	29.086	35.041	4.34	0.03		30	28.903	34.983	4.35	0.03		
	30	28.842	35.050	4.42	0.03		40	26.681	35.022	4.69	0.04		
	40	27.029	35.059	4.67	0.04		50	25.567	34.952	4.70	0.05		
	50	26.345	35.022	4.70	0.05		75	23.267	34.930	4.08	0.24		
	75	24.692	34.897	4.56	0.19		100	20.805	34.975	2.67	0.13		
	100	22.944	34.959	3.85	0.19		125	18.506	35.034	2.27	0.07		
	125	21.301	35.188	3.31	0.13		150	16.964	35.081	2.10	0.04		
	150	19.060	35.108	1.99	0.08		175	16.608	35.280	2.86	0.02		
	175	17.843	35.096	1.81	0.05		200	15.444	35.240	2.95	0.02		
	200	16.096	35.078	1.98	0.03		250	14.161	35.211	3.32	0.02		
	250	14.153	35.136	2.92	0.02		300	12.609	35.058	3.28	0.02		
	300	12.724	35.074	3.23	0.02		400	11.056	34.926	3.56	0.02		
	400	10.604	34.896	3.88	0.02		500	9.920	34.817	3.70	0.02		
500	9.590	34.809	3.40	0.02									
CTD data (BTL)						CTD data (BTL)							
BTL	Depth	Pres.	Temp.	Sal	DO	FLC	BTL	Depth	Pres.	Temp.	Sal	DO	FLC
No.	m	db	°C	(psu)	ml·l ⁻¹	μg/l	No.	m	db	°C	(psu)	ml·l ⁻¹	μg/l
Sur.	0	***	28.9	***	***	***	Sur.	0	***	29.0	***	***	***
1							1	200	201	15.284	35.276	3.21	0.02
2							2	200	202	15.277	35.278	3.23	0.02
3							3	201	203	15.270	35.281	3.24	0.02
4							4	74	75	23.089	34.936	4.04	0.24
5							5	75	75	23.377	34.926	4.23	0.24
6							6	74	75	23.345	34.927	4.22	0.24
7							7	74	75	23.369	34.928	4.23	0.24
8							8	74	75	23.360	34.929	4.23	0.24
9							9	74	75	23.373	34.930	4.24	0.24
10							10	74	75	23.374	34.930	4.24	0.24
11							11	99	100	21.338	34.975	2.94	0.14
12							12	100	100	21.224	34.975	2.91	0.15
13							13	100	100	21.229	34.976	2.81	0.14
14							14	100	101	21.181	34.979	2.74	0.14
15							15	100	101	21.061	34.978	2.75	0.13
16							16	100	101	20.932	34.977	2.78	0.14
17							17	49	50	25.664	34.944	4.67	0.05
18							18	49	50	25.573	34.951	4.67	0.05
19							19	49	50	25.592	34.949	4.68	0.05
20							20	49	50	25.691	34.958	4.69	0.05
21							21	49	49	25.657	34.955	4.70	0.06
22							22	49	50	25.717	34.956	4.69	0.05
23							23	5	5	28.962	34.984	4.35	0.02
24							24	5	5	28.966	34.985	4.34	0.02

KH-09-5_Leg5		St.14	Depth	4493 m		KH-09-5_Leg5		St.15	Depth	4081 m			
Date:	2010.2.2		Lat.	16 59.93S		Date:	2010.2.2		Lat.	16 00.14S			
Time:	10:30		Long.	56 00.11E		Time:	16:27		Long.	55 59.87E			
CTD data (LAY)	Pres.	Temp.	Sal	DO	FLC	CTD data (LAY)	Pres.	Temp.	Sal	DO	FLC		
	db	°C	(psu)	ml·l ⁻¹	μg/l		db	°C	(psu)	ml·l ⁻¹	μg/l		
	5	29.765	35.089	4.31	0.01		5	29.670	34.935	4.33	0.02		
	10	29.471	35.082	4.32	0.01		10	29.497	34.933	4.34	0.02		
	20	29.098	35.055	4.35	0.02		20	28.828	34.977	4.42	0.02		
	30	28.741	35.064	4.40	0.02		30	28.264	35.056	4.56	0.03		
	40	28.101	35.092	4.50	0.03		40	26.694	35.033	4.64	0.03		
	50	27.269	35.080	4.56	0.04		50	26.183	34.994	4.67	0.04		
	75	25.256	34.987	4.64	0.16		75	24.979	34.920	4.75	0.05		
	100	24.158	34.965	4.60	0.26		100	23.623	34.965	4.64	0.18		
	125	22.379	35.069	3.53	0.16		125	22.989	35.069	4.41	0.25		
	150	20.391	35.109	2.65	0.07		150	21.666	35.278	3.73	0.11		
	175	19.506	35.335	3.08	0.03		175	19.918	35.390	3.32	0.03		
	200	18.321	35.538	3.59	0.02		200	18.333	35.360	3.09	0.02		
	250	16.190	35.572	4.32	0.01		250	15.970	35.418	3.70	0.02		
	300	14.240	35.387	4.64	0.01		300	13.851	35.221	3.67	0.02		
	400	11.994	35.093	4.91	0.01		400	10.890	34.931	4.51	0.01		
500	10.488	34.872	5.06	0.01	500	9.443	34.751	4.44	0.01				
CTD data (BTL)						CTD data (BTL)							
BTL No.	Depth m	Pres. db	Temp. °C	Sal (psu)	DO ml·l ⁻¹	FLC μg/l	BTL No.	Depth m	Pres. db	Temp. °C	Sal (psu)	DO ml·l ⁻¹	FLC μg/l
Sur.	0	***	30.0	***	***	***	Sur.	0	***	29.9	***	***	***
1							1	199	200	18.447	35.367	3.11	0.02
2							2	199	200	18.462	35.368	3.10	0.02
3							3	199	201	18.442	35.367	3.12	0.02
4							4	105	105	23.375	35.007	4.52	0.26
5							5	105	105	23.373	35.009	4.52	0.26
6							6	104	105	23.378	35.008	4.53	0.25
7							7	104	105	23.397	35.004	4.55	0.26
8							8	104	105	23.401	35.004	4.53	0.27
9							9	105	105	23.380	35.009	4.53	0.26
10							10	104	105	23.407	35.003	4.53	0.26
11							11	100	100	23.606	34.965	4.63	0.23
12							12	99	100	23.618	34.965	4.63	0.21
13							13	99	100	23.616	34.965	4.64	0.22
14							14	100	100	23.613	34.965	4.63	0.21
15							15	99	100	23.617	34.965	4.64	0.21
16							16	100	100	23.612	34.965	4.64	0.21
17							17	51	51	26.245	34.997	4.67	0.03
18							18	50	51	26.229	34.996	4.67	0.03
19							19	50	51	26.254	34.998	4.67	0.04
20							20	50	51	26.249	34.999	4.67	0.03
21							21	50	51	26.242	34.998	4.67	0.04
22							22	51	51	26.247	34.998	4.66	0.04
23							23	4	4	29.609	34.930	4.33	0.02
24							24	5	5	29.610	34.932	4.33	0.02

KH-09-5_Leg5		St.18	Depth	4329 m	KH-09-5_Leg5	St.21	Depth	4312 m					
Date:	2010.2.3		Lat.	13 00.01S	Date:	2010.2.4		Lat.	15 00.29S				
Time:	11:21		Long.	55 59.93E	Time:	5:15		Long.	56 59.94E				
CTD data (LAY)		Pres.	Temp.	Sal	DO	FLC	CTD data (LAY)		Pres.	Temp.	Sal	DO	FLC
		db	°C	(psu)	ml·l ⁻¹	μg/l			db	°C	(psu)	ml·l ⁻¹	μg/l
		5	29.050	34.947	4.34	0.02			5	29.641	35.010	4.29	0.02
		10	29.001	34.944	4.33	0.02			10	29.597	35.005	4.31	0.02
		20	28.844	34.938	4.34	0.02			20	28.341	35.094	4.50	0.02
		30	28.432	34.984	4.48	0.03			30	27.913	35.106	4.58	0.02
		40	26.910	35.060	4.62	0.05			40	27.157	35.086	4.64	0.03
		50	25.975	35.063	4.70	0.08			50	26.370	35.030	4.67	0.04
		75	22.887	35.100	3.82	0.26			75	24.392	34.929	4.65	0.12
		100	21.239	35.243	3.49	0.13			100	22.859	35.018	4.04	0.20
		125	19.907	35.344	3.23	0.07			125	21.780	35.111	3.45	0.14
		150	18.185	35.443	3.30	0.03			150	20.098	35.201	2.99	0.08
		175	16.594	35.334	3.09	0.02			175	18.657	35.341	3.03	0.03
		200	15.437	35.295	3.23	0.02			200	16.990	35.311	2.98	0.02
		250	13.674	35.110	2.92	0.02			250	15.741	35.459	4.03	0.02
		300	12.445	34.980	2.72	0.02			300	13.709	35.270	4.18	0.02
		400	10.778	34.872	3.06	0.02			400	11.263	34.983	4.70	0.01
		500	9.568	34.788	3.33	0.02			500	9.402	34.744	4.62	0.01
CTD data (BTL)							CTD data (BTL)						
BTL No.	Depth m	Pres. db	Temp. °C	Sal (psu)	DO ml·l ⁻¹	FLC μg/l	BTL No.	Depth m	Pres. db	Temp. °C	Sal (psu)	DO ml·l ⁻¹	FLC μg/l
Sur.	0	***	29.2	***	***	***	Sur.	0	***	29.7	***	***	***
1	198	199	15.351	35.296	3.26	0.02	1	89	90	23.093	34.992	4.13	0.21
2	199	200	15.387	35.300	3.26	0.02	2	89	90	23.132	34.989	4.16	0.22
3	199	200	15.382	35.302	3.27	0.02	3	89	89	23.180	34.987	4.19	0.22
4	75	75	22.077	35.132	3.65	0.22	4	90	90	23.236	34.985	4.22	0.22
5	74	75	22.507	35.111	3.68	0.23	5	89	90	23.281	34.983	4.23	0.22
6	74	74	22.584	35.103	3.72	0.24	6	89	90	23.288	34.983	4.25	0.22
7	74	74	22.597	35.113	3.72	0.25	7	90	90	23.281	34.984	4.24	0.22
8	73	74	22.578	35.116	3.75	0.25	8	89	90	23.352	34.979	4.27	0.22
9	74	74	22.622	35.114	3.74	0.25	9	89	90	23.390	34.977	4.28	0.22
10	74	75	22.540	35.126	3.74	0.26	10	89	90	23.407	34.976	4.27	0.22
11	101	101	21.070	35.257	3.46	0.11	11	89	90	23.411	34.977	4.28	0.22
12	101	102	21.054	35.262	3.45	0.12	12	89	90	23.417	34.976	4.29	0.23
13	101	101	21.048	35.265	3.43	0.11	13	90	90	23.413	34.977	4.28	0.22
14	100	101	21.040	35.267	3.45	0.12	14	89	90	23.409	34.977	4.28	0.22
15	101	102	21.035	35.265	3.43	0.12	15	100	100	22.681	35.031	3.90	0.19
16	102	102	21.029	35.265	3.44	0.12	16	100	100	22.661	35.034	3.89	0.20
17	49	50	25.153	35.023	4.69	0.15	17	89	90	23.410	34.977	4.27	0.23
18	50	50	25.457	35.033	4.68	0.09	18	90	90	23.381	34.978	4.28	0.23
19	50	50	25.442	35.035	4.68	0.09	19	90	90	23.378	34.978	4.27	0.23
20	49	50	25.429	35.037	4.69	0.09	20	89	90	23.377	34.978	4.28	0.22
21	50	50	25.434	35.037	4.69	0.09	21	49	49	26.040	35.009	4.69	0.05
22	50	50	25.422	35.035	4.68	0.09	22	49	50	26.063	35.012	4.68	0.04
23	5	5	29.042	34.944	4.33	0.02	23	5	5	29.672	35.011	4.29	0.01
24	5	5	29.042	34.945	4.33	0.02	24	5	5	29.674	35.012	4.30	0.01

KH-09-5_Leg5		St.22		Depth		4343 m		KH-09-5_Leg5		St.25		Depth		3939 m	
Date:	2010.2.4		Lat.	16 00.25S		Date:	2010.2.5		Lat.	17 59.86S					
Time:	11:59		Long.	56 59.99E		Time:	5:59		Long.	58 00.02E					
CTD data (LAY)	Pres.	Temp.	Sal	DO	FLC	CTD data (LAY)	Pres.	Temp.	Sal	DO	FLC				
	db	°C	(psu)	ml·l ⁻¹	μg/l		db	°C	(psu)	ml·l ⁻¹	μg/l				
	5	29.600	34.999	4.31	0.02		5	28.857	34.920	4.34	0.01				
	10	29.022	35.026	4.35	0.02		10	28.860	34.922	4.35	0.01				
	20	28.563	35.088	4.44	0.02		20	28.718	34.914	4.36	0.02				
	30	27.503	35.115	4.58	0.03		30	28.098	34.910	4.41	0.02				
	40	26.345	35.055	4.65	0.05		40	27.975	34.927	4.43	0.02				
	50	25.812	35.088	4.59	0.09		50	26.960	34.969	4.63	0.03				
	75	24.174	35.025	4.29	0.27		75	24.465	35.041	4.80	0.05				
	100	22.682	35.108	3.81	0.21		100	23.737	35.139	4.78	0.07				
	125	21.462	35.210	3.45	0.11		125	23.006	35.249	4.73	0.11				
	150	20.581	35.346	3.44	0.05		150	22.391	35.356	4.54	0.15				
	175	19.073	35.432	3.38	0.02		175	20.999	35.529	4.08	0.05				
	200	17.016	35.185	2.47	0.03		200	20.238	35.613	4.04	0.02				
	250	15.933	35.366	3.50	0.02		250	18.477	35.585	3.78	0.01				
	300	13.828	35.272	4.04	0.01		300	16.019	35.487	3.89	0.01				
400	11.833	35.056	4.59	0.01	400	13.405	35.212	4.09	0.01						
500	10.180	34.838	4.62	0.01	500	11.246	34.981	4.74	0.01						
CTD data (BTL)						CTD data (BTL)									
BTL	Depth	Pres.	Temp.	Sal	DO	FLC	BTL	Depth	Pres.	Temp.	Sal	DO	FLC		
No.	m	db	°C	(psu)	ml·l ⁻¹	μg/l	No.	m	db	°C	(psu)	ml·l ⁻¹	μg/l		
Sur.	0	***	29.8	***	***	***	Sur.	0	***	29.2	***	***	***		
1	74	75	24.474	34.981	4.39	0.26	1	198	200	20.020	35.620	4.00	0.02		
2	74	75	24.515	34.977	4.49	0.27	2	199	200	20.048	35.619	4.02	0.02		
3	74	75	24.509	34.983	4.50	0.26	3	199	200	20.095	35.617	4.02	0.02		
4	75	75	24.548	34.982	4.49	0.28	4	138	139	22.654	35.323	4.69	0.16		
5	75	75	24.556	34.974	4.49	0.26	5	138	139	22.659	35.322	4.69	0.16		
6	75	75	24.595	34.976	4.50	0.26	6	138	138	22.658	35.323	4.68	0.16		
7	74	75	24.613	34.985	4.51	0.26	7	138	139	22.660	35.321	4.69	0.16		
8	75	75	24.607	34.977	4.54	0.25	8	138	139	22.658	35.323	4.69	0.16		
9	74	75	24.645	34.987	4.53	0.25	9	138	139	22.660	35.323	4.69	0.16		
10	75	75	24.665	34.975	4.53	0.26	10	138	139	22.664	35.322	4.69	0.16		
11	74	75	24.705	34.976	4.55	0.25	11	99	100	23.661	35.146	4.77	0.07		
12	75	75	24.681	34.977	4.54	0.26	12	99	100	23.828	35.111	4.77	0.07		
13	74	74	24.694	34.984	4.55	0.25	13	99	99	23.789	35.120	4.78	0.06		
14	74	75	24.742	34.986	4.55	0.25	14	99	99	23.836	35.103	4.78	0.07		
15	74	75	24.728	34.984	4.55	0.25	15	99	99	23.848	35.101	4.77	0.06		
16	74	75	24.722	34.981	4.54	0.27	16	99	99	23.850	35.101	4.77	0.06		
17	75	75	24.668	34.981	4.53	0.26	17	50	50	26.122	34.995	4.73	0.03		
18	75	75	24.700	34.977	4.55	0.26	18	49	50	26.231	34.992	4.72	0.03		
19	74	75	24.708	34.977	4.56	0.25	19	49	49	26.234	34.994	4.71	0.03		
20	74	75	24.746	34.980	4.56	0.25	20	49	50	26.351	34.991	4.70	0.03		
21	74	75	24.690	34.975	4.56	0.27	21	49	50	26.346	34.991	4.70	0.03		
22	74	75	24.598	34.978	4.54	0.26	22	49	50	26.343	34.991	4.70	0.03		
23	74	75	24.571	34.974	4.55	0.26	23	5	5	29.030	34.928	4.34	0.01		
24	74	74	24.592	34.974	4.55	0.27	24	5	5	28.965	34.929	4.35	0.01		

KH-09-5_Leg5		St.26		Depth		3524 m		KH-09-5_Leg5		St.27		Depth		4076 m	
Date:	2010.2.5		Lat.	17 00.06S		Date:	2010.2.5		Lat.	16 00.26S					
Time:	12:30		Long.	58 00.15E		Time:	18:37		Long.	58 00.25E					
CTD data (LAY)	Pres.	Temp.	Sal	DO	FLC	CTD data (LAY)	Pres.	Temp.	Sal	DO	FLC				
	db	°C	(psu)	ml·l ⁻¹	μg/l		db	°C	(psu)	ml·l ⁻¹	μg/l				
	5	29.468	34.950	4.31	0.02		5	29.672	34.979	4.29	0.02				
	10	29.406	34.943	4.32	0.02		10	29.669	34.978	4.29	0.02				
	20	28.491	34.961	4.43	0.03		20	28.263	35.048	4.50	0.03				
	30	27.937	35.000	4.48	0.03		30	27.455	35.069	4.58	0.03				
	40	27.208	35.034	4.55	0.05		40	26.217	35.006	4.67	0.04				
	50	26.520	35.036	4.59	0.07		50	25.348	34.963	4.71	0.05				
	75	25.290	35.022	4.58	0.23		75	23.694	34.938	4.68	0.08				
	100	23.742	35.000	4.31	0.26		100	22.866	35.083	4.34	0.23				
	125	22.216	35.143	3.92	0.17		125	21.949	35.231	3.85	0.14				
	150	20.951	35.262	3.22	0.07		150	20.143	35.237	3.01	0.06				
	175	19.257	35.489	3.51	0.02		175	19.016	35.235	2.69	0.04				
	200	18.135	35.518	3.62	0.02		200	17.994	35.252	2.71	0.03				
	250	15.617	35.347	3.45	0.02		250	16.568	35.469	3.74	0.01				
	300	14.412	35.332	3.99	0.01		300	14.469	35.332	3.96	0.02				
	400	11.518	35.018	4.79	0.01		400	11.191	34.971	4.68	0.01				
	500	10.207	34.836	4.84	0.01		500	9.279	34.733	4.51	0.01				
	CTD data (BTL)						CTD data (BTL)								
BTL No.	Depth	Pres.	Temp.	Sal	DO	FLC	BTL No.	Depth	Pres.	Temp.	Sal	DO	FLC		
	m	db	°C	(psu)	ml·l ⁻¹	μg/l		m	db	°C	(psu)	ml·l ⁻¹	μg/l		
Sur.	0	***	29.3	***	***	***	Sur.	0	***	29.6	***	***	***		
1							1	199	200	17.962	35.249	2.71	0.02		
2							2	199	200	17.964	35.250	2.71	0.03		
3							3	199	200	17.960	35.251	2.71	0.03		
4							4	99	100	22.739	35.090	4.22	0.21		
5							5	100	100	22.679	35.100	4.23	0.21		
6							6	100	100	22.710	35.099	4.21	0.21		
7							7	99	100	22.622	35.108	4.23	0.21		
8							8	100	100	22.715	35.096	4.23	0.21		
9							9	99	100	22.698	35.100	4.24	0.21		
10							10	99	100	22.718	35.098	4.24	0.21		
11							11	74	75	23.587	34.950	4.66	0.09		
12							12	75	75	23.595	34.951	4.66	0.09		
13							13	75	75	23.594	34.950	4.66	0.09		
14							14	74	75	23.596	34.950	4.65	0.09		
15							15	74	75	23.594	34.951	4.67	0.09		
16							16	75	75	23.594	34.954	4.67	0.09		
17							17	49	49	24.776	34.916	4.73	0.05		
18							18	49	50	24.846	34.920	4.74	0.05		
19							19	49	49	24.836	34.921	4.73	0.05		
20							20	49	49	24.857	34.922	4.74	0.05		
21							21	49	49	24.857	34.921	4.74	0.05		
22							22	50	50	24.860	34.922	4.74	0.05		
23							23	5	5	29.683	34.965	4.29	0.02		
24							24	5	5	29.682	34.966	4.29	0.02		

KH-09-5_Leg5		St.28	Depth	4162 m		KH-09-5_Leg5		St.29	Depth	4143 m			
Date:	2010.2.6		Lat.	14 59.90S		Date:	2010.2.6		Lat.	13 59.95S			
Time:	1:06		Long.	58 00.05E		Time:	6:55		Long.	58 00.08E			
CTD data (LAY)	Pres.	Temp.	Sal	DO	FLC	CTD data (LAY)	Pres.	Temp.	Sal	DO	FLC		
	db	°C	(psu)	ml·l ⁻¹	μg/l		db	°C	(psu)	ml·l ⁻¹	μg/l		
	5	29.400	35.026	4.29	0.03		5	29.225	34.919	4.32	0.01		
	10	29.399	35.025	4.29	0.02		10	29.179	34.919	4.32	0.02		
	20	28.612	34.955	4.36	0.03		20	28.808	34.934	4.39	0.02		
	30	27.883	35.029	4.47	0.04		30	27.443	35.099	4.56	0.03		
	40	25.881	35.030	4.57	0.15		40	26.327	35.119	4.63	0.03		
	50	25.472	35.044	4.45	0.23		50	24.682	34.919	4.69	0.06		
	75	23.690	34.932	4.63	0.19		75	22.972	35.024	4.57	0.21		
	100	22.595	35.087	4.16	0.17		100	22.737	35.069	4.53	0.15		
	125	21.761	35.173	3.56	0.12		125	21.292	35.206	3.40	0.10		
	150	20.494	35.363	3.42	0.05		150	20.472	35.286	3.22	0.07		
	175	19.377	35.391	3.15	0.03		175	19.328	35.454	3.37	0.04		
	200	18.205	35.357	3.09	0.02		200	17.233	35.174	2.30	0.03		
	250	16.442	35.440	3.68	0.02		250	14.693	35.136	2.69	0.02		
	300	14.490	35.300	3.89	0.02		300	13.388	35.087	2.99	0.02		
	400	11.611	35.014	4.24	0.01		400	11.538	34.987	3.72	0.02		
500	9.536	34.766	4.57	0.01	500	9.789	34.805	3.84	0.02				
CTD data (BTL)						CTD data (BTL)							
BTL	Depth	Pres.	Temp.	Sal	DO	FLC	BTL	Depth	Pres.	Temp.	Sal	DO	FLC
No.	m	db	°C	(psu)	ml·l ⁻¹	μg/l	No.	m	db	°C	(psu)	ml·l ⁻¹	μg/l
Sur.	0	***	29.2	***	***	***	Sur.	0	***	29.4	***	***	***
1							1						
2							2						
3							3						
4							4						
5							5						
6							6						
7							7						
8							8						
9							9						
10							10						
11							11						
12							12						
13							13						
14							14						
15							15						
16							16						
17							17						
18							18						
19							19						
20							20						
21							21						
22							22						
23							23						
24							24						

KH-09-5_Leg5		St.30	Depth	4282 m		KH-09-5_Leg5		St.33	Depth	2748 m			
Date:	2010.2.6		Lat.	12 59.96S		Date:	2010.2.7		Lat.	12 59.75S			
Time:	12:49		Long.	57 59.91E		Time:	8:33		Long.	61 29.90E			
CTD data (LAY)	Pres.	Temp.	Sal	DO	FLC	CTD data (LAY)	Pres.	Temp.	Sal	DO	FLC		
	db	°C	(psu)	ml·l ⁻¹	μg/l		db	°C	(psu)	ml·l ⁻¹	μg/l		
	5	29.353	34.631	4.33	0.02		5	29.378	34.604	4.31	0.02		
	10	29.248	34.630	4.34	0.02		10	29.294	34.614	4.33	0.02		
	20	29.149	34.641	4.35	0.03		20	29.028	34.642	4.36	0.02		
	30	28.416	34.703	4.50	0.04		30	27.846	34.704	4.55	0.03		
	40	26.907	34.796	4.60	0.09		40	25.202	34.796	4.74	0.05		
	50	25.709	34.855	4.55	0.27		50	24.476	34.825	4.74	0.07		
	75	22.271	35.063	3.65	0.29		75	23.016	34.962	4.41	0.31		
	100	19.543	35.064	2.51	0.14		100	21.702	35.276	3.93	0.14		
	125	18.739	35.424	3.34	0.05		125	20.439	35.295	3.27	0.08		
	150	17.465	35.430	3.27	0.03		150	18.181	35.160	2.57	0.04		
	175	15.366	35.173	2.62	0.03		175	17.159	35.150	2.36	0.03		
	200	14.621	35.163	2.73	0.02		200	16.905	35.357	2.99	0.02		
	250	13.341	35.120	3.08	0.03		250	15.181	35.444	4.07	0.02		
	300	12.202	35.045	3.41	0.02		300	13.530	35.295	4.58	0.01		
	400	10.511	34.882	3.52	0.02		400	11.072	34.955	4.66	0.01		
500	9.393	34.757	4.00	0.01	500	9.283	34.722	4.45	0.01				
CTD data (BTL)						CTD data (BTL)							
BTL	Depth	Pres.	Temp.	Sal	DO	FLC	BTL	Depth	Pres.	Temp.	Sal	DO	FLC
No.	m	db	°C	(psu)	ml·l ⁻¹	μg/l	No.	m	db	°C	(psu)	ml·l ⁻¹	μg/l
Sur.	0	***	29.4	***	***	***	Sur.	0	***	29.6	***	***	***
1	199	200	14.694	35.172	2.76	0.02	1	199	200	16.878	35.388	3.18	0.02
2	199	200	14.687	35.172	2.76	0.02	2	198	200	16.920	35.377	3.06	0.02
3	199	201	14.682	35.172	2.76	0.02	3	198	200	16.918	35.374	3.07	0.02
4	60	60	24.923	34.884	4.31	0.28	4	75	75	23.644	34.871	4.63	0.19
5	60	60	24.885	34.892	4.31	0.29	5	75	75	23.841	34.853	4.68	0.13
6	60	60	24.920	34.890	4.32	0.37	6	75	75	23.795	34.854	4.68	0.13
7	60	60	24.920	34.891	4.33	0.28	7	75	76	23.829	34.854	4.69	0.14
8	60	60	24.907	34.892	4.32	0.28	8	75	76	23.839	34.853	4.67	0.14
9	60	60	24.916	34.892	4.33	0.29	9	75	76	23.846	34.853	4.68	0.15
10	60	60	24.912	34.893	4.33	0.28	10	75	75	23.845	34.853	4.67	0.14
11	100	100	19.759	35.106	2.66	0.14	11	100	101	21.871	35.226	4.05	0.17
12	100	100	19.809	35.119	2.67	0.14	12	100	101	21.981	35.189	4.14	0.20
13	100	100	19.813	35.121	2.68	0.14	13	101	101	22.007	35.181	4.11	0.20
14	100	100	19.804	35.119	2.68	0.14	14	100	101	22.054	35.163	4.13	0.21
15	99	100	19.818	35.126	2.69	0.14	15	99	100	22.081	35.156	4.18	0.21
16	99	100	19.820	35.125	2.72	0.13	16	99	99	22.161	35.137	4.19	0.22
17	49	50	26.044	34.840	4.60	0.20	17	50	50	25.219	34.801	4.73	0.06
18	50	50	26.032	34.840	4.60	0.20	18	51	51	25.086	34.806	4.74	0.06
19	49	50	26.007	34.843	4.60	0.19	19	50	51	25.129	34.806	4.74	0.05
20	49	50	26.056	34.841	4.61	0.33	20	50	50	25.205	34.804	4.74	0.06
21	50	50	26.074	34.841	4.61	0.19	21	50	50	25.282	34.803	4.74	0.06
22	50	50	26.072	34.841	4.61	0.19	22	51	51	24.862	34.814	4.74	0.06
23	5	5	29.770	34.639	4.32	0.02	23	6	6	29.658	34.602	4.32	0.01
24	5	5	29.764	34.639	4.32	0.03	24	6	6	29.663	34.603	4.32	0.02

KH-09-5_Leg5		St.40	Depth	2460 m		
Date:	2010.2.9		Lat.	18 00.01S		
Time:	8:13		Long.	58 59.76E		
CTD data (LAY)	Pres.	Temp.	Sal	DO	FLC	
	db	°C	(psu)	ml·l ⁻¹	μg/l	
	5	29.368	34.736	4.33	0.02	
	10	29.197	34.697	4.33	0.02	
	20	28.081	34.762	4.42	0.02	
	30	27.867	34.877	4.46	0.02	
	40	27.239	34.994	4.56	0.02	
	50	26.190	35.007	4.66	0.03	
	75	24.678	34.979	4.72	0.06	
	100	23.337	35.016	4.68	0.13	
	125	22.225	35.319	4.31	0.12	
	150	21.106	35.359	3.59	0.06	
	175	20.490	35.372	3.42	0.05	
	200	18.588	35.349	3.05	0.03	
	250	17.556	35.487	3.52	0.02	
	300	16.035	35.528	4.02	0.01	
	400	12.740	35.184	4.52	0.01	
500	10.635	34.894	4.91	0.01		
CTD data (BTL)						
BTL No.	Depth m	Pres. db	Temp. °C	Sal (psu)	DO ml·l ⁻¹	FLC μg/l
Sur.	0	***	29.5	***	***	***
1	198	199	18.417	35.347	3.06	0.02
2	199	200	18.584	35.348	3.06	0.03
3	199	200	18.544	35.347	3.06	0.03
4	115	116	22.331	35.294	4.35	0.13
5	116	117	22.335	35.296	4.35	0.13
6	116	116	22.346	35.294	4.36	0.13
7	116	116	22.354	35.292	4.36	0.13
8	116	117	22.338	35.297	4.36	0.13
9	117	117	22.328	35.301	4.36	0.13
10	115	115	22.374	35.288	4.37	0.13
11	100	100	23.216	35.051	4.67	0.15
12	100	100	23.233	35.043	4.66	0.15
13	99	100	23.264	35.029	4.66	0.15
14	99	100	23.279	35.022	4.66	0.15
15	100	100	23.272	35.030	4.66	0.15
16	99	100	23.281	35.021	4.67	0.15
17	51	51	26.080	35.001	4.66	0.03
18	51	52	26.172	35.005	4.66	0.03
19	50	51	26.189	35.005	4.65	0.03
20	51	51	26.203	35.005	4.65	0.03
21	51	52	26.179	35.008	4.65	0.03
22	51	51	26.213	35.006	4.65	0.03
23	6	6	29.398	34.721	4.33	0.01
24	5	6	29.307	34.716	4.33	0.01

KH-09-5_Leg6		St.1		Depth	4853 m		KH-09-5_Leg6		St.2		Depth	4807 m	
Date:	2010.2.16		Lat.	17 10.17S		Date:	2010.2.17		Lat.	16 22.30S			
Time:	19:26		Long.	71 55.71E		Time:	14:27		Long.	76 09.48E			
CTD data (LAY)	Pres.	Temp.	Sal	DO	FLC	CTD data (LAY)	Pres.	Temp.	Sal	DO	FLC		
	db	°C	(psu)	ml·l ⁻¹	μg/l		db	°C	(psu)	ml·l ⁻¹	μg/l		
	10	28.820	34.392	4.33	0.01		5	28.875	34.202	4.32	0.02		
	20	28.822	34.392	4.34	0.02		10	28.882	34.202	4.32	0.03		
	30	28.822	34.394	4.33	0.02		20	28.885	34.202	4.32	0.02		
	40	28.721	34.402	4.35	0.02		30	28.837	34.206	4.33	0.03		
	50	27.412	34.602	4.56	0.03		40	28.698	34.228	4.37	0.04		
	75	25.339	34.729	4.66	0.06		50	28.041	34.339	4.43	0.06		
	100	22.676	35.021	4.11	0.34		75	26.975	34.838	4.18	0.18		
	125	21.315	35.113	3.30	0.15		100	23.112	34.912	3.75	0.25		
	150	20.889	35.267	3.61	0.07		125	21.138	34.961	2.72	0.12		
	175	20.112	35.494	3.63	0.03		150	19.397	35.040	2.67	0.05		
	200	19.723	35.623	3.84	0.02		175	19.767	35.447	3.45	0.03		
	250	18.167	35.691	3.99	0.01		200	18.632	35.533	3.58	0.02		
	300	16.382	35.623	4.23	0.01		250	17.491	35.681	4.03	0.01		
	400	12.602	35.182	4.79	0.01		300	15.274	35.459	4.00	0.01		
	500	10.555	34.883	5.03	0.01		400	11.626	35.023	4.34	0.01		
					500	9.628	34.749	4.99	0.01				
CTD data (BTL)						CTD data (BTL)							
BTL	Depth	Pres.	Temp.	Sal	DO	FLC	BTL	Depth	Pres.	Temp.	Sal	DO	FLC
No.	m	db	°C	(psu)	ml·l ⁻¹	μg/l	No.	m	db	°C	(psu)	ml·l ⁻¹	μg/l
Sur.	0	***	28.8	***	***	***	Sur.	0	***	28.7	***	***	***
1	198	200	19.695	35.626	3.86	0.02	1						
2	199	200	19.694	35.627	3.86	0.02	2						
3	199	200	19.7	35.628	3.85	0.02	3						
4	199	200	19.693	35.628	3.85	0.02	4						
5	199	200	19.694	35.628	3.86	0.02	5						
6	99	100	22.360	35.072	4.03	0.25	6						
7	99	99	22.326	35.071	4.04	0.28	7						
8	100	100	22.423	35.060	4.05	0.29	8						
9	100	100	22.496	35.050	4.09	0.29	9						
10	99	100	22.541	35.039	4.16	0.30	10						
11	74	74	24.333	34.856	4.63	0.12	11						
12	75	75	24.635	34.832	4.64	0.10	12						
13	75	76	24.736	34.818	4.65	0.10	13						
14	75	75	24.855	34.797	4.66	0.09	14						
15	74	74	24.983	34.779	4.66	0.08	15						
16	50	50	26.823	34.579	4.62	0.04	16						
17	50	50	27.126	34.584	4.60	0.04	17						
18	49	50	27.149	34.592	4.59	0.03	18						
19	49	50	27.326	34.601	4.57	0.03	19						
20	49	49	27.450	34.607	4.56	0.03	20						
21	4	4	28.809	34.391	4.34	0.02	21						
22	4	4	28.809	34.391	4.33	0.02	22						
23	4	4	28.807	34.391	4.33	0.02	23						
24	4	4	28.808	34.391	4.34	0.02	24						

KH-09-5_Leg6		St.4	Depth	5111 m		KH-09-5_Leg6		St.6	Depth	2553 m			
Date:	2010.2.18		Lat.	15 28.90S		Date:	2010.2.19		Lat.	14 30.36S			
Time:	13:22		Long.	80 55.26E		Time:	13:22		Long.	86 05.99E			
CTD data (LAY)	Pres.	Temp.	Sal	DO	FLC	CTD data (LAY)	Pres.	Temp.	Sal	DO	FLC		
	db	°C	(psu)	ml·l ⁻¹	μg/l		db	°C	(psu)	ml·l ⁻¹	μg/l		
	10	28.544	34.217	4.35	0.02		10	28.447	34.333	4.36	0.03		
	20	28.533	34.210	4.36	0.02		20	28.437	34.332	4.36	0.03		
	30	28.437	34.231	4.38	0.03		30	28.373	34.339	4.37	0.03		
	40	28.405	34.354	4.37	0.03		40	28.333	34.362	4.38	0.04		
	50	28.305	34.407	4.37	0.03		50	28.200	34.404	4.41	0.05		
	75	27.711	34.716	4.48	0.05		75	26.645	34.669	4.41	0.12		
	100	25.895	34.774	4.53	0.14		100	25.267	34.676	4.30	0.29		
	125	22.086	34.839	2.94	0.17		125	22.209	34.779	3.15	0.13		
	150	19.862	34.972	2.50	0.07		150	19.879	34.751	2.48	0.06		
	175	19.336	35.324	3.15	0.04		175	19.236	35.273	3.11	0.02		
	200	17.578	35.180	2.78	0.03		200	17.399	35.085	2.74	0.02		
	250	15.039	35.079	2.78	0.02		250	15.932	35.241	3.10	0.02		
	300	13.510	35.113	3.18	0.02		300	14.020	35.195	3.38	0.02		
	400	11.248	34.969	4.25	0.01		400	11.208	34.955	4.17	0.01		
	500	9.471	34.738	4.46	0.01		500	9.178	34.713	3.09	0.02		
CTD data (BTL)						CTD data (BTL)							
BTL	Depth	Pres.	Temp.	Sal	DO	FLC	BTL	Depth	Pres.	Temp.	Sal	DO	FLC
No.	m	db	°C	(psu)	ml·l ⁻¹	μg/l	No.	m	db	°C	(psu)	ml·l ⁻¹	μg/l
Sur.	0	***	28.6	***	***	***	Sur.	0	***	28.5	***	***	***
1	198	200	17.520	35.190	2.79	0.03	1						
2	199	200	17.617	35.177	2.78	0.03	2						
3	199	200	17.6	35.183	2.76	0.02	3						
4	199	200	17.718	35.185	2.75	0.03	4						
5	199	200	17.820	35.191	2.75	0.03	5						
6	112	113	23.002	34.854	3.94	0.23	6						
7	112	112	24.009	34.858	4.06	0.23	7						
8	112	113	24.031	34.862	4.06	0.23	8						
9	112	112	24.024	34.857	4.07	0.23	9						
10	112	113	24.026	34.872	4.05	0.23	10						
11	99	99	25.537	34.776	4.56	0.15	11						
12	99	99	25.693	34.785	4.55	0.15	12						
13	99	100	25.736	34.788	4.55	0.15	13						
14	99	100	25.756	34.790	4.55	0.14	14						
15	99	100	25.789	34.791	4.54	0.15	15						
16	50	50	28.281	34.403	4.36	0.03	16						
17	50	50	28.280	34.403	4.37	0.03	17						
18	50	50	28.279	34.403	4.37	0.04	18						
19	50	50	28.276	34.402	4.37	0.04	19						
20	50	50	28.277	34.403	4.37	0.03	20						
21	5	5	28.512	34.211	4.37	0.02	21						
22	4	4	28.514	34.212	4.37	0.02	22						
23	4	4	28.513	34.211	4.35	0.02	23						
24	4	4	28.514	34.212	4.36	0.03	24						

KH-09-5_Leg6		St.8		Depth		5217 m		KH-09-5_Leg6		St.10		Depth		3286 m	
Date:		2010.2.20		Lat.		13 33.03S		Date:		2010.2.22		Lat.		12 08.36S	
Time:		12:24		Long.		91 11.24E		Time:		11:35		Long.		102 31.36E	
CTD data (LAY)	Pres.	Temp.	Sal	DO	FLC	CTD data (LAY)	Pres.	Temp.	Sal	DO	FLC				
	db	°C	(psu)	ml·l ⁻¹	μ g/l		db	°C	(psu)	ml·l ⁻¹	μ g/l				
	10	28.981	34.064	4.33	0.03		5	29.831	34.413	4.31	0.02				
	20	28.866	34.169	4.36	0.03		10	29.836	34.415	4.31	0.02				
	30	27.956	34.365	4.48	0.04		20	29.127	34.573	4.37	0.03				
	40	27.545	34.528	4.50	0.05		30	28.789	34.596	4.39	0.03				
	50	27.112	34.513	4.50	0.06		40	27.513	34.482	4.59	0.03				
	75	25.788	34.677	4.50	0.14		50	26.880	34.506	4.64	0.05				
	100	23.503	34.881	3.93	0.22		75	25.281	34.499	4.54	0.11				
	125	21.952	35.015	3.65	0.10		100	23.212	34.597	3.58	0.27				
	150	20.131	34.996	3.09	0.05		125	20.623	34.691	2.80	0.09				
	175	17.957	34.947	2.71	0.02		150	18.861	34.755	2.58	0.05				
	200	16.672	34.937	2.65	0.02		175	15.469	34.626	2.39	0.02				
	250	14.804	35.069	2.98	0.02		200	14.114	34.653	2.31	0.02				
	300	13.491	35.276	4.32	0.02		250	12.337	34.649	2.26	0.03				
	400	10.005	34.802	5.11	0.01		300	11.488	34.724	2.35	0.03				
	500	8.437	34.614	4.52	0.01		400	9.757	34.720	2.83	0.02				
CTD data (BTL)						CTD data (BTL)									
BTL	Depth	Pres.	Temp.	Sal	DO	FLC	BTL	Depth	Pres.	Temp.	Sal	DO	FLC		
No.	m	db	°C	(psu)	ml·l ⁻¹	μ g/l	No.	m	db	°C	(psu)	ml·l ⁻¹	μ g/l		
Sur.	0	***	28.4	***	***	***	Sur.	0	***	29.8	***	***	***		
1	200	201	16.754	34.964	2.72	0.02	1	200	201	14.079	34.653	2.29	0.03		
2	200	201	16.772	34.968	2.69	0.02	2	199	200	14.088	34.654	2.30	0.02		
3	200	201	16.7	34.960	2.68	0.02	3	200	201	14.080	34.654	2.30	0.02		
4	200	201	16.711	34.956	2.68	0.02	4	199	201	14.094	34.653	2.30	0.03		
5	201	202	16.638	34.952	2.67	0.02	5	200	201	14.098	34.654	2.30	0.02		
6	90	90	24.960	34.767	4.42	0.26	6	91	91	23.600	34.553	3.82	0.31		
7	89	90	24.941	34.763	4.40	0.27	7	91	91	23.636	34.562	3.78	0.31		
8	90	90	24.953	34.763	4.47	0.26	8	90	90	23.687	34.564	3.84	0.31		
9	90	90	24.957	34.765	4.41	0.28	9	89	89	23.772	34.570	3.99	0.32		
10	90	91	24.958	34.767	4.45	0.26	10	89	90	23.787	34.571	4.03	0.32		
11	99	100	23.856	34.847	4.14	0.24	11	99	100	22.404	34.629	3.30	0.19		
12	100	100	23.876	34.848	4.15	0.24	12	99	100	22.430	34.623	3.30	0.20		
13	101	101	23.869	34.851	4.15	0.25	13	100	100	22.434	34.623	3.34	0.18		
14	100	101	23.939	34.842	4.16	0.25	14	100	101	22.382	34.654	3.29	0.19		
15	100	101	23.900	34.846	4.15	0.25	15	99	99	22.444	34.622	3.31	0.19		
16	49	49	27.432	34.537	4.52	0.06	16	50	50	27.233	34.562	4.62	0.05		
17	50	50	27.442	34.538	4.53	0.06	17	49	50	27.253	34.553	4.61	0.05		
18	49	49	27.440	34.537	4.52	0.06	18	50	50	27.237	34.565	4.62	0.05		
19	49	50	27.451	34.538	4.53	0.06	19	50	50	27.243	34.562	4.62	0.05		
20	49	49	27.458	34.536	4.52	0.06	20	49	50	27.301	34.541	4.62	0.05		
21	5	5	29.000	34.042	4.33	0.03	21	5	5	29.777	34.416	4.30	0.02		
22	5	5	28.998	34.042	4.34	0.03	22	5	5	29.786	34.416	4.31	0.02		
23	5	5	28.990	34.042	4.33	0.03	23	5	5	29.787	34.416	4.31	0.02		
24	5	6	28.997	34.045	4.33	0.03	24	5	5	29.782	34.416	4.30	0.02		

KH-09-5_Leg6	St.11	Depth	5194 m			
Date:	2010.2.23	Lat.	11	20.59S		
Time:	15:22	Long.	109	08.13E		
CTD data (LAY)	Pres.	Temp.	Sal	DO	FLC	
	db	°C	(psu)	ml·l ⁻¹	μg/l	
	5	30.291	33.603	4.31	0.03	
	10	30.238	33.606	4.30	0.03	
	20	30.204	33.690	4.32	0.03	
	30	29.819	34.358	4.39	0.03	
	40	29.226	34.448	4.41	0.04	
	50	29.035	34.602	4.42	0.06	
	75	25.058	34.457	3.88	0.30	
	100	22.294	34.622	3.09	0.12	
	125	18.525	34.567	2.74	0.04	
	150	17.561	34.584	2.67	0.03	
	175	15.963	34.541	2.63	0.02	
	200	15.092	34.711	2.45	0.02	
	250	12.333	34.677	2.34	0.03	
	300	11.342	34.757	2.48	0.02	
	400	9.619	34.725	2.06	0.03	
	500	8.163	34.649	2.34	0.03	
CTD data (BTL)						
BTL	Depth	Pres.	Temp.	Sal	DO	FLC
No.	m	db	°C	(psu)	ml·l ⁻¹	μg/l
Sur.	0	***	30.2	***	***	***
1	200	201	14.735	34.637	2.42	0.02
2	199	200	14.669	34.637	2.42	0.02
3	200	201	14.7	34.631	2.42	0.02
4	200	201	14.629	34.627	2.43	0.02
5	201	202	14.468	34.611	2.43	0.02
6	66	66	25.507	34.353	4.08	0.36
7	66	66	25.515	34.362	4.04	0.39
8	66	67	25.463	34.291	3.98	0.41
9	66	66	25.536	34.276	3.95	0.41
10	66	67	25.529	34.262	3.94	0.42
11	99	99	22.222	34.612	3.05	0.11
12	101	101	22.236	34.617	3.06	0.11
13	98	99	22.317	34.615	3.06	0.11
14	99	100	22.316	34.616	3.07	0.11
15	100	100	22.307	34.615	3.05	0.11
16	51	51	29.002	34.594	4.40	0.07
17	50	50	29.003	34.593	4.41	0.07
18	51	51	29.027	34.600	4.41	0.06
19	50	50	29.027	34.595	4.41	0.06
20	51	52	29.023	34.602	4.40	0.06
21	6	6	30.244	33.612	4.30	0.03
22	6	6	30.246	33.613	4.30	0.03
23	6	6	30.246	33.613	4.30	0.03
24	6	6	30.245	33.612	4.31	0.03

